Electron Microscopy of *Treponema pallidum*
Occurring in a Human Primary Lesion

LEWIS M. DRUSIN, GABRIELLE C. ROUILLER, AND GEORGE B. CHAPMAN

Department of Medicine, Georgetown University Medical School, Washington, D.C. 20007, National Communicable Disease Center, Atlanta, Georgia 30333, and Department of Biology, Georgetown University, Washington, D.C. 20007

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Early electron microscope studies of *Treponema pallidum* (5, 7, 12, 18, 19) were of no value in defining the morphological aspects of the host-parasite interrelationship because observations were made only of bacterial cells derived from suspensions. Such preparations, obviously, destroyed the anatomical relationships which existed between the host and the bacterium. Improvements in techniques, e.g., a combination of shadow-casting procedures with enzyme digestion, did enable several investigators (2, 3, 6, 14, 15) to describe more clearly the structures of certain spirochetes. Thus, for example, the association of a fibrillar bundle with the bacterial cells was established. It remained, however, for Ovchinnikov and Delektorsky (11) to locate convincingly the fibrillar bundle between the cell wall and the plasma membrane, to describe fine structural features of the fibrils, and to clarify their numbers. Also, within the last 5 years, there have been several reports of electron microscope studies which included ultrathin sections of Nichols pathogenic or Reiter strains of *T. pallidum* either in situ or extracted from rabbit testis (1, 9, 10, 13). In an effort to extend our knowledge by describing the appearance in the electron microscope of ultrathin sections of *T. pallidum* in a human lesion, this study of a biopsy of a human chance, and this tissue was immediately fixed in 5.0% glutaraldehyde buffered at pH 7.4 with 0.2 M cacodylate buffer (8). Postfixation was accomplished in 1.0% OsO₄, similarly buffered. The samples were dehydrated in increasing concentrations of ethyl alcohol and embedded in Epon 812 (4). Ultrathin sections were cut with a DuPont diamond knife on an MT-2 Porter-Blum ultramicrotome, picked up on collodion-coated copper grids, stained with uranyl acetate (17), and examined in an RCA EMU-2D electron microscope.

It should be noted that, although the host tissue represents an inflammatory reaction, sarcoidosis is associated with alteration of the immune mechanism. Consequently, this lesion may not constitute a typical example of a true inflammatory reaction (16).

The helical form characteristic of *T. pallidum* is readily apparent (Fig. 1–4), although the frequency and amplitude of the sine wave pattern are variable. These changes are probably due to differences in the plane of the section and to differences in the degree of shrinkage of the specimens, as well as to real differences in these characteristics. In each section, the bacterial cells are located in the extracellular space. We have never observed them within the cytoplasm of a host cell. The bacteria are found both in amorphous extracellular substance (Fig. 1, 2) and in association with collagen fibrils (Fig. 3, 4). No definite preferential location for the bacterial cells could be determined. Occasionally (Fig. 1, arrow), one observes a clear area adjacent to the bacterial cell. Whether this represents a shrinkage space or an area from which some bacterial or host substance has been removed in the preparation of the material cannot now be determined.

A cell wall may be observed most clearly in one nearly longitudinal section (Fig. 4, arrow). The three-layered wall structure reported by Ryter and Pillot (13) could not be seen. The total wall thickness of about 14 nm did not fall within the range (7–9 nm) of wall thicknesses reported.
FIG. 1. *Treponema pallidum* located in amorphous extracellular ground substance. A clear space is discernible (arrow) adjacent to the cell. Three fibrils are visible (double arrow) and a bacterial and fibroblast cytoplasm (F) rich in ribonucleoprotein particles. × 28,000. The magnification mark represents 1 μm.

FIG. 2. *Treponema pallidum* located in amorphous extracellular ground substance, but proximal to areas of collagen. One prominent inclusion is seen (arrow) in the bacterial cytoplasm. × 28,800. The magnification marker represents 1 μm.

FIG. 3. *Treponema pallidum* located in a group of collagen fibrils. × 28,800. The magnification marker represents 1 μm.
FIG. 4. Sections through several different Treponema pallidum cells. A cell wall may be seen (arrow) and a cytoplasmic membrane (double arrow). The bacterial cells are here located in an area of collagen fibrils. × 28,800. The magnification marker represents 1 μm.
by those authors, and it was appreciably less than that measured (20 nm) from Fig. 4 of Ovchinnikov and Deletersky (9). A thin cytoplasmic membrane, reported by both Abe (1) and by Ryter and Pillot (13), may be seen (Fig. 4, double arrow). It is about 7 nm thick. Only one of our sections passed through a cell at a plane appropriate to reveal the three fibrils (Fig. 1, double arrow). This compares to the three to seven reported by Ryter and Pillot (13), the two to eight reported by Ovchinnikov and Deletersky (9), and to the three to four reported by Swain (15). Abe (1), on the other hand, reported seeing no fibrils in the Nichols strain of T. pallidum. In our sections, the diameter of each fibril was about 14 nm. Ryter and Pillot (13) reported a fibril diameter of 20 nm, whereas Ovchinnikov and Deletersky (9) reported a fibril diameter of 10 to 20 nm. Along with Ryter and Pillot (13), we could find no substructure in the fibrils. We were unable to confirm, in this material, the location of the fibrils between the cell wall and plasma membrane, as reported by Ryter and Pillot (13) and by Ovchinnikov and Deletersky (11). However, in ultrathin sections of testis from a syphilitic rabbit, containing Nichols pathogenic strain of T. pallidum, the fibrils were found between the cell wall and plasma membrane (L. M. Drusin and G. B. Chapman, unpublished data).

The cytoplasm of the bacterial cells appears to be packed homogeneously with 10- to 15-nm diameter ribonucleoprotein (RNP) particles. Ovchinnikov and Deletersky (9) report the same size range for RNP particles. The distribution of RNP particle sizes seems to be somewhat smaller in the bacterial cytoplasm than in the human fibroblast cells (F), appearing in the same microscopic field (Fig. 1).

Apparent size differences noted in this report may be due to variations in preparative procedures. It is also possible that differences in appearances, noted below, may be attributed either to similar variations or to the fact that preservation of structures of bacteria fixed in situ in human tissue is a different matter from the preservation of comparable structures of bacteria fixed in suspension.

Although several inclusion bodies are present in some of the bacterial cells, we prefer not to consider even the one in Fig. 2 (arrow) as being a mesosome-like structure. Similarly differentiated structures were thus described by Ryter and Pillot (13) and by Ovchinnikov and Deletersky (9). We, also, did not find structures which could be accurately described as basal granules or round, spore-like structures as they had been identified by Ovchinnikov and Deletersky (9) and by Ryter and Pillot (13). In reality, it appears that the basal granules, "b," in plate 6, Fig. 3 and 4 of Ovchinnikov and Deletersky (9), are the same structures which Ryter and Pillot (13) called mesosomes. Furthermore, these structures, labeled "b," as noted above, are much larger than the similarly labeled structures in plate 4, Fig. 1 (9). The structures labeled "b" in plates 1, 3, 4 (9) appear more likely to be basal granules.

In none of our electron micrographs could we identify nuclear material in these bacteria, an observation shared by Abe (1). This phenomenon is in contrast with the obvious nuclear material found in the investigation of Ryter and Pillot (13) and material considered by Ovchinnikov and Deletersky (9) to represent a nucleoid (plate 2).

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LITERATURE CITED