

Ammonification in *Bacillus subtilis* Utilizing Dissimilatory Nitrite Reductase Is Dependent on *resDE*

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During anaerobic nitrate respiration *Bacillus subtilis* reduces nitrate via nitrite to ammonia. No denitrification products were observed. *B. subtilis* wild-type cells and a nitrate reductase mutant grew anaerobically with nitrite as an electron acceptor. Oxygen-sensitive dissimilatory nitrite reductase activity was demonstrated in cell extracts prepared from both strains with benzyl viologen as an electron donor and nitrite as an electron acceptor. The anaerobic expression of the discovered nitrite reductase activity was dependent on the regulatory system encoded by *resDE*. Mutation of the gene encoding the regulatory Fnr had no negative effect on dissimilatory nitrite reductase formation.

Changes in oxygen tension are a part of the environmental conditions in the habitat of the soil bacterium *Bacillus subtilis*. Oxygen is the major electron acceptor for aerobic growth. In the absence of oxygen *B. subtilis* develops alternative strategies for survival. Two general principles of anaerobic catabolism are known. Reducing equivalents (NADH and reduced flavin adenine dinucleotide) produced mainly by glycolysis and the Krebs cycle during the consumption of energy substrates like glucose are oxidized by membrane-localized electron transport chains. During these respiratory processes a transmembrane proton gradient is generated and used as the energy supply for ATP synthesis and transport of molecules through the cytoplasmic membrane. In the absence of external electron acceptors, the reducing equivalents are reoxidized by endogenous electron acceptors without proton gradient formation. This redox-balanced dismutation of the substrate is coupled directly on the substrate level to energy conservation.

The energetic yields of these fermentation processes are much lower than those of anaerobic respiration. However, anaerobic respiratory electron transport requires the presence and efficient utilization of alternative terminal electron acceptors (1). Recently, the utilization of nitrate as an alternative anaerobic electron acceptor was described for *B. subtilis* and genes for the respiratory nitrate reductase system (*narGHJ*) were cloned (2–4). A large variety of bacilli have been identified as respiring nitrate (10, 14). For *Bacillus licheniformis*, denitrification from nitrate via nitrite, NO, and N₂O to N₂ was shown (9). *Bacillus macerans* performs ammonification with the production of ammonia from nitrate (10). For the gram-positive model bacterium *B. subtilis*, anaerobic nitrate utilization has been demonstrated without elucidation of the biochemical fate of the reduced nitrate. Induction of anaerobic nitrate respiration in *B. subtilis* is dependent on the regulator Fnr (2). The currently known location of potential Fnr binding sites and the functional analysis of other anaerobically induced genes (6) indicate the limitation of Fnr function in *B. subtilis* to the regulation of nitrate respiration. Moreover, in contrast to Fnr function in enterobacteria, the *B. subtilis fnr* gene is under

the control of a second regulatory component, the multifunctional ResDE system (6, 8, 13). Besides its recently discovered importance for anaerobic metabolism, the *resDE* locus is involved in the regulation of various other central cellular functions, such as sporulation, carbon source utilization, the phosphate regulon, heme A synthesis, and formation of the cytochrome *bf* complex (5, 13).

With this study we identified *B. subtilis* as an ammonifying bacterium converting nitrate via nitrite into ammonia. A dissimilatory nitrite reductase activity which required *resDE* but not *fnr* for its formation under anaerobic growth conditions was identified and measured.

Anaerobic conversion of nitrate via nitrite into ammonia by *B. subtilis*. *B. subtilis* JH642 (*pheA1 trpC2*) (Bacillus Genetic Stock Center) and THB1 (*pheA1 trpC2 ΔnarGH::Tet Tet^r*) (4) were grown anaerobically at 37°C on Luria-Bertani medium (7) supplemented with 20 mM K₃PO₄ (pH 7), 2 mM (NH₄)₂SO₄, 1 mM L-glutamic acid, 1 mM L-tryptophan, 0.8 mM L-phenylalanine, 0.005% (wt/vol) ammonium iron(III) citrate, 1 mM glucose, and, when indicated, 10 mM nitrite or nitrate (4, 6). The bacteria were incubated in completely filled flasks with rubber stoppers and with shaking at 180 rpm in an incubation shaker to minimize aggregation of the bacteria. Inoculation was performed aerobically at a 1:100 ratio of aerobically grown overnight culture and prewarmed medium. Anaerobic conditions were achieved after a short time through consumption of residual oxygen by the inoculated bacteria (11). Nitrate and ammonia concentrations in the growth media were monitored with standard enzymatic test systems (Boehringer, Mannheim, Germany). The ammonia concentration of the employed medium was subtracted from the measured values. Nitrite concentrations were measured by the diazotation procedure (12).

