

Characterization of the HspR-Mediated Stress Response in *Helicobacter pylori*

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Received 17 December 2001/Accepted 12 March 2002

The major heat shock genes of *Helicobacter pylori* are regulated by the HspR repressor. In the present study we characterize the transcriptional response of the three known HspR-dependent promoters P_{cbp} , P_{gro} , and P_{hrc} to different environmental stresses. A temperature shift from 37 to 42°C causes a typical heat shock response at all three promoters characterized by an immediate and strong induction phase of transcription and a subsequent adaptation phase, which is specific for each promoter and whose onset is determined partially by the half-lives of the respective mRNAs. Exposure to high osmolarity induces a similar response on the P_{gro} and P_{cbp} promoters while no such response is detectable at the P_{hrc} promoter. Puromycin treatment induces transcription from all three HspR-dependent promoters, indicating that different environmental stresses are intracellularly sensed by the regulatory machinery through the accumulation of nonnative proteins. The implications of these data for the regulatory network controlling the heat shock response in *H. pylori* are discussed.

The heat shock proteins of the gastric pathogen *Helicobacter pylori* have been studied in some detail both because of their general role in the protection of the bacteria from the hostile environment represented by the human stomach and because of their involvement in specific pathogenic processes. The *H. pylori* GroEL (Hsp60) homologue (12, 20) has been proposed to play a role in regulating the activity of the nickel-dependent urease enzyme, which generates ammonia ions from the hydrolysis of urea and therefore protects the bacteria from the low pH of the stomach lumen (4, 6). Its cochaperone GroES (Hsp10) is thought to contribute to this regulation by controlling the availability of nickel ions by means of its intrinsic metal-binding activity (10, 20). GroEL as well as another major heat shock protein, DnaK (Hsp70), can furthermore be found in association with the outer membrane, and this surface localization has been suggested to modify the glycolipid binding specificity of *H. pylori* cells at a low pH (5, 9, 15).

Because of their multiple functions in both the general stress response and in specific disease mechanisms, the *H. pylori* heat shock proteins are expected to be tightly regulated in their expression levels. It was recently demonstrated that transcription of the *groESL*, *hrcA-grpE-dnaK*, and *cbpA-hspR-orf* operons encoding the major chaperones of *H. pylori* is negatively regulated by HspR (17). HspR is a homologue of the repressor of the *dnaK* operon of *Streptomyces coelicolor* (2, 7). By RNA analyses and footprint experiments with purified protein preparations, we have shown that HspR represses transcription by binding to regions of 75 bp on the three chaperone gene-transcribing promoters P_{gro} , P_{hrc} , and P_{cbp} . In an attempt to

identify the environmental stresses, which induce transcription from these promoters, we subjected *H. pylori* cultures to both osmotic shock and thermal shock at 45°C. While the P_{gro} and P_{cbp} promoters were found to be strongly inducible by treatment with 300 mM NaCl, no induction could be observed on either promoter by incubation of cultures at 45°C, indicating that heat shock does not induce transcription from HspR-dependent promoters. Recently, however, Homuth and co-workers (8) have shown that the P_{gro} and P_{hrc} promoters are strongly inducible by a mild heat shock at 42°C, suggesting that HspR can indeed mediate the transcriptional response to a sudden temperature increase. In the present work we determine in detail the transcriptional response of the three HspR-dependent promoters to different environmental stresses. We show that a temperature shift to 42°C causes a typical heat shock response at all three promoters, characterized by a fast and strong induction of transcription and a subsequent shutoff phase, whose onset is determined largely by the stability of the respective mRNAs. Treatment with 300 mM NaCl causes a similar response on both P_{gro} and P_{cbp} while transcription from P_{hrc} is essentially unaffected under these conditions. Puromycin treatment also induced transcription from the HspR-regulated promoters, indicating that misfolded proteins represent the intracellular signal that is sensed by the transcriptional machinery.

