Identification of the Outer Membrane Porin of *Thermus thermophilus* HB8: the Channel-Forming Complex Has an Unusually High Molecular Mass and an Extremely Large Single-Channel Conductance

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The outer membrane of the thermophilic bacterium *Thermus thermophilus* was isolated using sucrose step gradient centrifugation. Its detergent extracts contained an ion-permeable channel with an extremely high single-channel conductance of 20 nS in 1 M KCl. The channel protein was purified by preparative sodium dodecyl sulfate (SDS)-polyacrylamide gel electrophoresis. It has a high molecular mass of 185 kDa, and its channel-forming ability resists boiling in SDS for 10 min.

Porins are membrane proteins that form channels for small hydrophilic solutes in the outer membrane of gram-negative bacteria (1, 26, 27) and in the cell wall of the mycolata (20, 34, 35). Porins mediate the passive diffusion of ions and small hydrophilic nutrient molecules across the outer membrane or the cell wall according to their molecular masses (exclusion limit of about 600 Da in *Escherichia coli*) or according to kind of the substrate when the porins are substrate specific (specific porins), such as LamB for carbohydrates (5, 22) or Tsx for nucleosides (4). Specific porins have binding sites for the diffusion of specific classes of nutrients across the cell wall (24). Porins of the outer membrane of gram-negative bacteria consist of three identical subunits, which are stable against denaturing conditions due to their predominating β-barrel structure and embedding in the membrane (25). Some of the outer membrane porins from different organisms have been crystallized (12, 31, 37). According to their three-dimensional structure, each of the three monomers contains a channel with a diameter of between 0.8 and 1.4 nm. These porins have a single-channel conductance of between 150 pS and 3.5 nS in 1 M KCl and are only moderately ion selective (1).

Enteric gram-negative bacteria such as *E. coli* and *Salmonella enterica* serovar Typhimurium have been well characterized for the presence of porins with molecular masses of between 30 and 60 kDa in the outer membrane (1). comparatively little is known about the porins of other gram-negative bacteria, such as the thermophilic *Thermus thermophilus*. Here, besides interesting studies of S-layer proteins (10, 13, 14, 28) and the composition of the peptidoglycan (29), little is known about the structure of the outer membrane and about the existence of porins. In this study, we present the identification and purification of the first outer membrane porin of the thermophilic gram-negative bacterium *T. thermophilus*.

*T. thermophilus* strain HB8 was obtained from the Deutsche Stammssammlung. It was grown in batch cultures at 70°C using a New Brunswick shaker at 120 rpm for about 1 to 2 days until the cells reached the stationary phase. The growth medium was composed of either standard medium (8) or Luria-Bertani medium. The cells were harvested by centrifugation (10,000 rpm for 10 min in a Beckman J2-21M/E centrifuge [rotor JA20]) and washed twice in 50 mM Tris-HCl (pH 8.0). About 5 g of cells (wet weight) was suspended in 50 ml of 50 mM Tris-HCl (pH 8.0) and kept on ice. The cells were disrupted with a Branson sonifier (8). Unbroken cells were removed by centrifugation at 12,500 × g for 10 min at 4°C. The cell envelopes (cytoplasmic membrane, murein, and outer membrane) were obtained by centrifugation of the supernatant at 170,000 × g for 60 min (Beckman Omega 90 XL ultracentrifuge, rotor 70.1 Ti) at 4°C. The pellet was resuspended in 3 ml of 50 mM Tris-HCl (pH 8.0) and centrifuged again under the same conditions for 30 min. The final pellet was resuspended in 0.5 ml of 50 mM Tris-HCl (pH 8) and applied to a step gradient of 30% (3 ml), 40% (4 ml), 50% (2 ml), 55% (2 ml), and 65% (1 ml) sucrose. The gradient was centrifuged at 110,000 × g for 16 h in a Beckman Optima 90 XL ultracentrifuge (rotor SW40Ti) at 4°C.

