

## *Salmonella enteritidis* *agfBAC* Operon Encoding Thin, Aggregative Fimbriae

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*Salmonella enteritidis* produces thin, aggregative fimbriae, named SEF17, which are composed of polymerized AgfA fimbrin proteins. DNA sequence analysis of a 2-kb region of *S. enteritidis* DNA revealed three contiguous genes, *agfBAC*. The 453-bp *agfA* gene encodes the AgfA fimbrin, which was predicted to be 74% identical and 86% similar in primary sequence to the *Escherichia coli* curli structural protein, CsgA. pHAG, a pUC18 derivative containing a 3.0-kb *HindIII* fragment encoding *agfBAC*, directed the *in vitro* expression of the major AgfA fimbrin, with an  $M_r$  of 17,000, and a minor AgfB protein, with an  $M_r$  of 16,000, encoded by the 453-bp *agfB* gene. AgfA was not expressed from pDAG, a pUC18 derivative containing a 3.1-kb *DraI* DNA fragment encoding *agfA* but not *agfB*. Primer extension analysis identified two adjacent transcription start sites located immediately upstream of *agfB* in positions analogous to those of the *E. coli* curli *csgBA* operon. No transcription start sites were located immediately upstream of *agfA* or *agfC*. Northern (RNA) blot analysis confirmed that transcription of *agfA* was initiated from the *agfB* promoter region. Secondary-structure analysis of the putative mRNA transcript for *agfBAC* predicted the formation of a stem-loop structure ( $\Delta G^\circ$ , -22 kcal/mol [-91 kJ/mol]) in the intercistronic region between *agfA* and *agfC*, which may be involved in stabilization of the *agfBA* portion of the *agfBAC* transcript. *agfBAC* and flanking regions had a high degree of sequence similarity with those counterparts of the *E. coli* curli *csgBA* region for which sequence data are available. These data are demonstrative of the high degree of similarity between *S. enteritidis* SEF17 fimbriae and *E. coli* curli with respect to fimbrin amino acid sequence and genetic organization and, therefore, are indicative of a common and relatively recent ancestry.

Fimbriae are filamentous surface structures composed of a repeated major subunit protein (fimbrin) and sometimes several minor subunit-like proteins (24). Fimbrins are polymerized by hydrophobic and hydrophilic interactions to form either thick (7- to 8-nm), rigid structures, thin (2- to 4-nm), flexible filaments, or composites of both (13, 16, 19). The characterization of fimbrial biosynthesis has shown that the regulation and assembly of some fimbriae are accomplished by several accessory proteins (17). The genes encoding structural and accessory proteins are typically arranged in 7- to 9-kb clusters organized as a series of operons and individual genes (4, 7, 18, 23).

*Salmonella enteritidis* is an important, food-borne, enteric pathogen which produces several fimbrial types (5). Characterization of the various fimbriae of *S. enteritidis* 27655-3b led to the discovery of thin, aggregative fimbriae (SEF17) that mediate fibronectin binding (10, 11). SEF17 fimbriae are composed mainly of a fimbrin with an  $M_r$  of 17,000, AgfA. These fimbriae are highly stable structures, requiring treatment with 90% formic acid for depolymerization (11). Diarrheagenic *Escherichia coli* strains produce thin, aggregative fimbriae that are biochemically and serologically related to those of SEF17 (12). However, the degree of DNA sequence dissimilarity between the respective fimbrin genes is sufficient that *agfA*-based

nucleotide probes hybridize only to *Salmonella* DNA, thereby providing a valuable, genus-specific diagnostic for *Salmonella* spp. (14).

*E. coli* HB101 also produces an AgfA-related fimbrin (3). The corresponding thin, aggregative fimbriae have been named curli. *csgA*, the gene encoding the major curli subunit protein, has recently been cloned and sequenced (22). Transcriptional analysis revealed that *csgA* is transcribed as a dicistronic operon from the promoter of the upstream gene, *csgB* (2). The *csgBA* promoter requires an AT-rich upstream activating sequence, which is recognized by both  $\sigma^s$  and  $\sigma^{70}$  sigma factors and is repressed by H-NS, which prevents the formation of transcription initiation complexes with  $\sigma^{70}$  under conditions under which  $\sigma^s$  is not expressed, including temperatures above 26°C, high osmolarity, and non-stationary-phase growth (2).