Under anaerobic growth conditions *B. subtilis* JH642 almost completely converted nitrate into nitrite over a period of 6 h until the end of the log phase (Fig. 1A and 2A). Immediately upon the appearance of nitrite as the product of nitrate reduction, its further conversion into ammonia started (Fig. 2A). Under the employed conditions, approximately 65% of the produced nitrite was converted into ammonia (Fig. 2A). Almost complete conversion of nitrite into ammonia was observed only for the nitrate reductase mutant THB1 (Fig. 2B) and for wild-type cells (data not shown) when nitrite was sup-

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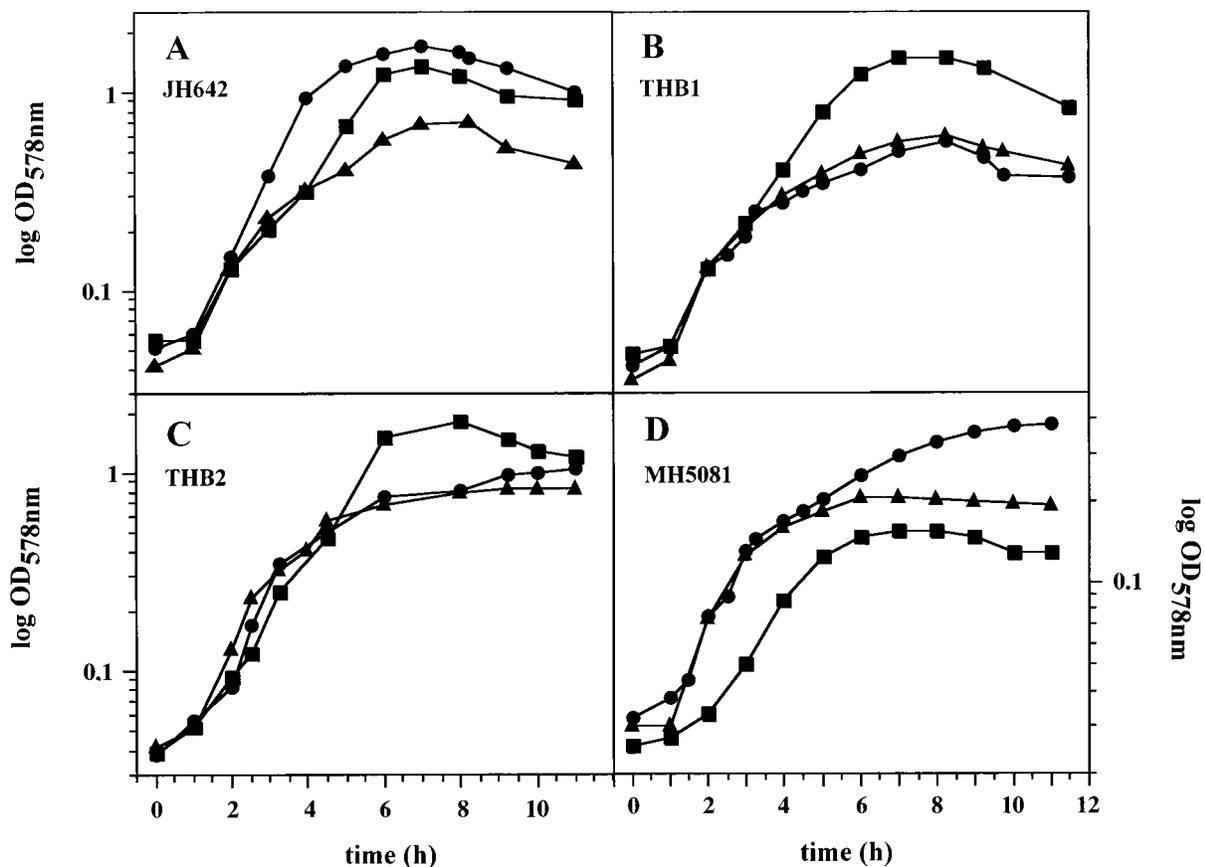


FIG. 1. Anaerobic growth of *B. subtilis* JH642 (wild type) (A), THB1 (nitrate reductase-deficient mutant) (B), THB2 (*fur* mutant) (C), and MH5081 (*resDE* mutant) (D) with 10 mM nitrate (●) or 10 mM nitrite (■) as a terminal electron acceptor and fermentation in the absence of external electron acceptors (▲). Growth was monitored by determination of the optical density at 578 nm (OD_{578nm}) at the indicated time points. Values reported are the averages from at least 10 independent experiments performed in triplicate.

