Membrane Association and Multimerization of TcpT, the Cognate ATPase Ortholog of the Vibrio cholerae Toxin-Coregulated-Pilus Biogenesis Apparatus

Shital A. Tripathi and Ronald K. Taylor*

Department of Microbiology and Immunology, Dartmouth Medical School, Hanover, New Hampshire 03755

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The toxin-coregulated pilus (TCP) is one of the major virulence factors of Vibrio cholerae. Biogenesis of this type 4 pilus (Tfp) requires a number of structural components encoded by the tcp operon. TcpT, the cognate ATPase, is required for TCP biogenesis and all TCP-mediated functions. We studied the stability and localization of TcpT in cells containing in-frame deletions in each of the tcp genes. TcpT was detectable in each of the biogenesis mutants except the ΔtcpT strain. TcpT was localized to the inner membrane (IM) in a TcpR-dependent manner. TcpR is a predicted bitopic inner membrane protein of the TCP biogenesis apparatus. Using metal affinity pull-down experiments, we demonstrated interaction between TcpT and TcpR. Using Escherichia coli as a heterologous system, we investigated direct interaction between TcpR and TcpT. We report that TcpR is sufficient for TcpT IM localization per se; however, stable IM localization of TcpT requires an additional V. cholerae-specific factor(s). A LexA-based two-hybrid system was utilized to define interaction domains of the two proteins. We demonstrate a strong interaction between the cytoplasmic domain of TcpR and the N-terminal 100 amino acid residues of TcpT. We also demonstrated the ability of the C-terminal domain of TcpT to multimerize.

Cholera, an acute gastrointestinal diarrheal disease, continues to threaten the world as a global public health problem, resulting in hundreds of thousands of cases per year (14). Vibrio cholerae, the causative agent of cholera, is a motile, gram-negative, comma-shaped bacterium that can be acquired by humans via consumption of contaminated food or water (22). Upon ingestion and survival of the gastric acid barrier, V. cholerae colonizes the upper intestine, manifesting pathogenesis by expressing various virulence factors. The toxin-coregulated pilus (TCP) is the major intestinal colonization factor of V. cholerae. TCP-deficient V. cholerae mutants are essentially completely defective in colonization of both humans and infant mice, the latter of which are used as a cholera model (17, 45, 47). TCP not only facilitates colonization directly, via mediating microcolony formation, but also mediates secretion of a soluble colonization factor, TcpF (24, 25). Upon colonization, V. cholerae secretes cholera toxin (CT), which deregulates ion transport by intestinal epithelial cells, leading to a massive loss of water and electrolytes, resulting in a characteristic “rice water stool” (22). TCP serves as the receptor for CTXφ, which carries the CT genes (49). TCP provides a model for study of the biogenesis of type 4 pili (Tfp), enabling us to understand how microbes assemble a sophisticated macromolecular structure that participates in such various functions as colonization, extracellular secretion, and bacteriophage uptake. Based on the N-terminal sequence of the pilin and properties of the assembled filament, TCP shares the structural and functional characteristics of Tfp (44) and is classified into the subclass of type 4b pili, which are often associated with intestinal infections, for example, the CFA/III pilus of enterotoxigenic Escherichia coli (ETEC), the CFC pilus of Citrobacter rodentium, and the bundle-forming pilus, BFP, of enteropathogenic E. coli (EPEC) (13, 29, 41, 46).

TCP biogenesis requires the products of at least 10 TCP-specific genes (tcpA, tcpB, tcpQ, tcpC, tcpR, tcpD, tcpS, tcpT, tcpE, and tcpJ), which are located, along with tcpF, in the tcp operon (23, 24, 27). In-frame deletion mutants of each of these tcp genes, though not tcpF, fail to assemble TCP (24). The tcpF gene encodes a colonization factor, TcpF, that is secreted via the TCP apparatus (24). One of the tcp genes, tcpT, encodes the cognate putative ATPase (20, 21). The presence of similar cognate ATPases, termed type II secretion ATPases, is a conserved feature found to be essential for Tfp biogenesis, and related type II secretion systems (T2SSs), in numerous gram-negative bacteria (for a review, see reference 37). TCP biogenesis has been shown to be dependent on TcpT (5). TcpT, like all Tcp and T2SS ATPases, does not contain any predicted transmembrane domain and is therefore hypothesized to peripherally associate to the TCP apparatus via direct interaction with an inner membrane (IM) protein component(s). TcpD, TcpE, TcpJ, and TcpR are the four tcp-encoded predicted IM proteins of the TCP apparatus. Topology prediction programs such as TMHMM (http://www.cbs.dtu.dk/services/TMHMM/) (43) indicate a bitopic topology for TcpD and TcpR and a polytopic topology for TcpJ and TcpE.