Eight fractions of the gradient (F1 to F8, from top to bottom) were collected, each with a volume of about 1.5 ml, and analyzed for protein content by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and for the presence of pore-forming activity by reconstitution experiments in lipid bilayer membranes. Fractions F3, F4, F5, and F6 were either yellow or orange and contained the inner membrane. Fraction F7 contained two white bands. The highest pore-forming activity in the lipid bilayer assay was measured for fraction F7. This fraction contained only a few protein bands. One predominant one had a very high molecular mass of about 180 kDa. Some minor channel-forming activity was found in...
The gel was stained with Coomassie brilliant blue.

of protein of the supernatant was solubilized at 30°C for 10 min in 15 mM CaCl2. This fraction also contained a high-molecular-

mass band as a prominent protein (Fig. 1). In further experi-

ments, we subjected the 0.6% LDAO fraction to preparative SDS-PAGE (7% gels, similar to Fig. 1) and eluted six different regions of the gel with 1% Genapol–10 mM Tris (pH 8) overnight at 4°C. The eluted proteins of the SDS-PAGE gel showed no channel-forming activity for molecular mass regions below 120 kDa. However, high activity was found for protein eluted from the 185-kDa band. The 185-kDa fractions were collected and electrophoresed again. The 185-kDa protein was found to be essentially free of contaminant protein, as Fig. 2 clearly demonstrates. It is the outer membrane porin of *T. thermophilus*. We investigated its biochemical properties and found it to be extremely stable. Even heating to 100°C for more than 10 min in 1% SDS and treatment with organic solvents did not affect its channel-forming activity. Furthermore, its molecular mass did not change as the result of such treatments. The results presented here indicate that the outer membrane porin of *T. thermophilus* is indeed extremely stable.

The channel formation mediated by the 185-kDa porin was further analyzed in single-channel experiments. Small amounts of the porin protein (about 5 ng/ml) were added to the aqueous phase on one or both sides of the membrane. After a delay of 1 to 2 min, probably caused by slow aqueous diffusion of the protein, the current increased in a stepwise fashion similar to that observed previously for gram-negative bacterial porins (2).

Figure 3 demonstrates that the channels had an extremely high single-channel conductance compared to that of the enteric bacteria (1, 2). The channels had a long lifetime, because only on-steps were observed within at least several minutes. Interestingly, the single-channel distribution was fairly homogeneous, and more than 80% of the channels were localized within the conductance range between 17.5 and 22.5 nS.

The giant pore from the *T. thermophilus* outer membrane was only moderately cation selective. This can be derived from single-channel experiments in which KCl was replaced by LiCl or potassium acetate (KAc), i.e., the mobile ions K+ and Cl− were replaced by the less-mobile ions Li+ and acetate − (Table 1). The single-channel conductance in 1 M LiCl and 1 M KAc decreased on average by a factor of about 2 compared with the conductance in 1 M KCl. This result indicated that the channel conductance followed the bulk aqueous conductivity of these salt solutions because they are about half that in 1 M KCl. Furthermore, we observed a linear dependence of the single-channel conductance as a function of the bulk aqueous conductivity (Table 1). This is of course expected when we assume that the outer membrane porin of *T. thermophilus* is wide and water-filled, as is suggested by its huge single-channel conductance in 1 M KCl.

The results of the single-channel experiments agree with zero-current membrane potential measurements in the presence of salt gradients (3). After incorporation of a large number of channels in membranes bathed in 50 mM KCl, 10-fold salt gradients were established across the membranes by addition of small amounts of concentrated KCl solution to one side of the membrane. For KCl and KAc, the more diluted side of the membrane became slightly positive, which indicated pref-

![Image](http://jb.asm.org/)

FIG. 1. SDS–7% PAGE of the supernatant of cell envelopes treated with 10 mM Tris-HCl, 10 mM CaCl2 (pH 8), and 0.6% LDAO. Lane 1, molecular mass markers (66, 45, and 36 kDa). Lane 2, 50 μg of protein of the supernatant was solubilized at 30°C for 10 min in 15 μl of sample buffer. The gel was stained with Coomassie brilliant blue.

