Genome Sequence of the Plant Growth-Promoting Bacterium

Enterobacter cloacae GS1

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Here, we present the genome sequence of Enterobacter cloacae GS1. This strain proficiently colonizes rice roots and promotes plant growth by improving plant nutrition. Analyses of the E. cloacae GS1 genome will throw light on the genetic factors involved in root colonization, growth promotion, and ecological success of this rhizobacterium.

Enterobacter cloacae GS1, a rice root-colonizing bacterium, was enriched in planta from a rhizospheric microbial suspension. This strain has been deposited in the Microbial Type Collection Centre, Chandigarh, India, with the accession number MTCC 5698. Earlier, we had shown that E. cloacae GS1 outcompetes other innate microbial flora, colonizes rice roots, and promotes plant growth (6). We sequenced the genome of E. cloacae GS1 to gain a better understanding of the genes involved in rhizosphere colonization.

Total genomic DNA from a stationary-phase culture of E. cloacae GS1 was isolated using the Qiagen DNeasy protocol according to the manufacturer’s instructions. A total of 298,439 reads with an average read length of 526 bp were generated by Roche 454 pyrosequencing at the Research and Testing Laboratory, Lubbock, TX. The reads added up to 157,085,169 sequenced bases, indicating an ~34-fold coverage of the ~4.5-Mb genome. The obtained reads were de novo assembled using MIRA (Mimicking Intelligent Read Assembly) version 3.4 (2), which yielded 60 contigs (N50 length = 150 kb). The assembly was visualized using the Staden package version 2.0 (7), screened for misassemblies, and manually curated. Contigs with significant overlaps were joined based on consensus quality and coverage at the ends. Finally, 48 contigs were obtained, the longest and shortest of them being 1,527,101 bp and 545 bp, respectively.

The draft genome of E. cloacae GS1 is 4,500,707 bp long with a G+C content of 55.5%. The genome was annotated using the Rapid Annotations using Subsystems Technology (RAST) (1) server employing the GLIMMER gene caller. A total of 4,683 protein-encoding genes (PEGs) distributed in 548 metabolic subsystems were predicted along with 119 RNA coding regions. E. cloacae GS1 contains genes vital for motility, chemotaxis, adhesion, polysaccharide biosynthesis, and biofilm formation, which are required in various stages of root colonization (4). In the rhizosphere, survival of a bioinoculant depends on its ability to elicit density-dependent behavior and compete with rhizospheric microflora for the niche and nutrients. E. cloacae GS1 has the luxS gene encoding S-ribosylhomocysteine lyase, which produces the autoinducer-2 quorum sensing (QS) signal that regulates population density-dependent gene expression (5). Similar to other enteric bacteria, E. cloacae GS1 lacks a luxI homolog required for biosynthesis of N-acyl homoserine lactone (NAHL) QS signals. However, seven LuxR family transcriptional regulators were identified. The best studied of them is sdiA, which is involved in the detection of and response to environmental NAHLs (3). E. cloacae GS1 contains seven predicted β-lactamases, fusaric acid resistance determinants, and enterobactin production and uptake mechanisms, which could help in competition against soil microbiota. The existence of pathways for the synthesis of root elongation factors like indole acetic acid, 2,3-butanediol, and phosphate- soluble organic acids correlates with the rice growth-promoting ability of this strain. Thus, the E. cloacae GS1 genome encodes the traits essential for a successful bioinoculant.

Nucleotide sequence accession numbers. The Enterobacter cloacae subsp. cloacae GS1 whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession AJXP00000000. The version described in this paper is the first version, AJXP01000000.

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REFERENCES