In this study, we provide evidence for the inner membrane localization of TcpT. TcpT was mislocalized to the cytoplasm in a tcpR deletion strain but not in tcpD or tcpE deletion mutants, indicating the necessity of TcpR for membrane tethering of TcpT. We have demonstrated interaction between TcpR and TcpT by using metal affinity pull-down experiments. This interaction was shown to be direct using TcpR-PhoA and

* Corresponding author. Mailing address: Dartmouth Medical School, Department of Microbiology and Immunology, HB7550, Hanover, NH 03755. Phone: (603) 650-1632. Fax: (603) 650-1318. E-mail: ronald.k.taylor@dartmouth.edu.

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TABLE 1. Strains and plasmids used in this study

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TcpT expressed in *E. coli*. The interactive domains of each were defined with a LexA-based bacterial heterologous-protein two-hybrid system. The ability to make homodimers was demonstrated for the C-terminal domain (CTD) of TcpT by using the LexA-based homologous-protein two-hybrid system.

MATERIALS AND METHODS

Bacterial strains, plasmids, culture conditions, and DNA manipulations. The strains and plasmids used in this study are listed in Table 1 except for the in-frame tcp deletion mutants, which have been reported previously (6, 24). *V. cholerae* and *E. coli* strains were maintained at ~70°C in LB medium containing 30% (vol/vol) glycerol. *E. coli* cultures were grown in LB broth with a starting pH of 7.0 at 37°C for 12 to 16 h. In order to express TCP, *V. cholerae* strains were grown at 30°C in LB broth with a starting pH of 6.5 for 12 to 16 h. Antibiotics, when necessary, were used at the following concentrations: ampicillin (Ap), 100 μg/ml; tetracycline (Tc), 3 μg/ml; chloramphenicol (Cm), 15 μg/ml; tetracycline (Tc), 15 μg/ml; streptomycin (Sm), 100 μg/ml. When required, IPTG (isopropyl-β-D-thiogalactopyranoside) was added to LB broth to 1 mM and arabinose was added to the growth media at concentrations ranging from 0.02% to 0.1%. All DNA manipulations were performed by standard molecular and genetic techniques (3, 26).

**Subcellular fractionation.** To prepare whole-cell extracts, cultures grown for 12 to 16 h were centrifuged and cells were resuspended in 2% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) buffer with or without 2-mercaptoethanol and bromophenol blue (the latter to be used for protein estimation) and were boiled for 10 min. For membrane fractionation, a modification of a protocol based on differential solubility of membrane proteins in 2-mercaptoethanol and bromophenol blue was used to separate soluble cytoplasm from insoluble total membrane fraction. The insoluble membrane fraction was resuspended in 1 ml of 10 mM Tris-Cl–10 mM NaCl (pH 8.0) containing 2.5% Sarkosyl, followed by a 30-min incubation on a platform rocker at room temperature. The insoluble outer membrane (OM) proteins were separated from the soluble IM proteins by ultracentrifugation at 200,000 × g for 1 h at 12°C. Pellets containing OM fractions were resuspended in 20 μl of 10 mM Tris-Cl (pH 8.0).