MATERIALS AND METHODS

Media and growth conditions. *H. pylori* G27 (24) cells were recovered from frozen stocks on Columbia agar plates containing 5% horse blood, 0.2% cyclodextrin, and Dent's or Skirrow's antibiotic supplement under microaerophilic conditions (Oxoid) for 2 to 3 days. After passage on fresh plates, bacteria were cultured in a 5% CO₂-95% air atmosphere. Liquid cultures of *H. pylori* were grown in modified brucella broth containing Dent's or Skirrow's antibiotic supplement and 5% fetal calf serum in a 5% CO₂-95% air atmosphere. When required, rifampin or puromycin was added to a final concentration of 80 or 20 µg/ml, respectively. Osmotic shock of *H. pylori* cultures was performed by adding NaCl to a final concentration of 300 mM NaCl. Heat shock was achieved by

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incubation of *H. pylori* cultures in a water bath at 42°C. It should be noted that previous heat shock experiments in our laboratory were performed by incubation of liquid cultures in microaerophilic jars (17). Differences in the induction profiles of heat shock promoters with respect to previously published experiments might therefore be attributable to these different growth conditions.

RNA preparation. *H. pylori* G27 was grown in 260 ml of modified brucella broth at 37°C to mid-log phase and then exposed to various stresses. At different time intervals after stress exposure, 25-ml samples were harvested and stored at -20°C. Cells were lysed in 3.7 ml of 100 mM Tris-HCl (pH 7.5), 2 mM Na₂-EDTA, and 1% sodium dodecyl sulfate for 5 min at 95°C. After 10 min of incubation on ice in the presence of 80 mM KCl, cellular debris was removed by centrifugation at 8,000 rpm for 10 min in a JA20 rotor (Beckman). To 3.5 ml of supernatant, 4.56 g of CsCl was added, and the RNA was sedimented by centrifugation in an SW65 rotor (Beckman) for 15 to 20 h at 35,000 rpm. The RNA pellet was then resuspended in 500 μl of TE (10 mM Tris-HCl [pH 8], 1 mM Na₂-EDTA), extracted once with an equal volume of phenol-chloroform (1:1), ethanol precipitated, resuspended in 200 μl of TE, reprecipitated, and stored at -20°C.

Quantification of transcripts. P_{gro}- and P_{hrc}-specific transcripts were detected by reverse transcription with oligonucleotides groS (5'-GACCCCTTCTCCTA ATGGCTG) and hrcA (5'-CAAACGCATCTAACAACTCTC) as described by Spohn and Scarlato (17). P_{cbp}-specific transcripts were detected by S1 nuclease mapping with a 674-bp-specific DNA probe (17). This DNA fragment was cloned into the pGem3 vector (Promega) as a PCR product with oligonucleotide pair cbp1-cbp2 (5'-attattggatccACCCAAGACGCGCTAAAGCCC and 5'-attattgAATTCTTGGGGTTAGGGGGATTTAAGGG, capital letters indicate *H. pylori* sequences and underlined sequences represent the *Bam*HI and *Eco*RI recognition sites, respectively) and comprises the 5' regions of the *cbpA* gene and the upstream open reading frame HP1023. Briefly, the DNA fragment was 5' end labeled at its *Eco*RI site in the presence of [³²P]ATP (5,000 Ci/mmol; Amersham) and T4 polynucleotide kinase (New England Biolabs). Approximately 20 fmol of the labeled fragment was coprecipitated with 15 μg of total *H. pylori* RNA and resuspended in 20 μl of hybridization buffer (80% formamide, 60 mM Tris-Cl [pH 7.5], 400 mM NaCl, 0.4 mM EDTA). The mixture was denatured at 100°C for 3 min and then incubated at the annealing temperature of 43°C. After 16 h of incubation, 180 μl of ice-cold S1 buffer (33 mM Na-acetate [pH 5.2], 5 mM ZnSO₄, 250 mM NaCl) and 1 μl of S1 nuclease (400 U/μl) were added and S1 digestion was carried out for 30 min at 37°C. Samples were then extracted once with phenol-chloroform, ethanol precipitated, resuspended in 5 μl of sequencing loading buffer, heat denatured, subjected to 6% urea polyacrylamide gel electrophoresis, autoradiographed, and exposed to a PhosphorImager (Molecular Dynamics) for radioactivity quantification. As a control experiment, the *flaB* transcription was assayed by primer extension with oligonucleotide *flaB* as described in Spohn and Scarlato (18).