The structural similarities of curli and SEF17 fimbriae and similarities in the N-terminal fimbrin sequences suggest that these related fimbriae are assembled by analogous mechanisms. In an effort to further understand the biosynthesis of SEF17 fimbriae, the DNA sequence of the *agfBAC* operon was determined and the expression of these genes was examined.

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### MATERIALS AND METHODS

**Bacterial strains and growth conditions.** *S. enteritidis* 27655-3b was routinely grown on solid T medium at 37°C (11). To assay for SEF17 production, Congo red (100 µg/ml) was incorporated into T medium (10, 12). *E. coli* recombinant

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clones were grown on Luria-Bertani medium containing ampicillin (100 µg/ml) (26).

**Plasmid construction.** *S. enteritidis* chromosomal DNA, purified by CsCl gradient centrifugation (26), was digested with *DraI* or *HindIII*. *AgfA*-encoding fragments were identified by hybridization to a <sup>32</sup>P-labeled (26), 374-bp, *AgfA*-encoding DNA fragment amplified from *S. enteritidis* *TnphoA* 2-7f by PCR (10). Hybridizing fragments were subcloned into pUC18 (29) and introduced into *E. coli* DH5α by transformation (26). Plasmids were purified by standard alkaline lysis procedures (26) or on Qiagen columns according to the manufacturer (Qiagen Inc., Chatsworth, Calif.).

**DNA sequence analyses.** DNA sequences were determined with Sequenase version 2.0 (United States Biochemical Corp., Cleveland, Ohio) and custom oligonucleotide primers synthesized on a PCR-MATE EP model 391 DNA synthesizer (Applied Biosystems Inc., Foster City, Calif.). GeneWorks version 2.0 (IntelliGenetics Inc., Mountainview, Calif.) was used to order overlapping DNA sequences. DNA Strider 1.1 (20) was used to identify open reading frames within the composite sequence. The programs of the PC/GENE suite (IntelliGenetics Inc.) were used to locate putative promoters (25) and Shine-Dalgarno motifs (15) and to predict RNA and protein secondary structures. The predicted amino acid sequence for each open reading frame was compared with protein sequences listed in the National Center for Biotechnology Information (NCBI) databases with the program BLASTX (1).

**SDS-PAGE and Western blot (immunoblot) analysis.** *AgfA* was solubilized from *S. enteritidis* or *E. coli* recombinant clones and detected by Western blot analysis according to the method of Collinson et al. (10, 11). Proteins contained in the sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) sample buffer-glycine extracts of whole cells and the formic acid-digested cell material were separated by SDS-PAGE, electrophoretically transferred to nitrocellulose, and screened with rabbit anti-SEF17 immune serum. Immunoreactive *AgfA* proteins were detected with goat anti-rabbit immunoglobulin G-alkaline phosphatase conjugates and visualized with 5-bromo-4-chloro-3-indolylphosphate and the enhancer Nitro Blue Tetrazolium.

**In vitro transcription-translation analysis.** Proteins encoded by *agfBAC* were labeled with [<sup>35</sup>S]methionine with a coupled, in vitro transcription-translation system (Promega Corp., Madison, Wis.). Plasmid templates (4 µg) were used in reaction mixtures, which were incubated first for 1.5 h at 37°C and then for 10 min at 0°C. A 20-µl aliquot was removed from the transcription-translation reaction mixture, added to 80 µl of acetone, and held on ice for 15 min. The acetone-precipitated proteins were recovered by centrifugation (14,000 × g, 5 min, 4°C), dried for 15 min under vacuum, resuspended in 50 µl of SDS-PAGE sample buffer, and heat denatured (100°C, 5 min). [<sup>35</sup>S]methionine-labeled proteins in a 15-µl aliquot were separated by SDS-PAGE. The 12.5% acrylamide gel was fixed in 7% acetic acid for 1 h, dried onto 3-mm-thick Whatman paper for 2 h at 80°C, and then exposed to Kodak X-Omat AR5 film for autoradiographic detection of labeled proteins.