The purity of subcellular fractions was determined with immunoblotting and enzyme assays for known localization markers, as follows. EpsL and TcpC immunoblots were used as IM and OM localization markers, respectively (6, 38). As a cytoplasmic marker, glucos-6-phosphate dehydrogenase (G6PDH) activity was assayed (18). G6PDH substrate vials containing glucos-6-phosphate along with the cofactors NADP, PMS, and 2,6-dichlorophenol indophenol (DCP-IP) (Sigma) were reconstituted with 0.3 M Tris-Cl (pH 8.0) just prior to use, as recommended by the manufacturer. One milliliter of a 1:2 dilution of cytoplasmatic fraction or 1 ml of IM fraction was added to a 4.5-ml cuvette containing the reconstituted substrate, immediately followed by an overlay of 2 ml of mineral oil. The decrease in the optical density at 600 nm (OD600) was measured at 1-min intervals for 5 min to monitor the reduction of blue-colored DCP-IP to a colorless form (2). Specific activity was calculated as nanomoles of DCP-IP reduced per minute per milligram of total protein.

**SDS-PAGE and immunoblot analysis.** Protein concentrations of whole-cell extracts and subcellular fractions were determined with a BCA Protein Assay Kit (Pierce). Samples containing equal amounts of total protein were resolved by 12.5% SDS-PAGE. Proteins were transferred to nitrocellulose membranes and probed individually with polyclonal antipeptide or monoclonal antibodies. Reactive bands were visualized with ECL detection reagents (Amersham Pharmacia).

**Metal affinity chromatography.** Cells were grown overnight under TCP-expressing conditions with or without the addition of 0.2% arabinose to induce expression of TcpR-6xHis. Cells were harvested and resuspended in extraction buffer (50 mM sodium phosphate–300 mM sodium chloride, pH 7.0) containing 1% vol/vol Triton X-100, 20 mM MgCl2, and DNease added to a final concentration of 20 μg/ml. The cells were broken by three passages through a French pressure cell at 8,000 lb/in2. Unbroken cells were removed by low-speed centrifugation. The lysates were incubated on a platform rocker at room temperature for 30 min. The clear lysates were mixed at a 1:1 ratio with TALON metal affinity
resin (Clontech) and allowed to mix on a rotating shaker for 1 h at 4°C. The protein purification protocol specified by the manufacturer was followed. The elution fraction was obtained with an extraction buffer containing 150 mM imidazole.

β-Galactosidase assay. For LexA-based homodimerization/heterodimerization experiments, expression of sulA:lexZ fusions was assayed from *E. coli* cultures grown at 37°C in LB, pH 7.0, containing 1 mM IPTG to an OD600 of approximately 0.8 (10). β-Galactosidase activity was expressed as Miller units (28).

Protein localization prediction. The TMHMM program (http://www.cbs.dtu.dk/services/TMHMM/) (43) was used to predict the transmembrane helix topology of TcpR, TcpD, TcpJ, and TcpE.

**RESULTS**

TcpT is detectable in all of the individual TCP biogenesis mutants except the ΔtcpT strain. The stability of the cognate ATPases of some Tfp systems and those of T2SSs have been shown to be affected by the absence of one or more accessory protein components of the cognate systems (4, 32, 36, 40). These reports led us to investigate the steady-state levels of TcpT in in-frame deletion mutants of each of the tcp genes. We observed similar abundances of TcpT in all of the IM tcp biogenesis mutants, i.e., in the ΔtcpR, ΔtcpD, ΔtcpE, and ΔtcpJ strains, compared to the wild-type (WT) strain (Fig. 1). However, we repeatedly noticed a decrease in the detectable amount of TcpT in a ΔtcpS strain and a slight increase in the detectable amount of TcpT in a ΔtcpA strain, as seen in Fig. 1. We did not observe any reproducibly significant difference in steady-state levels of TcpT in any of the other biogenesis mutants. The detectable levels of TcpT in all of the TCP biogenesis mutants, including the ΔtcpS strain, provided us the opportunity to determine the cellular localization of TcpT in each of the tcp biogenesis mutants.