RESULTS

Three HspR-regulated promoters exhibit a typical heat shock response at 42°C. The structural organization of the three HspR-regulated operons and the respective promoters are schematically depicted in Fig. 1. P_{gro} transcribes one bicistronic mRNA encoding the chaperonin machinery GroESL; P_{hrc} transcribes a tricistronic mRNA encoding the putative repressor HrcA, the chaperone DnaK, and its cochaperone GrpE (8, 17); and P_{cbp} is likely to transcribe a tricistronic mRNA encoding the DnaJ homologue CbpA, the HspR repressor, and a putative DNA helicase (17). To investigate the transcriptional response of these promoters to a sudden temperature increase, total RNA was isolated from *H. pylori* strain G27 grown at 37°C and at different time points after upshift of the culture to 42°C. Promoter-specific mRNA was then detected by reverse transcription with specific oligonucleotides or by S1 nuclease mapping with specific DNA probes, respectively, and quantified by exposure of urea-acrylamide gels to a PhosphorImager. The results of this analysis are shown in Fig. 2. Figure 2A shows that the amount of each transcript increases rapidly after temperature upshift, starting at 2 min and reaching a maximum at 10 to 15 min. As evident from Fig. 2B,

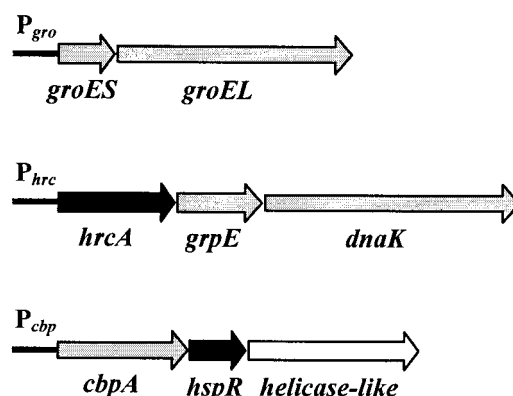


FIG. 1. Structural organization of *H. pylori* chaperone genes (17, 21). Grey arrow bars indicate chaperone genes, black arrow bars indicate regulatory genes (*hspR* and *hrcA*), and the white arrow bar indicates a putative helicase-like gene. Chaperone genes *groES* and *groEL* code for the HspA (Hsp10) and HspB (Hsp60) proteins (20); *cbpA* encodes a protein with 30% amino acid identity (in a 288-amino-acid overlap) to the chaperone curved DNA binding protein CbpA from *E. coli*, a homologue of DnaJ (22); *hspR* encodes a regulatory protein with 46% identity (in a 91-amino-acid overlap) to HspR, the negative regulator of heat shock genes in *S. coelicolor* (2); the helicase-like open reading frame encodes a protein with 30% identity within 421 amino acids to a hypothetical helicase-like protein from *Haemophilus influenzae*; *hrcA* encodes a protein with 28% identity (in a 71-amino-acid overlap) to the heat-inducible transcriptional repressor HrcA from *B. subtilis* (23); and *grpE* and *dnaK* encode the GrpE and DnaK (Hsp70) chaperones, respectively.

the ratio of this increase of transcription is different for the three promoters, ranging from 5-fold for P_{gro} to 7-fold for P_{cbp} and 11-fold for P_{hrc}. The phase of maximal induction is followed by a decline phase during which transcript amounts gradually decrease. In the case of P_{cbp}, this shutoff phase starts immediately after the peak of induction at about 10 min and brings back transcript amounts to about twice the pre-heat shock levels within 1 h and to a level comparable to the unstressed situation after 3 h. P_{hrc} transcript amounts accumulate at the maximal level for about 10 min and then begin to decline, reaching a new steady-state level of transcription within 90 min, which is about fourfold higher than the uninduced level. Conversely, P_{gro} transcript levels do not decrease significantly after the initial induction phase and remain at a high level until the end of the time course at 3 h after temperature upshift.

We conclude that the three HspR-regulated promoters of *H. pylori* exhibit a typical heat shock response at 42°C. All three promoters are strongly and rapidly induced after temperature upshift. The adaptation phase following this initial induction phase follows different kinetics for the different promoters.

