

Yop Fusions to Tightly Folded Protein Domains and Their Effects on *Yersinia enterocolitica* Type III Secretion

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***Yersinia enterocolitica* organisms secrete Yop proteins via the type III pathway. Translational fusion of *yop* genes to ubiquitin or dihydrofolate reductase results in hybrid proteins that cannot be secreted. The folding of hybrids prevents their own transport, but it does not hinder the type III secretion of other Yops.**

The type III secretion machinery of pathogenic yersiniae (*Yersinia pestis*, *Y. pseudotuberculosis*, and *Y. enterocolitica*) provides for bacterial escape from phagocytic killing and is encoded by a 70-kb virulence plasmid (15). Twenty-one *ysc* (mnemonic for Yop secretion) genes encode secretion machinery components that promote transport of 14 Yops (mnemonic for *Yersinia* outer proteins) across the bacterial double-membrane envelope (15). The secretion signal of Yop proteins has been mapped by fusing the open reading frame of *yop* proteins to the 5' end of coding sequence for reporter proteins, thereby generating translational hybrids (38). Fusion of *yop* coding sequences to the 5' end of the adenylate cyclase-encoding domain of *cya* (23) results in the secretion of Yop-Cya fusions in a manner resembling that of the type III transport of native Yops (51). *lacZ* encodes β -galactosidase (29), a large (116-kDa) cytoplasmic polypeptide that assembles into a tetrameric structure (12). The type III secretion of *yop* fusions to full-length *lacZ* has not yet been examined. The first 92 codons of *lacZ* (*lacZ'*) specify the α -peptide, a module that unfolds readily but also provides for the alpha complementation phenotype of certain *lacZ* alleles (33). Fusion of *lacZ'* to the 3' end of *Yersinia* Yops results in the transport Yop-LacZ' hybrids into the extracellular medium (38). *npt* encodes neomycin phosphotransferase, a cytoplasmic protein that confers bacterial resistance to aminoglycoside antibiotics (47). Similarly to Cya and LacZ' fusions, Yop-Npt hybrids are also transported by the type III secretion machinery (3).

Fusion of the first 15 codons of *yopE*, *yopH*, *yopN*, or *yopQ* to the 5' end of *cya* or *npt* leads to the type III secretion of hybrid Yop proteins (1, 3, 5, 49, 50). Sory and Cornelis proposed that the amino acid sequence generated by the first 15 codons functions as a signal peptide and mediates substrate recognition by the type III machinery (51). Lloyd et al. developed this model further, predicting an amphipathic helical structure as a common substrate property of all Yop signal peptides (35, 36). *yop* secretion signals (codons 1 to 15) have been altered by frameshift mutations immediately following

the AUG start codon, while reporter expression was restored by suppressor mutations at the fusion site. Several of these frameshift mutations do not affect secretion signaling, albeit the peptide sequence is completely altered (1–3, 5). *yopQ* signal mutations with a defect in secretion in which the mRNA sequence is changed without affecting the codon specificity (synonymous mutations) have been described (44). Thus, it appears that *yop* mRNA may function as a signal for the type III secretion of Yop proteins (4). The two models vary considerably in predicting the mode of substrate recognition. Future work will need to provide more definitive proof to reveal the mechanisms of type III secretion.

Much of the thought on type III secretion is influenced by observations that were previously made with other protein secretion pathways (9). Signal peptides are hydrophobic sequences at the N terminus of precursor proteins that initiate polypeptides into the secretory pathway, a mechanism that translocates proteins across membranes (10). Signal peptides provide for substrate recognition by interacting with the SecYEG translocon in the plasma membranes of bacterial cells (7, 21). Once the precursor protein is translocated across the membrane, the signal peptide is removed by signal peptidase (leader peptidase) and the mature polypeptide is released from the secretion machinery (17). Transport by the Sec machinery occurs only when proteins assume an unfolded state and often requires the association of substrate with chaperones (32). Chaperones bind to the hydrophobic core residues of unfolded globular proteins (45) and are dissociated from the secretion substrate when a mobile machinery component, the ATPase SecA, moves precursor proteins across the plasma membrane (18, 24). A second transport pathway that involves the recognition of signal peptides in nascent polypeptides has been revealed (55). The signal recognition particle (SRP) binds to the signal peptide and the ribosome (42, 56). The complex between SRP, signal peptide of nascent polypeptide, and ribosome docks on the SRP receptor (34, 37). The ribosome is subsequently lodged onto the SecYEG secretion channel to promote cotranslational secretion of the remainder of the polypeptide chain (43, 57).