**RNA extraction.** Total RNA was prepared from *S. enteritidis* cells grown on T medium overnight at 37°C by a modification of the procedure of McCormick et al. (21) as described by Clouthier et al. (9). Two gene probes for Northern (RNA) blot analysis were prepared by PCR amplification (10) of either a 192-bp *agfB* fragment, by using the primer pair TAF41 (5'-GCAAGTCTTCATTTAAT CAG3') and TAF43 (5'-CTGGCATCGTTGGCATTGCC3'), or a 183-bp *agfC* fragment, by using the primer pair TAF38 (5'-CGATATTTACACGGTGAT CC3') and TAF30 (5'-GTCTCTGAAGATATATTATCCA3'). A 394-bp *agfA* probe was prepared from an *EcoRI* fragment of pAGF3 (14) that was purified with the Sephaglas BandPrep kit (Pharmacia Biotech, Uppsala, Sweden). The gene probes were labeled with [<sup>32</sup>P]dATP by random priming (26).

**Primer extension analysis.** Custom sequencing primers complementary to the coding strands of *agfB* (TAF44, 5'-GTGCGGTTGCAATCCAG3'), *agfA* (TAF42, 5'-GTGGAACGACGCCCA3'), and *agfC* (TAF26, 5'-GTCTCT GAAGATATATTTACT3') were end labeled with [<sup>32</sup>P]ATP and used in primer extension reactions as previously described (9, 27).

**Nucleotide sequence accession numbers.** The nucleotide sequences reported herein for *agfB*, *agfA*, and *agfC* were submitted to GenBank and given accession no. U43280.

## RESULTS

**Cloning and sequencing of *agfBAC* region.** To identify chromosomal fragments encoding proteins responsible for SEF17 biosynthesis, genomic digests of *S. enteritidis* 27655-3b prepared with *HindIII* or *DraI* were probed with a PCR fragment containing a 333-bp *agfA* sequence. The hybridizing 3.0-kb *HindIII* fragment and the hybridizing 3.1-kb *DraI* fragment were each subcloned into pUC18 to form pHAG and pDAG, respectively (Fig. 1A). Sequencing of approximately 2 kb of DNA surrounding the overlapping region of the *HindIII* and *DraI* fragments resulted in recognition of the *agfA* gene and two flanking open reading frames (Fig. 1B). *agfA* was identified by comparison with a previously sequenced 333-bp region of