Differences in the adaptation kinetics are largely due to differences in mRNA stabilities. To establish whether the kinetic differences observed in the shutoff phase of transcription are consequences of a possible differential regulation of the three heat shock promoters by the HspR repressor or of regulatory events at the posttranscriptional level, we decided to estimate the half-lives of the respective mRNAs. For that purpose, *H. pylori* G27 cells were first exposed to a 15-min heat

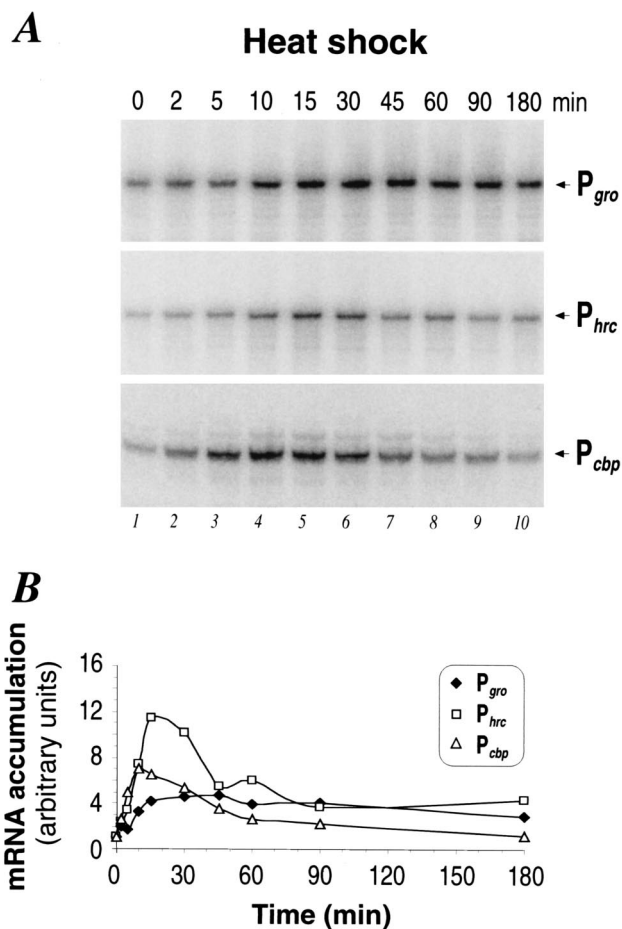


FIG. 2. Heat shock response of chaperone genes. (A) Primer extension (P_{gro} , P_{hrc}) and S1 nuclease (P_{cbp}) analyses of *H. pylori* RNA extracted from cells grown at 37°C (0 min) (lane 1) or upon temperature shift to 42°C (lanes 2 to 10). The time interval at which RNA was extracted is indicated, in minutes, above each lane. The products of RNA elongation or protection by S1 nuclease are marked on the rightmost side of the autoradiography by the names of the corresponding promoters: P_{gro} , P_{hrc} , and P_{cbp} . (B) Pattern of RNA accumulation at the indicated promoters as obtained by PhosphorImager analysis quantifications of the radioactive bands shown in panel A. It is worth mentioning that primer extensions conducted on the *cbpA* mRNA gave unsatisfactory results. By contrast, S1 nuclease mapping turned out to function very well in mapping both the transcription start site and the regulation of transcription upon stress conditions.

shock at 42°C to induce transcription and then treated with rifampin to stop transcriptional initiation. RNA was prepared before and at different time intervals after the addition of rifampin, and the amount of each specific transcript was monitored by reverse transcription and S1 nuclease mapping. Figure 3 shows that the amount of P_{cbp} -specific transcript decreases rapidly after rifampin treatment, with a half-life which calculates to ~6 min. P_{hrc} and P_{gro} transcripts on the other hand appear to be particularly stable; after 30 min of rifampin treatment, 65% of the initial amount of P_{hrc} -specific transcript and 85% of P_{gro} -specific transcript can still be detected. By extrapolation, the half-lives of the P_{hrc} and P_{gro} transcripts can therefore be estimated to 40 to 45 min and more than 90 min, respectively. These are unusually long half-lives. In fact, in

Bacillus subtilis the *groE* transcript shows an estimated half-life of 5 min under heat shock and non-heat shock conditions (26).