TcpT is localized to the IM of WT *V. cholerae* but is mislocalized to the cytoplasm in a ΔtcpR strain. A previous model of TCP biogenesis (27) proposed that TcpT was localized to the IM via TcpE; however, this had not been examined experimentally. To test this model, we used subcellular fractionation to first demonstrate that the majority of TcpT is in fact localized to the IM in WT *V. cholerae* (Fig. 2A). The subcellular fractionation protocol was able to provide fairly pure fractions, as shown by the controls in Fig. 2A. TcpT was localized to the IM in each of the IM TCP biogenesis mutants except for the ΔtcpR strain, in which it was found to be almost entirely in the cytoplasm (Fig. 2B). Therefore, TcpR appears to be the major component that tethers TcpT to the IM. Complementation of the ΔtcpR strain using an arabinose-inducible pTcpR-6xHis construct restored IM localization of TcpT, as shown in Fig. 2C. Expression of this TcpR-6xHis construct in the ΔtcpR strain partially restored the WT TCP-associated phenotypes, such as autoagglutination, CTXΦ transduction, and TcpF secretion, indicating that His-tagged TcpR is functional (data not shown).

**Steady-state levels of TcpT are higher in a ΔtcpA strain than in the WT strain.** As shown in Fig. 1, there appeared to be a greater amount of TcpT detected in a ΔtcpA (pilin-negative) strain than in the WT. In order to determine if this was due to an overall increase in the steady-state level of membrane-bound TcpT or if it was also accompanied by mislocalization of TcpT, we performed a quantitative Western blot analysis of the cytoplasmic and IM fractions of a ΔtcpA deletion mutant complemented with pTcpR-6xHis grown under inducing and noninducing conditions and probed with anti-TcpT antibody.
normal in the ΔtcpA strain (6, 24). Moreover, there have not been any reports of strains or conditions wherein the steady-state level of any of the TCP biogenesis apparatus components is either increased or decreased due to a selective regulatory effect on gene expression. Therefore, it is likely that TcpT turnover is decreased specifically in the absence of pilin, as no other biogenesis mutants exhibit this effect.

TcpT interacts with TcpR. To determine if the role of TcpR in tethering TcpT to the IM is through interaction between the two proteins, we utilized metal affinity pull-down experiments to determine if TcpR-6xHis could pull down TcpT. Cell lysates were prepared from a ΔtcpR strain and the same strain complemented with pTcpR-6xHis grown under tcp-inducing conditions in the presence or absence of arabinose. Membrane proteins were solubilized with Triton X-100, and the resulting lysates were incubated with TALON metal affinity resin. The flowthrough and eluates from the metal affinity resin were subjected to immunoblot analysis (Fig. 4). The tcpR deletion strain served as a control to indicate that TcpT does not have any inherent affinity for binding to the metal affinity resin in the absence of TcpR. As seen in Fig. 4, upon induction, TcpR-6xHis was able to bind to the metal affinity resin and fractionate with the eluate, as shown by the anti-His immunoblot. Moreover, TcpR-6xHis was able to pull down TcpT, as indicated by coelution of TcpT. These data suggest that TcpR and TcpT are able to interact with each other in either a direct or an indirect manner.

TcpR is sufficient to direct TcpT to the IM but cannot tether it in a stable form in the absence of additional components. To address the ability of full-length TcpT to interact with full-length TcpR in the absence of other TCP proteins, we utilized E. coli as a heterologous system and attempted to coexpress TcpR-6xHis and TcpT. Our aim was to investigate TcpR-dependent IM localization of TcpT in the absence of the rest of the TCP proteins. We used the same TcpR-6xHis construct that was used to complement the tcpR deletion strain. However, we found that the TcpR-6xHis protein was undetectable when expressed in E. coli grown at either 30°C or 37°C, indicating that TcpR might be stabilized or stably held in the IM by another V. cholerae protein(s) (Fig. 5A). Consistent with this idea, the V. cholerae ΔtcpP strain complemented with the TcpR-6xHis construct expressed detectable amounts of TcpR-6xHis when the cells were grown under tcp-inducing Conditions (30°C) but not when the expression of the tcp operon was only partially expressed (37°C) (Fig. 5A). Finally, we used a tcpP deletion strain, which lacks a transcriptional activator of toxT expression (30). ToxT is the direct transcriptional activator of the tcp operon (16). We found that TcpR-6xHis could not be detected at either temperature in this strain (Fig. 5A).