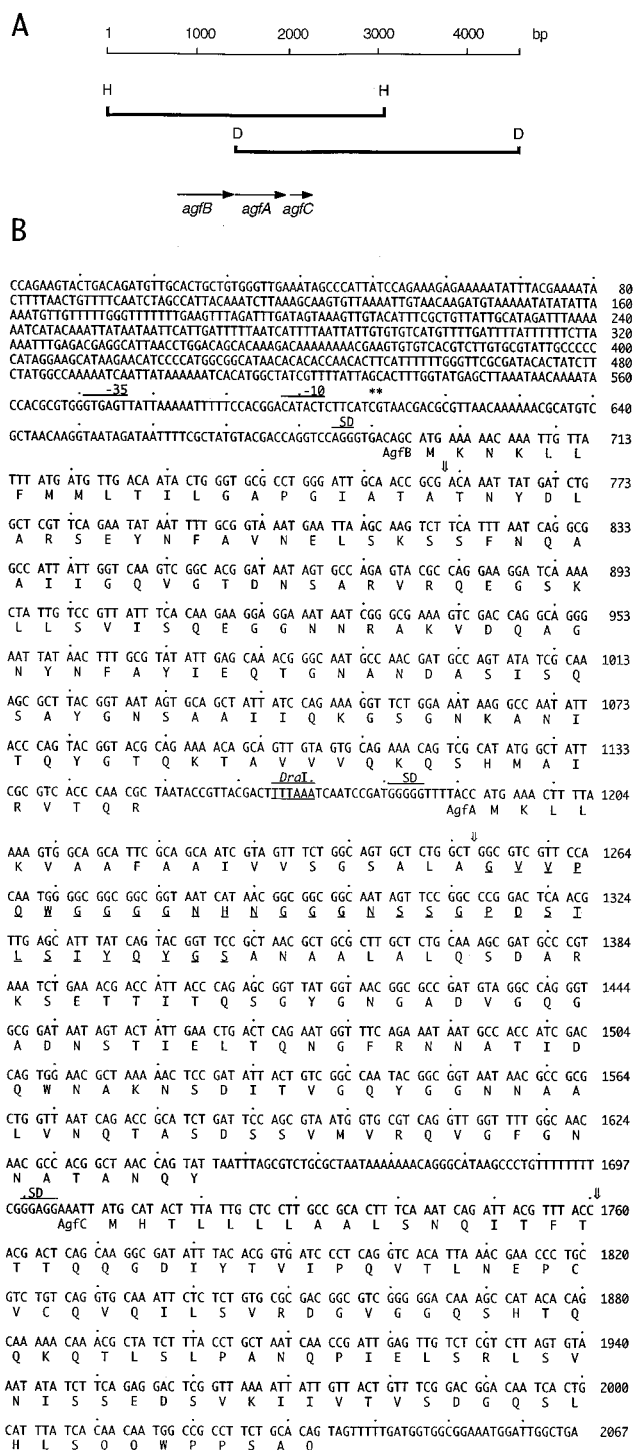


FIG. 1. Cloning and sequence determination of the region of the *S. enteritidis* chromosome surrounding *agfA*. (A) Schematic indicating the locations and orientations of *agfBAC* on two recombinant pUC18 plasmid derivatives, pHAG and pDAG, containing overlapping 3.0-kb *HindIII* (H) and 3.1-kb *DraI* (D) fragments, respectively. (B) DNA sequences and predicted amino acid sequences of *agfBAC*. The Shine-Dalgarno motifs (SD) and -35 and -10 promoter motifs (25) are overscored. Arrows mark the predicted signal sequence cleavage sites in *AgfB*, *AgfA*, and *AgfC*. The *AgfA* sequence previously determined by N-terminal amino acid sequence analysis is underscored. Transcriptional start sites (Fig. 4) are noted by asterisks.



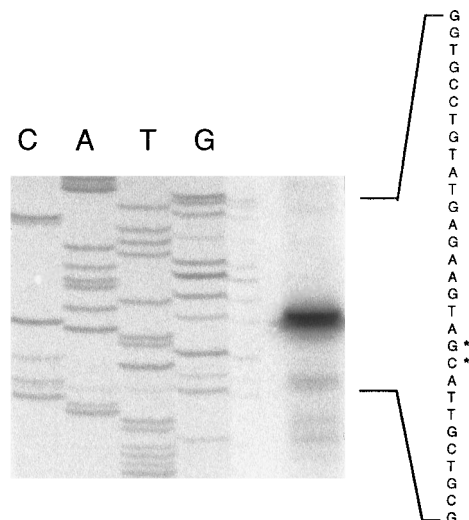


FIG. 4. Primer extension analysis to determine the transcription initiation site(s) for *agfBAC*. The primer TAF 44 (5'-GTCGCGGTTGCAATCCCAG3'), located 46 bp downstream of the *agfB* start codon, was used to direct sequencing reactions of *agfB* encoded by pHAG and to direct the primer extension reaction on total *S. enteritidis* RNA prepared as indicated in Materials and Methods. Asterisks mark the transcriptional start sites, which are located 85 and 86 nt upstream of the *agfB* start codon (Fig. 1).

*agfC* (Fig. 5B), which might stabilize the *agfBA* and *agfA* transcripts.

**Comparison of the *S. enteritidis agfBAC* and *E. coli csgBA* regions.** The DNA sequences of *agfBAC* and flanking regions were compared with the partial DNA sequences determined for the *E. coli csgBA* region (2, 22). The approximately 700-bp region immediately upstream of *agfB* had 63% similarity with sequences immediately upstream of *csgB* (Fig. 6A). The level of similarity was 76% over the 5' 275-bp regions, 39% over the central regions, and 82% in the 246-bp regions adjacent to the translational start codons of *agfB* and *csgB* (Fig. 6A). Notably, *agfBA* and *csgBA* shared the same two transcriptional start sites (Fig. 6A). The predicted N-terminal amino acid sequence of AgfB was identical to the 21-amino-acid N-terminal sequence predicted for CsgB (Fig. 6A). Similarly, 10 of the 12 C-terminal residues predicted for CsgB were identical to those of AgfB (Fig. 6B). The limited sequence data published for *csgB* (2) prevented further comparisons. The *agfBA* intergenic region had 93% similarity to the *csgBA* intergenic region (Fig. 6B). The sequence immediately downstream of *agfA* was 71% identical to the *agfAC* intergenic region (Fig. 6C). Comparison of *agfC* with the partially sequenced region downstream of *agfA* indicated that *E. coli* encoded an amino acid sequence of which 15 of 17 residues matched the N-terminal amino acid signal sequence predicted for AgfC (Fig. 6C).