plied as the electron acceptor. The gas phase of the *B. subtilis* growth chamber was carefully analyzed for gases resulting from potential denitrification processes. Growth experiments with denitrifying *Pseudomonas aeruginosa* (PAO1) and ammonifying *Escherichia coli* (K-12) served as controls. No significant amounts of N_2O or N_2 were detected in the gas phase of the *B. subtilis* and *E. coli* growth chambers, while the gas phase of the *P. aeruginosa* growth chamber contained substantial amounts of the various denitrification gases (data not shown). Clearly, *B. subtilis* should be regarded as an ammonifying facultative anaerobe.

In the absence of external terminal electron acceptors, fermentative growth of *B. subtilis* JH642 up to an optical density at 578 nm of 0.7 was observed (Fig. 1). The exact nature of the observed fermentation processes remains to be determined in future experiments.

***B. subtilis* contains a dissimilatory nitrite reductase.** To determine whether the conversion of nitrite into ammonia sustains anaerobic growth, *B. subtilis* JH642 was incubated anaerobically on medium containing nitrite. *B. subtilis* efficiently grew anaerobically with nitrite as an electron acceptor (Fig. 1A). To exclude any residual or side activity of the nitrate reducing system, the dissimilatory-nitrate-reductase-deficient mutant THB1 was incubated anaerobically with nitrite. As shown in Fig. 1B, THB1 grew efficiently with nitrite as an electron acceptor, demonstrating the presence of a dissimilatory nitrite reductase in *B. subtilis*. Almost all supplied nitrite

was converted into ammonia by THB1 (Fig. 2B). In control experiments, the mutant THB1 failed to grow efficiently on nitrate as an electron acceptor (Fig. 1B) and consequently no nitrate conversion was detected (Fig. 2B). Observed residual growth in the absence of external terminal electron acceptors was on the level of fermentative growth (Fig. 1B).

An oxygen-sensitive benzyl viologen-dependent nitrite reductase activity is present in cell extracts prepared from *B. subtilis*. To measure the newly discovered enzymatic activity directly in *B. subtilis* cell extracts, an in vitro test system had to be established. For this purpose, *B. subtilis* cultures grown anaerobically in the presence of 10 mM nitrate and nitrite were used at the end of the exponential growth phase. Harvesting, washing, and disruption of cells were performed under strictly anaerobic conditions. Cells were harvested and washed with 10 mM Tris (pH 7.4) containing 1 mg of $MgCl_2 \cdot 6H_2O$ per ml. Cells were anaerobically disrupted with a French press (20,000 lb/in²). To obtain cell extracts, the suspensions were filtered through a 0.2- μ m-pore-size sterile filter. Dissimilatory nitrite reductase activity was measured spectroscopically by the nitrite-dependent oxidation of reduced benzyl viologen in anoxic cuvettes containing 50 mM Tris (pH 7.4), 2 mM benzyl viologen reduced with 0.2 μ M sodium dithionite, 1 mM KNO_2 , and 5 to 50 μ l of cell extract (containing 2 mg of protein per ml) at room temperature. Reaction mixtures were incubated in the absence of external electron acceptors to determine the endogenous rate of benzyl viologen oxidation. No obvious back-