![Image](http://jb.asm.org/)

FIG. 2. SDS–5% PAGE of the 185-kDa protein of the outer membrane of *T. thermophilus* obtained by elution from preparative SDS-polyacrylamide gels. Lane 1, high-molecular-mass markers (205, 116, 97, 84, and 66 kDa). Lane 2, 5 μg of the pure protein solubilized at 100°C for 10 min in 7 μl of sample buffer. The gel was stained with Coomassie brilliant blue.
The gated channels formed by the monomeric outer membrane receptors such as FhuA (11) and BtuB12 (33) have higher molecular masses (around 60 to 80 kDa) than the porins. The gated channels formed by the outer membrane porins of gram-negative bacteria investi-
gates a general diffusion pore.

FIG. 3. Single-channel recording of the 185-kDa porin from the outer membrane of T. thermophilus. Channel-forming activity was measured with lipid bilayer membranes made of a 1% (wt/vol) solution of diphytanoyl phosphatidylcholine (Avanti Polar Lipids, Alabaster, Ala.) in n-decane (2). The aqueous phase contained 1 M KCl and 5 ng of protein per ml. The applied membrane potential was 20 mV; T = 20°C. The single-channel recordings were performed using Ag/AgCl electrodes (with salt bridges) connected in series to a voltage source and a Keithley 427 current amplifier. The amplified signal was monitored on a digital oscilloscope and recorded on a strip chart or tape recorder.

The protein was purified on a digital oscilloscope and recorded on a strip chart or tape recorder. When we used LiCl, the more dilute side became slightly negative. The zero-current membrane potential for the salts mentioned above had values of −3.4 mV (LiCl), 12 mV (KCl), and 26 mV (KAc). Analysis of these potentials using the Goldman-Hodgkin-Katz equation (3) suggested that the outer membrane channel has only a small selectivity, because the ratios of the permeability Pcation and Pion were 0.85 (LiCl), 1.8 (KCl), and 3.7 (KAc), which indicates that the outer membrane pore of T. thermophilus is indeed a general diffusion pore.

The outer membrane porin of T. thermophilus was purified to homogeneity using preparative SDS-PAGE, and its properties were studied in lipid bilayer membranes. The protein has an extremely large molecular mass (185 kDa) compared with the outer membrane porins of gram-negative bacteria investigated to date, which normally range between 30 kDa for the general diffusion porin of Rhodobacter capsulatus (6) and 3.5 nS for the general diffusion porin of Rhodobacter capsulatus (6) under otherwise identical conditions.

Earlier investigations into the structure of the outer membrane of T. thermophilus have already suggested that it has a normal membrane-like structure because it undergoes thermotropic phase transitions dependent on the growth temperature (36). In other investigations, it has been demonstrated that calcium plays a major role in the stability of proteins from the cell envelope of T. thermophilus (7). In our study, we showed that the proteins of the outer membrane have quite different properties from those of enteric bacteria, because we could not find any peptidoglycan-associated proteins, and the wash of the cell envelope with SDS and isolation of the peptidoglycan-associated protein (23) failed. The S-layer protein of T. thermophilus HB8, which has a molecular mass of about 100 kDa, has been studied in some detail in recent years (13, 14, 16, 28) and is probably already lost upon washing either with the Tris-HCl buffer or with the first wash with 0.01% LDAO, because we did not find the 100-kDa band in the outer membrane fraction of the sucrose density centrifugation or in the supernatants of the different LDAO steps. Although it has been claimed that the S-layer protein may form porin-like structures (10), it seems clear that it is not responsible for the permeability properties of the outer membrane, because the 100-kDa protein is not an integral outer membrane protein, although it contains a domain that interacts with the peptidoglycan layer (28).

![Diagram of single-channel recording](image)

<table>
<thead>
<tr>
<th>Salt</th>
<th>Concentration (M)</th>
<th>Vm (mV)</th>
<th>G (nS)</th>
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<tbody>
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<tr>
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*The membranes were formed from 1% diphytanoyl phosphatidylcholine dissolved in n-decane; T = 20°C. The pH of the aqueous salt solutions was around 6 unless otherwise indicated. G is given as the mean of at least 100 single steps similar to those shown in Fig. 3.
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REFERENCES