## DISCUSSION

This study has resulted in the characterization of the *S. enteritidis agfBA* gene sequences which are required for the expression of AgfA, the major structural fimbrin component of an unusual, thin, aggregative fimbria, SEF17. *agfA* is transcribed as part of an operon which includes the upstream gene, *agfB*. A third open reading frame, *agfC*, also belongs to this operon since no transcriptional start site between *agfA* and *agfC* was found. Consistent with these experimental observations, no sequences similar to prokaryotic promoter motifs were recognized within the *agfBAC* intragenic regions. The

inability to detect an *agfC* transcript was likely indicative of a very short half-life of a portion of a transcript (4, 6). A potentially stable stem-loop structure in the intercistronic region between *agfA* and *agfC* could contribute to the stabilization of the *agfBA* transcript and the individual *agfA* transcripts (6, 8). The need for production of large amounts of the structural fimbrin AgfA proteins of SEF17 fimbriae is a plausible biological rationale for the processing and enhanced half-life of the *agfA* region of the *agfBAC* transcript.

The role of AgfA as the major structural fimbrin subunit protein of SEF17 is unequivocal (10, 11). The function of AgfB is unknown. According to the predicted amino acid sequences, AgfB matches AgfA in size and resembles AgfA in primary amino acid sequence. The differences between the two proteins include the presence of two tryptophan and proline residues in AgfA which are lacking in AgfB and the fact that AgfB has half the glycine residues of AgfA and more than double the number of the basic residues arginine and lysine. Although these differences between the primary structures exist, secondary-structure predictions indicate that these two proteins fold similarly (12a). These results suggest that AgfB is a fimbrin-like protein. *agfB* appears to encode a signal sequence characteristic of an exported protein, but AgfB did not copurify with AgfA in intact SEF17 in amounts detectable by N-terminal amino acid sequence analysis (11). This cannot be taken as proof that AgfB is absent from the fimbrial fiber, as AgfB may be a structural component of SEF17 that is present in the minute amounts characteristic of minor fimbrial tip proteins (17, 24) or as AgfB may be loosely associated with SEF17 filaments and therefore be readily dissociated during the rigorous, unconventional purification procedure used to obtain SEF17 filaments containing AgfA (11).

At present, it is unclear how many genes are required for the biosynthesis of SEF17. The *agfBAC* operon does not encode proteins characteristic of chaperone or usher proteins typically required for fimbrial biosynthesis (17), and analysis of the DNA sequences 0.7 kb upstream or 2.3 kb downstream of

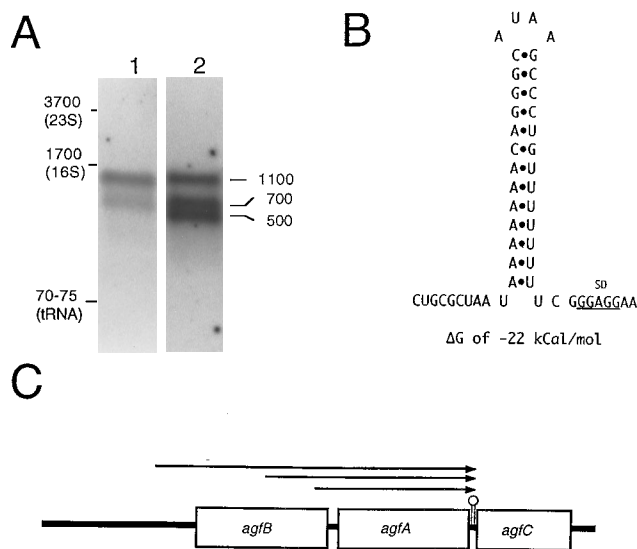


FIG. 5. Transcriptional analysis of *agfBAC*. (A) Total *S. enteritidis* RNA was probed by Northern blot analysis with gene probes to *agfB* (lane 1) and *agfA* (lane 2). The migrations of 23S RNA, 16S RNA, and tRNA species are noted on the left, and the approximate sizes (in nucleotides) of the transcripts are noted on the right. (B) The predicted stem-loop structure within the transcript arising from the *agfAC* intercistronic region is presented. (C) Schematic of the relative positions of the detected transcripts.