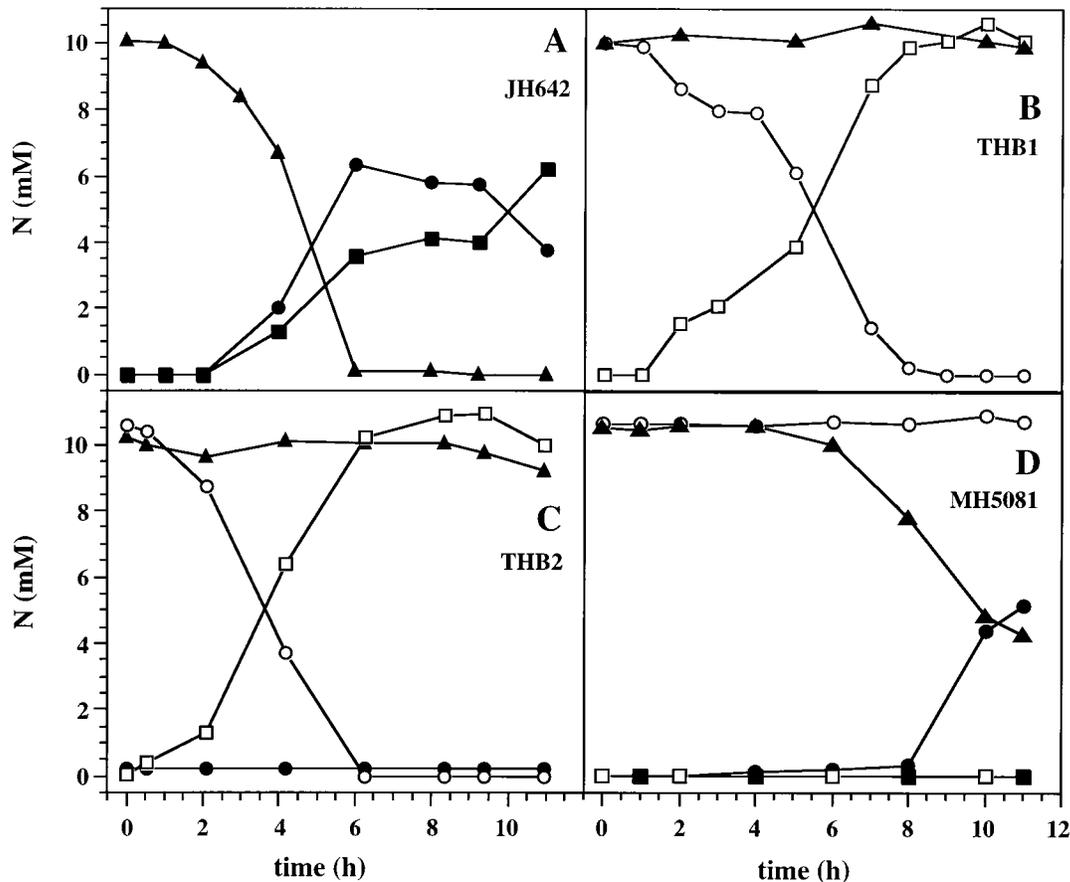


FIG. 2. Changing concentrations of nitrate (▲), nitrite (● and ○), and ammonia (■ and □) during anaerobic growth of *B. subtilis* JH642 (wild type) (A), THB1 (nitrate reductase-deficient mutant) (B), THB2 (*fnr* mutant) (C), and MH5081 (*resDE* mutant) (D). The concentrations given were determined at the indicated time points in the media of *B. subtilis* cultures grown with 10 mM nitrate (closed symbols) or 10 mM nitrite (opened symbols). Values reported are the averages from at least 10 independent experiments performed in triplicate.

ground activities were observed. Reactions were started by the addition of nitrite. Enzymatic oxidation of benzyl viologen was monitored spectroscopically at a wavelength of 578 nm. The extinction coefficient of benzyl viologen was used to determine the amount of reduced nitrite. One unit of activity corresponded to the reduction of 1 μmol of nitrite per min. Important for successful enzyme activity measurements was the strictly anaerobic handling during extract preparation. The nitrite reductase activity of approximately 5.6 U/mg of protein in cell extracts prepared from *B. subtilis* THB1 (Table 1) was reduced to approximately 2.9 U/mg of protein after 15 min of aerobic treatment and further to approximately 0.7 U/mg of protein after 90 min of exposure to aerobic conditions.

For all analyzed *B. subtilis* strains respiratory nitrate reductase activities were also measured. Since this enzyme activity was much less oxygen sensitive, cell extracts were prepared aerobically. However, no obvious benzyl viologen-dependent nitrite reductase activity was measured under these conditions. Strictly anaerobic handling of the protein fractions did not significantly increase nitrate reductase activities (data not shown). Cells were aerobically harvested, washed, disrupted with a French press (20,000 lb/in²), and subsequently centrifuged to give cell extracts. Respiratory nitrate reductase activity was measured spectroscopically by the nitrate-dependent oxidation of reduced benzyl viologen in anoxic cuvettes containing 50 mM Tris (pH 7.4), 2 mM benzyl viologen reduced

with 0.2 μM sodium dithionite, 1 mM KNO₃, and 100 to 200 μl of cell extract (containing 2 mg of protein per ml) at room temperature (10). Reactions were started by the addition of nitrate. Enzymatic oxidation of benzyl viologen was monitored spectroscopically at a wavelength of 578 nm. The extinction coefficient of benzyl viologen was used to determine the amount of reduced nitrate. One unit of activity corresponded to the reduction of 1 μmol of nitrate per min.