The differences in the adaptation kinetics between the three heat shock promoters can therefore probably be attributed to differences in the stabilities of the respective mRNAs. Transcriptional shutoff becomes evident immediately at the P_{cbp} promoter because of the short half-life of this mRNA, whereas the long half-lives of the P_{hrc} and P_{gro} mRNAs allow detection of shutoff only with a delay or not at all, respectively.

In order to investigate whether the initial induction of transcription detected after heat shock at 42°C is in part mediated by temperature-dependent changes in mRNA stability, we also measured the half-lives of the mRNAs transcribed from P_{gro} and P_{hrc} at 37°C. No significant differences could be observed in the stabilities of these mRNAs between heat shock and non-heat shock conditions (data not shown), indicating that the initial induction of transcription is solely the result of regulatory events at the level of transcription initiation.

The P_{cbp} and P_{gro} promoters are also induced by osmotic shock. Previous work had shown that salt treatment of *H. pylori* cells can induce transcription from the P_{cbp} and P_{gro} promoters but not from the P_{hrc} promoter (17). To study in more detail the transcriptional response of the three HspR-regulated promoters to a sudden increase in osmolarity, we isolated total RNA from *H. pylori* G27 immediately before and at different time intervals after the addition of 300 mM NaCl and assayed transcript levels as before by primer extension and S1 nuclease mapping and subsequent PhosphorImager analysis. Figure 4A shows that the P_{cbp} and P_{gro} promoters are induced rapidly after salt treatment, reaching a maximum after 10 and ~30 min, respectively. As shown in Fig. 4B, the induction ratios for both promoters are similar to those found for heat shock treatment, i.e., ~6- to 7-fold for P_{cbp} and 5- to 6-fold for P_{gro} . Also, the induction kinetics resemble those observed after thermal upshift, with P_{cbp} starting shutoff of transcription immediately

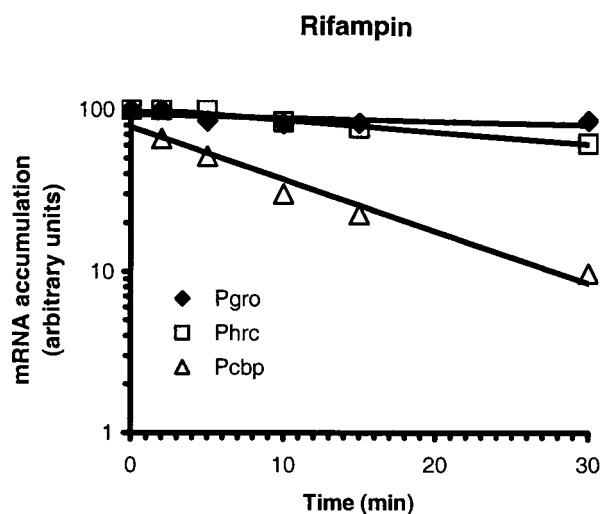


FIG. 3. Estimation of mRNA half-lives at 42°C. Data points represent PhosphorImager analysis quantifications of radioactive bands obtained by primer extension or protection by S1 nuclease of RNA extracted before (0 min) and after the addition of rifampin to growing cells at the indicated time intervals.