One of the possible reasons for the observed instability of TcpR could be its unique predicted topology, with the IM domain at the extreme C terminus, leaving only one amino acid predicted to be exposed to the periplasm. Thus, it is likely that other IM proteins interact with TcpR to stabilize it in the IM. Interaction among the IM proteins of T2SSs via the IM domains has been reported previously with respect to the GSP (general secretion pathway) (which encompasses a T2SS of V. cholerae (39)). As we wanted to investigate whether TcpR on its own could direct TcpT to the IM, we attempted to stabilize TcpR by fusing it to PhoA at its C terminus. The phoA allele used to create this fusion lacks its own signal sequence. As shown in Fig. 5B, the fusion protein (corresponding to a size of about 49 kDa) was indeed stably localized to the IM of E. coli with or without coexpression of TcpT, as detected by subcellular fractionation followed by anti-PhoA immunoblotting.

The fusion construct also produced alkaline phosphatase activity (data not shown), confirming the predicted topology of TcpR, i.e., the C terminus being exposed to the periplasm. As predicted, the expression of a tcpT-encoding construct alone resulted in localization of TcpT to the cytoplasm in E. coli (Fig. 5C). The coexpression of TcpT and TcpR-PhoA in E. coli resulted in the disappearance of the band corresponding to TcpT from the cytoplasmic fraction. Intriguingly, instead of concomitant localization of TcpT to the IM fraction, a complete disappearance of the band corresponding to the full-length TcpT protein was observed (Fig. 5C). Moreover, we observed the appearance of a new band (designated TcpT*), which is most likely a degradation product of TcpT, localized to the IM fraction of these cells (Fig. 5C). Taken together, these results suggest that TcpR-PhoA is able to interact with TcpT in the absence of other TCP components and is able to direct tethering of TcpT to IM per se but not in a stable form. Furthermore, we recapitulated these results for V. cholerae by using the ΔtcpP strain. We observed that expression of tcpT in this strain resulted in stable TcpT with a localization defect (Fig. 5D). Coexpression of TcpT with TcpR-PhoA in the ΔtcpP strain resulted in the complete disappearance of TcpT from both the IM and cytoplasmic fractions. Consistent with the E. coli results, the majority of TcpR-PhoA was localized to the IM.
fraction in this strain (Fig. 5D, bottom). Conversely, expression of TcpR-PhoA was able to complement a ΔtcpR strain, as shown by nearly complete restoration of localization of TcpT to the IM (Fig. 5D) as well as by partial restoration of other functional TCP phenotypes (data not shown). Taken together, these results show that TcpR is sufficient to direct TcpT to the IM but that stable localization of TcpT to the IM and the stability of TcpR itself require other V. cholerae-specific factors, most likely other IM components of the TCP biogenesis apparatus.