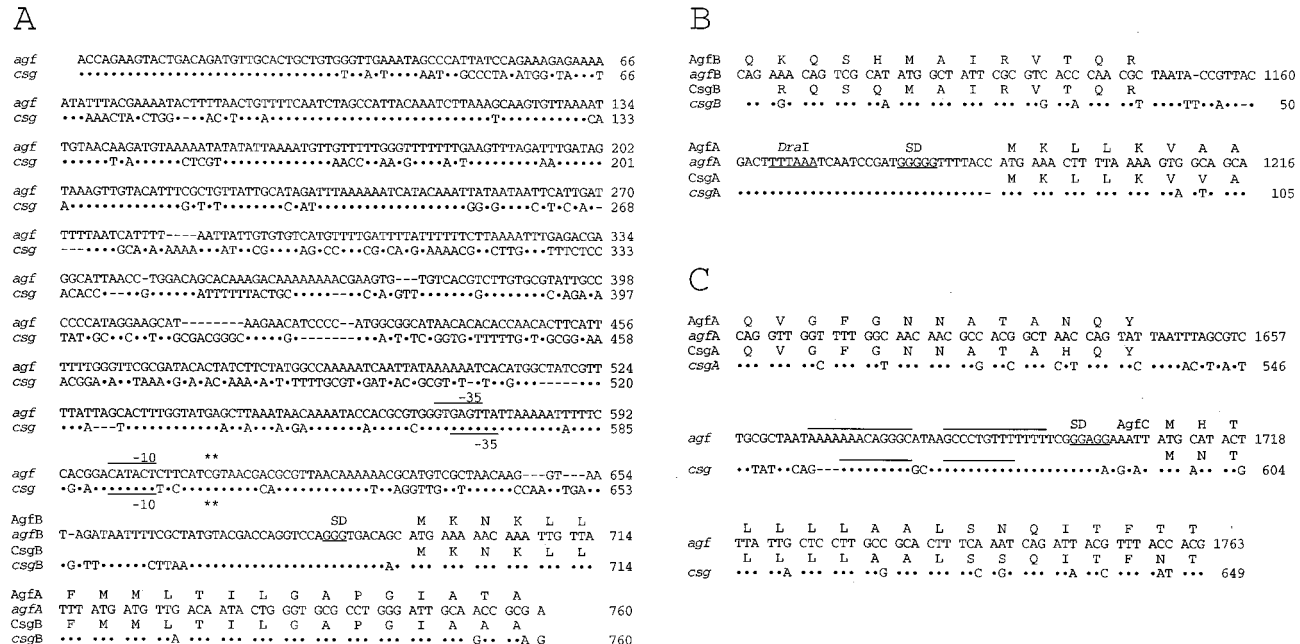


FIG. 6. DNA sequence comparison of the *S. enteritidis* 27655-3b *agfBAC* region with the *E. coli* *csgBA* region. Spaces (dashes) were introduced into both sequences to maximize alignments. Bases in the *E. coli* sequence matching those in the *S. enteritidis* sequence are denoted by dots. (A) Comparison of the *agfB* upstream region with that upstream of *csgB* (2). Promoter motif  $-10$  and  $-35$  regions (25) and a Shine-Dalgarno (SD) motif (15) for ribosome binding are underlined. The transcriptional start sites are noted with asterisks in both sequences. (B) Comparison of the *agfBA* intergenic region with the sequence upstream of *E. coli* *csgA* (2, 22). (C) Comparison of the *agfAC* intergenic region with the sequence downstream of *csgA* (22). Palindrome sequences indicative of potential stem-loop structures are overlined in both sequences.

*agfBAC* similarly did not identify open reading frames encoding other obvious fimbrial biosynthetic proteins (12a), suggesting that assembly and biosynthesis of SEF17 may proceed by a novel mechanism.