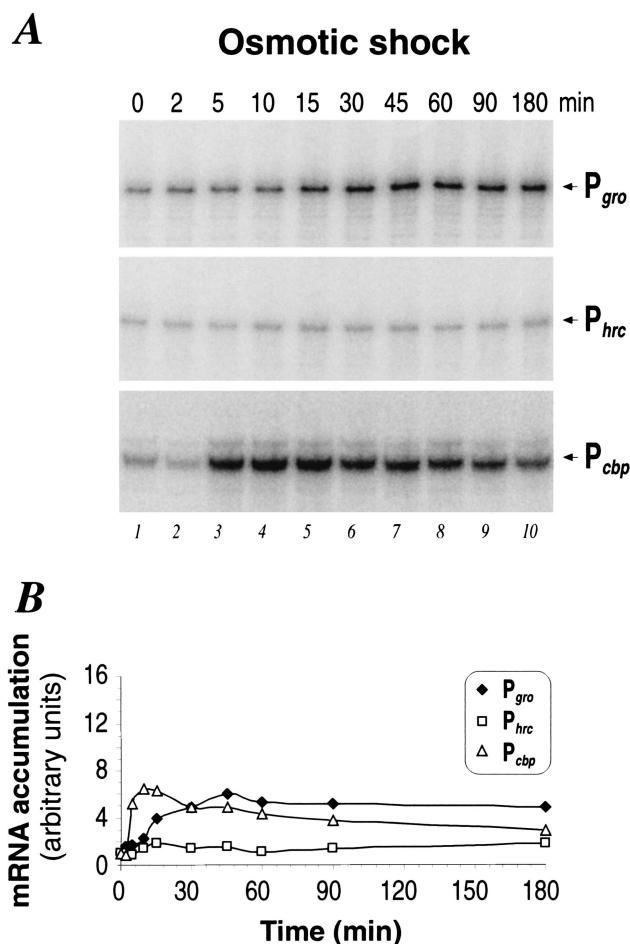


FIG. 4. Osmotic shock response of chaperone genes. (A) Primer extension (P_{gro} , P_{hrc}) and S1 nuclease (P_{cbp}) analyses of *H. pylori* RNA extracted from cells grown at 37°C (0 min) (lane 1) in normal medium or upon addition of 300 mM NaCl (lanes 2 to 10). Symbols are as described in the legend to Fig. 2. (B) Pattern of RNA accumulation at the indicated promoters as obtained by PhosphorImager analysis quantifications of the radioactive bands shown in panel A.

after the peak of induction and P_{gro} transcript levels remaining at the maximal induced level for up to 3 h after temperature upshift. In agreement with previously published results (17), no significant induction could be observed at the P_{hrc} promoter. Transcription of this promoter remains at the basal level during the whole time course of osmotic shock treatment. Similar results have been obtained by adding 440 mM sucrose to growing cells (data not shown).

Nonnative proteins induce an HspR-dependent stress response. The fact that environmental stresses as diverse as osmolarity changes and temperature increases can induce the same transcriptional response at two of the HspR-regulated promoters supports the hypothesis that these stresses are not sensed directly by the regulatory apparatus but converge into a common intracellular signal which is then perceived and transduced into a transcriptional response. In *Escherichia coli* and *B. subtilis* this intracellular signal is believed to be represented by unfolded or aberrant proteins that accumulate under various conditions of environmental stress, triggering in this way the onset of the heat shock response. To investigate if nonna-

tive proteins can induce a heat shock response of the HspR-dependent promoters, we isolated total RNA from *H. pylori* cells before and at different time intervals after the addition of puromycin, a tRNA analog that incorporates into nascent polypeptide chains and leads to premature translation termination. Transcription from the three HspR-dependent promoters was monitored as before by primer extension and S1 mapping and subsequent PhosphorImager analysis of urea-acrylamide gels. Figure 5A shows that all three promoters are induced rapidly after the addition of puromycin, with transcript levels reaching a maximum after about 30 min. As shown in Fig. 5B, P_{hrc} and P_{cbp} transcript amounts increase about 10- to 12-fold, whereas P_{gro} -specific transcripts increase only about 3-fold. In contrast to the heat and osmotic shock responses shown in Fig. 2 and 4, respectively, transcript levels fail to decline after this initial induction phase and remain at the maximal level for the whole time course of the experiment. As a control, the P_{112} promoter transcribing the *flaB* mRNA (18,