The cytoplasmic N-terminal domain of TcpR interacts with the N-terminal domain of TcpT. TcpR is a relatively small protein, consisting of 151 amino acid residues. Localization and topology prediction programs such as TMHMM (43) predict a bitopic TcpR topology in which TcpR spans the IM a single time, with the N-terminal 130 residues residing in the cytoplasm and only the last C-terminal residue being exposed on the periplasmic side. Thus, we hypothesized that the cytoplasmic 130-amino-acid region of TcpR might include the domain that directly interacts with TcpT. Unlike TcpR, TcpT is a bigger protein, consisting of 503 amino acid residues. We performed a primary sequence alignment of TcpT with the cognate ATPases of similar systems, such as EpsE, of the T2SS of V. cholerae, and OutE, of the T2SS of Erwinia chrysanthemi, and found three distinct domains of the proteins with respect to sequence conservation. The first domain, the N-terminal 100-amino-acid domain, represents a region of weak alignment (5 to 6% identity and 12 to 16% similarity). This is followed by the middle, more-conserved region, from amino acid 101 to 350 (25 to 26% identity and 40 to 42% similarity), which includes the predicted conserved sequence of catalytic importance, and then the CTD, from residues 351 to 503, which exhibits an intermediate alignment (18 to 19% identity and 30 to 33% similarity). It has been shown that the cognate membrane protein association domains of EpsE and OutE are present within the N-terminal 100 amino acid residues, whereas the C-terminal region is involved in homodimerization (34, 37). To investigate the possibility of the first (N-terminal) 100 residues of TcpT interacting with the predicted cytoplasmic domain of TcpR, we created LexA fusion constructs of various domains of TcpR and TcpT for use in a bacterial two-hybrid protein system (10) (Fig. 6A). This system allows detection of heterodimeric interactions because (for example, in our case), the TcpR cytoplasmic fragment is fused to a WT LexA DNA binding domain (DBD) that recognizes a WT half-site and the TcpT fragment is fused to a WT LexA DNA binding domain (DBD) that recognizes a mutant half-site. The reporter sulA::lacZ carries a hybrid LexA binding site. One half-site is WT and the other is recognized only by the mutant LexA DBD that recognizes a mutant half-site. The reporter sulA::lacZ carries a hybrid LexA binding site. One half-site is WT and the other is recognized only by the mutant LexA DBD (10). Plasmids encoding fusion proteins carrying different domains of TcpT and TcpR (Fig. 6A) were cotransformed into the E. coli SU202 strain carrying a sulA::lacZ reporter gene with the hybrid LexA binding site. The possible interactions were measured in terms of repression of sulA::lacZ expression (Fig. 6B). Each sample was also analyzed by immunoblotting with anti-LexA antibodies to determine whether the hybrid constructs were all expressed at similar levels. As shown in Fig. 6B, the N-terminal 100-amino-acid-residue-containing domain of TcpT, in combination with the cytoplasmic domain of TcpR, resulted in almost 20-fold repression of sulA::lacZ expression compared to the vector control and the control strains expressing either construct individually. The LexA fusion construct
FIG. 6. The N terminus of TcpT interacts with the predicted cytoplasmic domain of TcpR, as demonstrated by a LexA-based two-hybrid protein system. (A) Representation of various TcpT and TcpR constructs fused to the N terminus of LexA. The letter designation for each of the constructs corresponds to the data shown in panel B. The grey box represents the transmembrane domain of TcpR. (B) E. coli SU202 carrying various constructs was grown to an OD 600 of approximately 0.8 in the presence of 1 mM IPTG, and expression of sulA::lacZ was measured by β-galactosidase assays. n, negative control, i.e., coexpression of empty vectors; a, TcpR (amino acids 1 to 130) cytoplasmic domain; b, TcpT (amino acids 1 to 100) N-terminal domain; c, TcpT (amino acids 101 to 503) CTD; d, TcpT full-length protein; e, coexpression of a and b; f, coexpression of a and c; g, coexpression of a and d. The stability of each of the fusion proteins was investigated in triplicates) is shown at the base of the graph for each of the respective strains.

In the present study, using biochemical and genetic approaches, we have investigated the stability, localization, and protein-protein interaction of TcpT with respect to the TCP biogenesis apparatus components. We present evidence for IM localization of the cognate ATPases of other Tfp and T2SS complexes (4, 8, 32, 34, 40). We used a collection of in-frame tcp single-gene deletion mutants to investigate the requirement of the corresponding TCP components in tethering TcpT to the IM and found that TcpR was necessary for TcpT association with the IM. TcpR is a predicted bitopic IM protein that is a GspL homolog (33). Our results add to a common theme emerging from various studies showing IM localization of the cognate ATPase by a bitopic IM protein, GspL, of the IM platform of a T2SS or a Tfp (4, 8, 32, 35, 40).

**DISCUSSION**

in Fig. 6B to compare sample panels a and c with f). This result indicates that the TcpT CTD (amino acids 101 to 503) may contain another TcpR interaction site that could result in a nonproductive interaction, i.e., leading to instability of the fusion proteins. Coexpression of the TcpR cytoplasmic domain fusion protein with full-length TcpT failed to exhibit a strong interaction, as indicated by only a twofold repression of sulA::lacZ expression (Fig. 6B). Similar results for a full-length ATPase fusion construct interaction with its respective IM protein fusion construct in a yeast two-hybrid system have been reported for OutE (34). The inability of the full-length protein to exhibit a strong positive interaction with the cytoplasmic domain of TcpR could be due to the fact that TcpT also exhibits characteristic homodimerization involving the CTD of the protein (see below), which may interfere with the heterodimerization assay. Alternatively, it is possible that the full-length TcpT requires the IM domain of TcpR in order to interact with TcpR.