The data presented here support a close relationship between *S. enteritidis* SEF17 fimbriae and *E. coli* filaments called curli. The genetic organization of the *agfBAC* operon, which is partially responsible for SEF17 biosynthesis, closely resembles that of the *E. coli* *agfBA* region, which is necessary for the production of curli. The major structural protein genes, *agfA* and *csgA*, are homologous. Both genes are transcribed as part of an operon from promoters upstream of the respective flanking genes, *agfB* and *csgB* (2), which direct transcription from identical start sites. Although the complete DNA sequences for the genes flanking *csgA* of *E. coli* have not been published, it is clear from the sequence data available (2, 22) that *agfB* is homologous to *csgB*. These findings are not completely unexpected, since SEF17 and curli are morphologically, structurally, biochemically, and serologically related and are likely functionally similar as well (4, 11, 12, 22). However, the degree and extent of the DNA sequence conservation is somewhat surprising. The highly related regions of DNA sequence include intragenic regions, the 5' sequences of open reading frames considered to encode signal peptides typical of those of exported prokaryotic proteins (28), and large portions of the upstream regions, including that encompassing or corresponding to the *csgA* "upstream activating sequence" (2). These results provide strong evidence that the genes for SEF17 and curli biosynthesis have a common ancestry and that there was a relatively recent gene transfer to *E. coli*, to *S. enteritidis*, or to both from a common ancestor.

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#### REFERENCES

- Altschel, S. F., W. Gish, W. Miller, E. W. Myers, and D. J. Lipman. 1990. Basic local alignment search tool. *J. Mol. Biol.* 215:403-410.
- Arnqvist, A., A. Olsén, and S. Normark. 1994.  $\sigma^{70}$ -dependent growth-phase induction of the *csgBA* promoter in *Escherichia coli* can be achieved in vivo by  $\sigma^{70}$  in the absence of the nucleoid-associated protein H-NS. *Mol. Microbiol.* 13:1021-1032.
- Arnqvist, A., A. Olsén, J. Pfeifer, D. G. Russell, and S. Normark. 1992. The Crl protein activates cryptic genes for curli formation and fibronectin binding in *Escherichia coli* HB101. *Mol. Microbiol.* 6:2443-2452.
- Båga, M., M. Göransson, S. Normark, and B. E. Uhlin. 1988. Processed mRNA with differential stability in the regulation of *E. coli* pilin gene expression. *Cell* 52:197-206.
- Bäumler, A. J., and F. Heffron. 1995. Identification and sequence analysis of *lpfABCDE*, a putative fimbrial operon of *Salmonella typhimurium*. *J. Bacteriol.* 177:2087-2097.
- Belasco, J. G., and C. F. Higgins. 1988. Mechanisms of mRNA decay in bacteria: a perspective. *Gene* 72:15-23.
- Bilge, S. S., J. M. Apostol, Jr., M. A. Aldape, and S. L. Moseley. 1993. mRNA processing independent of RNaseIII and RNaseE in the expression of the F1845 fimbrial adhesin of *Escherichia coli*. *Proc. Natl. Acad. Sci. USA* 90:1455-1459.
- Chen, C.-Y. A., J. T. Beatty, S. N. Cohen, and J. G. Belasco. 1988. An intergenic stem-loop structure functions as an mRNA decay terminator necessary but insufficient for *puf* mRNA stability. *Cell* 52:609-619.
- Clouthier, S. C., K.-H. Müller, J. L. Doran, S. K. Collinson, and W. W. Kay. 1993. Characterization of three fimbrial genes, *sefABC*, of *Salmonella enteritidis*. *J. Bacteriol.* 175:2523-2533.
- Collinson, S. K., S. C. Clouthier, J. L. Doran, P. A. Bamser, and W. W. Kay. Characterization of the *agfBA* fimbrial operon encoding thin aggregative fimbriae of *Salmonella enteritidis*. In P. S. Paul, D. H. Francis, and D. Benfield