TABLE 1. Enzymatic determination of benzyl viologen-dependent nitrate and nitrite reductase activities

<i>B. subtilis</i> strain	Avg activity \pm SD (10^{-3} U/mg of protein) ^a	
	Nitrate reductase	Nitrite reductase
JH642 (wild type)	330 \pm 10	6,100 \pm 220
THB1 (<i>narG</i> mutant)	ND	5,600 \pm 280
THB2 (<i>fnr</i> mutant)	ND	9,900 \pm 310
MH5081 (<i>resDE</i> mutant)	50 \pm 10	ND

^a Benzyl viologen-dependent dissimilatory activities determined in vitro with cell extracts prepared from the *B. subtilis* strains after anaerobic growth on nitrate- and nitrite-containing medium. One unit of activity corresponded to the reduction of 1 μmol of nitrite or nitrate per min. See the text for detailed assay conditions. Values reported are the averages from at least three independent experiments performed in triplicate. ND, not detectable.

As shown in Table 1, wild-type *B. subtilis* (JH642) contained both dissimilatory nitrite and nitrate reductase activities. Control reactions with boiled enzyme fraction and without the addition of nitrite or nitrate clearly demonstrated the enzymatic nature of the observed reaction and its electron acceptor dependence (data not shown). The electron acceptor specificity of the used nitrite reductase assay was tested in a nitrate reductase-free background with extracts prepared from the nitrate reductase-deficient mutant THB1. THB1 contained only nitrite reductase activity (Table 1). These results demonstrated the presence of a dissimilatory nitrite reductase activity in *B. subtilis*.

The regulatory system *resDE* but not *fnr* is required for induction of dissimilatory nitrite reductase activity. The two-component system encoded by *resDE* is responsible for induction of respiratory nitrate reductase partially via transcriptional activation of *fnr* (8, 13). To determine the influence of *resDE* and *fnr* on dissimilatory nitrite reductase formation, the *resDE* mutant MH5081 (*pheA1 trpC2 resDE::pRC22 Tet^r*) (13) and the *fnr* mutant THB2 (*pheA1 trpC2 fnr::spec Spec^r*) were incubated anaerobically in the presence of nitrite.

The overall anaerobic growth of the *resDE* mutant in the absence of external electron acceptors was already reduced compared to that of wild-type *B. subtilis*, indicating a potential role of *resDE* in the regulation of fermentative processes (Fig. 1D). Moreover, the pleiotropic phenotype of the *resDE* mutation might contribute to the observed reduced growth. No increase of anaerobic growth with nitrite as the electron acceptor was detected, while a basal level even below normal fermentative growth remained (Fig. 1D). Moreover, the nitrite concentration in the growth media remained unchanged over the whole incubation period of 13 h (Fig. 2D). In vitro dissimilatory nitrite reductase activity tests (Table 1) using cell extract prepared from the *resDE* mutant failed to detect significant enzyme activity. These results demonstrated the dependence of nitrite reductase activity formation on *resDE*.

In control experiments the mutant MH5081 was incubated with nitrate (Fig. 1D and 2D). From the previously published experiments concerning *resDE*-dependent *narGHII* expression, a result similar to that of the growth experiment using nitrite was expected (8, 13). Interestingly, the *resDE* mutant MH5081 started to grow on nitrate after 6 h and at the same time conversion of nitrate into nitrite and in vitro nitrate reductase activity were observed (Fig. 1D and 2D; Table 1). In agreement with results of nitrite growth experiments described above, no further conversion of the produced nitrite into ammonia was observed, since the respiratory nitrite reductase was still not activated. Since the previous investigations concerning the *resDE* influence on respiratory nitrate reductase formation were ended 5 h after inoculation, the authors might have missed this late low-level expression of the enzyme (8).

The *fnr* mutant of *B. subtilis* JH642, THB2, was constructed as described before (2). Anaerobic growth of the *fnr* mutant with nitrite as an electron acceptor was slightly better than growth of the wild-type strain (Fig. 1C). Consequently, efficient nitrite conversion by the *fnr* mutant (Fig. 2C) and an increased

respiratory nitrite reductase activity in cell extracts prepared from the *fnr* mutant were observed (Table 1). In agreement with results of previous investigations (2), *fnr* mutation totally abolished respiratory growth using nitrate as an electron acceptor (Fig. 1C). No nitrate reductase activity was measured in cell extracts of the *fnr* mutant (Table 1). These experiments exclude a stimulatory role of *fnr* on dissimilatory nitrite reductase expression as observed for the *narGHII* genes encoding nitrate reductase.

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