The C terminus of TcpT can mediate homodimerization. Using a LexA-based homologous two-hybrid system (10), we investigated the domains involved in possible multimerization of TcpT. As shown in the Fig. 7A, approximately half of the TcpT protein from either the C terminus or the N terminus was fused with the N terminus of the WT LexA DBD by using pSR662 (10). Possible homodimerization was quantified in terms of repression of lacZ expression in E. coli SU101 carrying a sulA::lacZ reporter gene containing two WT LexA half-sites in the promoter region. As shown in Fig. 7B, the fusion protein containing the C-terminal half, but not the N-terminal half, of TcpT exhibited a strong repression of sulA::lacZ, indicating homodimerization, similar to that seen with the AraC fusion construct used as a positive control. However, full-length TcpT failed to exhibit sulA::lacZ repression, similar to the results reported previously for OutE (34). It is not clear why full-length TcpT would not mediate homodimerization and thus repression of sulA::lacZ as well as the CTD alone. One speculation is that the bulkier size of the full-length protein might cause a steric hindrance between two LexA monomers that interferes with DNA binding, a possible limitation of the LexA-based hybrid protein system that has been discussed previously (10).
In this study, we have demonstrated interaction between TcpT and TcpR by using metal affinity pull-down experiments. In order to investigate whether this interaction is direct or not, we coexpressed TcpR-PhoA and TcpT in the absence of other TCP proteins either in a heterologous system such as *E. coli* or in a ΔtcpP strain. In these cases, TcpR, by itself, directed TcpT to the membrane but in a manner that led to its degradation. Such conformational changes upon interaction between the ATPase and the cognate IM proteins of both a T2SS and a Tfp have been reported, such as the examples of the ATPases BfpD and EpsE becoming unstable when coexpressed with the cognate bitopic IM proteins BfpC and EpsL, respectively (1, 8).

The use of a LexA-based two-hybrid system that is designed to investigate heterodimerization among proteins allowed us to determine the regions of TcpT that are capable of interacting with the cytoplasmic domain of TcpR. We showed that the direct stable interaction between TcpR and TcpT involves the N-terminal portion (amino acids 1 to 100) of TcpT and the cytoplasmic domain of TcpR. This result is consistent with the evidence for other GspE-GspL cognate interactions in T2SSs (32, 34, 40). The second interaction between TcpT and TcpR seems to involve the CTD (amino acids 101 to 503) of TcpT and the cytoplasmic domain of TcpR. Although the LexA fusions containing these two regions of TcpT and TcpR did not exhibit a positive interaction, as indicated by the expression of *sulA*:lacZ reporter system, the coexpression of these fusion proteins resulted in instability of both, indicating a nonproductive or antagonistic interaction between these two regions of TcpT and TcpR. Therefore, in addition to the first N-terminal 100 residues, TcpT (amino acids 101 to 503) might contain a second site of TcpR interaction that could be a physiologically significant interaction. The former interaction might form a stable complex between TcpR and TcpT at the IM, whereas the latter (TcpT [amino acids 101 to 503]-TcpR) interaction might result in a conformational change in both of the proteins, which might mimic an active (ready for ATP hydrolysis) complex that needs to be stabilized by other TCP components. In the case of BFP, noted above, the reason for destabilization of BfpD upon coexpression with BfpC was shown to be due to mutual conformational changes in both proteins (8). The regions of BfpD interacting with BfpC are not yet mapped.