- (ed.), Mechanisms in the pathogenesis of enteric diseases. Proceedings of the First International Rushmore Conference on Pathogenic Mechanisms in Enteric Diseases. Rapid City, S.Dak.
10. Collinson, S. K., P. C. Doig, J. L. Doran, S. C. Clouthier, T. J. Trust, and W. W. Kay. 1993. Thin aggregative fimbriae mediate binding of *Salmonella enteritidis* to fibronectin. *J. Bacteriol.* **175**:12–18.
  11. Collinson, S. K., L. Emödy, K.-H. Müller, T. J. Trust, and W. W. Kay. 1991. Purification and characterization of thin, aggregative fimbriae from *Salmonella enteritidis*. *J. Bacteriol.* **175**:4773–4781.
  12. Collinson, S. K., L. Emödy, T. J. Trust, and W. W. Kay. 1992. Thin aggregative fimbriae from diarrheagenic *Escherichia coli*. *J. Bacteriol.* **174**:4490–4495.
  - 12a. Collinson, S. K., and W. W. Kay. Unpublished data.
  13. De Graaf, F. K. 1988. Fimbrial structures of enterotoxigenic *E. coli*. *Antonie van Leeuwenhoek* **54**:395–404.
  14. Doran, J. L., S. K. Collinson, J. Burian, G. Sarlós, E. C. D. Todd, C. K. Munro, C. M. Kay, P. A. Banser, P. I. Peterkin, and W. W. Kay. 1993. DNA-based diagnostic tests for *Salmonella* species targeting *agfA*, the structural gene for thin, aggregative fimbriae. *J. Clin. Microbiol.* **31**:2263–2273.
  15. Gold, L. 1988. Posttranscriptional regulatory mechanisms in *Escherichia coli*. *Annu. Rev. Biochem.* **57**:199–233.
  16. Gong, M., and L. Makowski. 1992. Helical structure of P pili from *Escherichia coli*. *J. Mol. Biol.* **228**:735–742.
  17. Hultgren, S. J., S. Normark, and S. N. Abraham. 1991. Chaperone-assisted assembly and molecular architecture of adhesive pili. *Annu. Rev. Microbiol.* **45**:383–415.
  18. Jordi, B. J. A. M., I. E. L. op den Camp, L. A. M. de Haan, B. A. M. van der Zeijst, and W. Gastra. 1993. Differential decay of RNA of the CFA/I fimbrial operon and control of relative gene expression. *J. Bacteriol.* **175**:7976–7981.
  19. Kuehn, M. J., J. Heuser, S. Normark, and S. J. Hultgren. 1992. P pili in uropathogenic *E. coli* are composite fibres with distinct fibrillar adhesive tips. *Nature (London)* **356**:252–255.
  20. Marck, C. 1988. "DNA Strider": a "C" program for the fast analysis of DNA and protein sequences on the Apple Macintosh family of computers. *Nucleic Acids Res.* **16**:1829–1836.
  21. McCormick, J. R., J. M. Zengel, and L. Lindahl. 1991. Intermediates in the degradation of mRNA from the lactose operon of *Escherichia coli*. *Nucleic Acids Res.* **19**:2767–2776.
  22. Olsén, A., A. Arnqvist, M. Hammar, S. Sukupolvi, and S. Normark. 1993. The RpoS sigma factor relieves H-NS-mediated transcriptional repression of *csgA*, the subunit gene of fibronectin-binding curli in *Escherichia coli*. *Mol. Microbiol.* **7**:523–536.
  23. Oudega, B., and F. K. De Graaf. 1988. Genetic organization and biogenesis of adhesive fimbriae of *Escherichia coli*. *Antonie van Leeuwenhoek* **54**:285–299.
  24. Paranchych, W., and L. S. Frost. 1988. The physiology and biochemistry of pili. *Adv. Microb. Physiol.* **29**:53–114.
  25. Reznikoff, W. S., D. A. Siegle, D. W. Cowing, and C. A. Gross. 1985. The regulation of transcription initiation in bacteria. *Annu. Rev. Genet.* **19**:355–387.
  26. Sambrook, J., E. F. Fritsch, and T. Maniatis. 1989. *Molecular cloning: a laboratory manual*, 2nd ed. Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.
  27. Sawers, G., and A. Böck. 1989. Novel transcriptional control of the pyruvate formate-lyase gene: upstream regulatory sequences and multiple promoters regulate anaerobic expression. *J. Bacteriol.* **171**:2485–2498.
  28. von Heijne, G. 1990. The signal peptide. *J. Membr. Biol.* **115**:195–201.
  29. Yanisch-Perron, C., J. Vieira, and J. Messing. 1985. Improved M13 phage cloning vectors and host strains: nucleotide sequences of the M13mp18 and pUC19 vectors. *Gene* **33**:103–119.