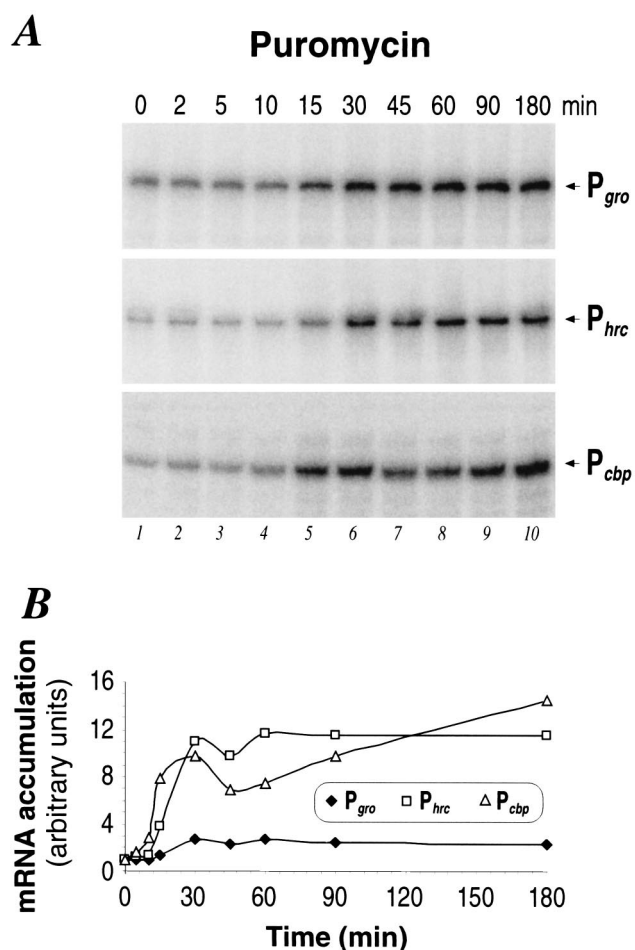


FIG. 5. Transcriptional response of chaperone genes to accumulation of misfolded polypeptides. (A) Primer extension (P_{gro} , P_{hrc}) and S1 nuclease (P_{cbp}) analyses of *H. pylori* RNA extracted from cells grown at 37°C (0 min) (lane 1) or upon the addition of puromycin (lanes 2 to 10). Symbols are as described in the legend to Fig. 2. (B) Pattern of RNA accumulation at the indicated promoters as obtained by PhosphorImager analysis quantifications of the radioactive bands shown in panel A.

19) was also included in this analysis, showing that transcription from this promoter was essentially unaffected by puromycin treatment (data not shown).

We conclude that aberrant proteins induced by the addition of puromycin can elicit a heat shock response of the HspR-regulated promoters of *H. pylori*.

DISCUSSION

Bacteria respond to conditions of stress by synthesizing chaperones that protect the cell from damage by preventing protein denaturation, aggregation, or misfolding (see reference 11 and references therein). The chaperones of the human gastric pathogen *H. pylori* have been studied in some detail because of their possible involvement in specific virulence mechanisms (9, 12, 20, 25). It has been shown that the *groESL*, *hrcA-grpE-dnaK*, and *cbpA-hspR-orf* operons, encoding the major chaperones of *H. pylori*, are transcribed by an RNA polymerase containing the vegetative sigma factor σ^{80} and regulated negatively by the transcriptional repressor HspR (17). Very recently, by using an *in vitro* selection and amplification approach, two new chromosomal binding sites of the HspR protein were identified (3). Surprisingly, these HspR binding sites are located at the 3' ends of two genes coding for predicted proteins with functions unrelated to those of chaperones. This suggests that *H. pylori* HspR may regulate the expression of genes encoding proteins with diverse functions. The control of transcription by HspR depends on the binding of this protein to large operators mapping to different positions surrounding the promoters (17). In addition to this regulation, the possibility that another transcriptional repressor, a homolog of the *B. subtilis* HrcA protein (16) encoded by the *hrcA-grpE-dnaK* locus, might also be involved in controlling stress response has been postulated (8, 17). Evidences that chaperone gene transcription in this system responds to an increase in salinity and temperature elevation to 42°C, but not to 45°C, have been recently reported (8, 17).