Based on our results, we predict that a nonproductive interaction such as that reported here (TcpT [amino acids 101 to 503]-TcpR) might cause mutual conformational changes, resulting in destabilization of both BfpD and BfpC. Consistent with this notion, Abendroth et al. attempted to crystallize the EpsE ATPase bound to the cytoplasmic domain of the cognate IM protein EpsL and found that the full-length ATPase was by β-galactosidase assays. n, negative control, i.e., empty vector; p, positive control AraC-LexA construct; a, TcpR cytoplasmic domain (amino acids 1 to 130); b, TcpR transmembrane domain (amino acids 131 to 151); c, TcpT N-terminal domain (amino acids 1 to 250); d, TcpT CTD (amino acids 251 to 503); e, TcpT full-length protein. The stability of each of the fusion proteins was detected by anti-LexA immunoblotting, and a composite that includes a representative sample for each strain (the strains were analyzed in triplicate) is shown at the base of the graph for each of the respective strains.
highly susceptible to proteolysis, resulting in a small stable fragment consisting of the N-terminal 1- to 88-amino-acid domain of the protein that could be co-crystallized with the cytoplasmic domain of EpsL (1). Based on our results and these reported studies, we predict that an additional GspL interaction site(s), besides the N-terminal 100 residues in the cognate ATPases of Gsp (GspE), could indeed be a common feature of the interaction of the cognate ATPases with the cognate bitopic membrane proteins.

Using a LexA-based homodimerization system, we demonstrated the ability of the C terminus of TcpT to mediate homodimerization. Homodimerization has been reported for various GspE homologs of T2SSs (32, 34, 48). It is becoming clear that most of these cognate ATPases of T2SSs, as well as the homologs of Tcp, such as BfpD, are capable of forming homo-hexamers (7, 9, 37, 42). Homodimerizations of the TcpR homolog of the BFP IM platform, BfpC, and T2SS (GspL) have also been reported (8, 32, 34, 38). Using the LexA-based homodimerization system, we were unable to detect TcpR homodimerization mediated by either the cytoplasmic domain or the membrane domain of TcpR. However, these results could reflect limitations of the LexA-based homodimerization system to detect interactions among transmembrane proteins. Therefore, additional approaches will be needed to investigate the possible homodimerization of TcpR.

An interesting outcome of our study was the finding that, in the absence of pilin (in a tcpA deletion strain), the steady-state levels of TcpT increased with no gross mislocalization. To explain the relationship between the pilin monomer and TcpT turnover, one hypothesis is that there is a direct interaction between TcpT and the pilin (the substrate monomers) that induces a conformational change in TcpT, perhaps analogous to that seen for SecA and its substrates (11, 31). This TcpA-mediated TcpT conformational change might affect TcpT stability. Another possibility is that in the absence of TcpA, the conformational status of the TCP IM platform is different in such a way that the TcpR-mediated tethering of TcpT results in an increasingly stable conformation of TcpT, resulting in increased steady-state levels of TcpT in the ΔtcpA strain.

We observed decreased steady-state levels of TcpT in a tcpS deletion strain. This result is entirely consistent with the BFP system, in which the absence of the cognate periplasmic protein BfpU resulted in a decrease in the detection of the cognate ATPase, BfpD, compared to the WT EPEC strain (36).

The fact that switching motility has not been observed under any experimental conditions for V. cholerae and that the tcp operon lacks a pilT (encoding an ATPase associated with retraction) ortholog and genes encoding more than one pseudo-pilin suggests that the TCP biogenesis apparatus structure and function might differ from those of the retractile Tfp elaborated by bacteria such as EPEC, P. aeruginosa, or N. gonorrhoeae. In agreement with this notion is the recent finding that Tcp gene clusters encoding the CFA/III pilus of ETEC and the CFC pilus of Citrobacter rodentium (29, 46) exhibit a greater resemblance to tcp in terms of the genetic organization of the respective operons. Furthermore, the primary sequences of the respective pilus apparatus proteins exhibited close analogy to the TCP proteins, indicating that TCP might represent a functional prototype of this sub-subclass of Tfp. Understanding the molecular interactions among TCP apparatus components and their impact on the assembly and function of the TCP apparatus is therefore a key to studying these specialized macromolecular assemblies, which are elaborated by various pathogenic bacteria.

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