In this work we determined in detail the temporal pattern of transcriptional response for the three HspR-dependent promoters to different environmental stresses. A temperature shift to 42°C causes a typical heat shock response at the P_{gro} , P_{hrc} , and P_{cbp} promoters characterized by a fast and strong induction of transcription and a subsequent shutoff phase (Fig. 2). The initial induction is a true consequence of HspR-mediated derepression of transcription initiation, as stabilities of the different mRNAs were found to remain unchanged after temperature upshift with respect to the nonstressed situation. The absolute differences in mRNA stabilities between the three promoters appear, however, to influence the onset and temporal pattern of the adaptation phase of transcription: the P_{cbp} promoter, which transcribes the mRNA with the shortest half-life of the three heat shock promoters (6 min) (Fig. 3), shows the earliest onset of the shutoff phase of transcription at about 10 min after temperature upshift while the P_{hrc} and P_{gro} promoters, transcribing mRNAs with significantly longer half-lives (45 and >90 min, respectively), appear to enter this phase only at later stages or not at all, respectively. Despite these differences in mRNA stabilities, however, it is reasonable to assume that at the level of transcriptional initiation, HspR-dependent repression is restored on all three promoters shortly after the

initial induction of transcription. Likely, at this stage of the heat shock response, the HspR repressor begins to regain its active, repression-competent conformation, possibly aided by the action of one or more specialized molecular chaperones. In support of this hypothesis is the recent finding that the DnaK chaperone of *S. coelicolor* can act as a corepressor together with the HspR protein (1). In this model, a DnaK-HspR complex efficiently represses transcription under non-heat shock conditions while a sudden increase in temperature leads to the transient sequestration of DnaK by accumulating unfolded proteins, thereby freeing the HspR repressor, which thus becomes inefficient in DNA binding and transcriptional repression. The increased amounts of DnaK protein produced after this induction of transcription then lead to efficient refolding of denatured proteins and restoration of the HspR-DnaK complex, which reestablishes transcriptional repression. Although in *H. pylori* no indications for the existence of such a complex have been found, it is likely that DnaK or another molecular chaperone participates in regulation of the DNA binding activity of HspR. In support of this hypothesis is the fact that puromycin treatment of *H. pylori* cells leads to a strong and irreversible induction of transcription at all three HspR-regulated promoters (Fig. 5). This induction could be interpreted as a result of the permanent sequestration of DnaK by the puromycin-induced aberrant proteins, and the consequent irreversible loss of DNA binding activity by the HspR repressor. Under physiological conditions DnaK would therefore represent the intracellular stress sensor, which mediates the appropriate transcriptional response according to the level of misfolded proteins present in the cytoplasm. All extracellular stresses, which produce aberrant or misfolded proteins in the cytoplasm, should therefore elicit a heat shock response at the HspR-dependent promoters via titration of the DnaK protein. As shown in Fig. 4, environmental stresses different from temperature changes can indeed induce a transcriptional response at the HspR-regulated promoters. The P_{cbp} and P_{gro} promoters are strongly induced by the addition of 300 mM NaCl to the growth medium, and the kinetics of induction correspond to the ones observed under heat shock conditions.

In contrast to the P_{cbp} and P_{gro} promoters, transcription from the P_{hrc} promoter is insensitive to osmotic shock treatment, possibly indicating that this stimulus is not sufficient to destabilize the bound protein from this operator. In support of this hypothesis is the finding that HspR binds to the P_{hrc} -associated operator with two- and fourfold-higher affinities than the P_{cbp} - and P_{gro} -associated operators, respectively (17). Alternatively, a more complex mechanism of transcriptional control might be operating on the P_{hrc} promoter, possibly involving HrcA, the repressor encoded by the first gene of the *dnaK* operon. HspR and HrcA might act as corepressors on this promoter, and this dual control might lead to a differentiation of the transcriptional response according to the environmental stimulus perceived. The involvement of a second repressor, though, also implies a second negative modulator of the heat shock response. In *B. subtilis* it has been shown that the GroEL chaperonin functions as a negative regulator of heat shock gene expression (13, 14). In this system, GroEL is necessary for the HrcA repressor protein to function efficiently as a repressor of the *dnaK* operon by maintaining HrcA in a properly folded state. We speculate that an HrcA-unknown

chaperone and the HspR-DnaK systems in *H. pylori* are required to fine-tune chaperone gene transcription in response to a sudden change in the environmental growth condition.

ACKNOWLEDGMENTS

We thank C. Mallia for editing the manuscript and G. Corsi for artwork.

This work has been supported partially by EU-TMR grant FMRX-CT980164, Chiron, MURST, and University of Bologna. I.D. is the recipient of an EU-TMR fellowship (FMRX-CT980164).

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