To identify bacterial machinery components that interact with signal peptides, previous work used several different strategies. Translational fusions between *lamB* and *malE*, encoding two secreted (signal peptide-bearing) proteins whose expres-

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TABLE 1. List of strains and plasmids used in this study

Strain or plasmid	Property	Reference or source
<i>Y. enterocolitica</i> W22703	Human clinical isolate, wild type	16
<i>Y. enterocolitica</i> LC2	Isogenic W22703 variant, $\Delta(\text{sycE})$	13
pDA14	<i>yopE</i> promoter and full-length ORF fused to 5' end of <i>lacZ</i>	This study
pDA36	<i>yopE</i> promoter and full-length ORF fused to 5' end of <i>npt</i>	3
pDA46	<i>yopE</i> promoter and codons 1 to 15 fused to 5' end of <i>npt</i>	3
pDA182	<i>yopQ</i> promoter and full-length ORF fused to 5' end of <i>npt</i>	5
pDA184	<i>yopQ</i> promoter and codons 1 to 15 fused to 5' end of <i>npt</i>	5
pVL9	<i>yopE</i> promoter and full-length ORF fused to 5' end of Ub gene- <i>npt</i>	This study
pVL13	<i>yopE</i> promoter and codons 1 to 15 fused to 5' end of Ub gene- <i>npt</i>	This study
pVL14	<i>yopE</i> promoter and full-length ORF fused to 5' end of Ub _{Gly3,13} gene- <i>npt</i>	This study
pVL15	<i>yopE</i> promoter and codons 1 to 15 fused to 5' end of Ub _{Gly3,13} gene- <i>npt</i>	This study
pVL16	<i>yopQ</i> promoter and full-length ORF fused to 5' end of Ub gene- <i>npt</i>	This study
pVL17	<i>yopQ</i> promoter and codons 1 to 15 fused to 5' end of Ub gene- <i>npt</i>	This study
pVL18	<i>yopQ</i> promoter and full-length ORF fused to 5' end of Ub _{Gly3,13} gene- <i>npt</i>	This study
pVL19	<i>yopQ</i> promoter and codons 1 to 15 fused to 5' end of Ub _{Gly3,13} gene- <i>npt</i>	This study
pVL20	<i>yopE</i> promoter and full-length ORF fused to 5' end of DHFR gene	This study
pVL21	<i>yopE</i> promoter and codons 1 to 15 fused to 5' end of DHFR gene	This study

sion is induced by maltose, and *lacZ*, specifying a cytoplasmic protein, cause the resulting hybrid proteins to jam the secretion pathway and to confer a maltose-sensitive growth phenotype (27, 41). Selection for maltose resistance led to the isolation of mutations that abolished the signal peptide function of *lamB* (20, 30). Mutations in secretion genes could be identified as conditional lethal mutations that confer increased LacZ activity due to the decreased initiation of the hybrids into the secretory pathway (8, 41, 48). The genetic strategies of identifying secretion machinery components are based on a single principle: signal peptide-bearing polypeptides are by default initiated into the secretion pathway and cannot be discarded unless the signal peptide is removed. Mutations in one secretion gene, *secB*, encoding the secretion chaperone, represent an important exception to this rule. *Escherichia coli* lacking the *secB* gene allows premature folding of the MalE precursor without initiating this polypeptide into the secretion pathway (46). This report examines whether the principle of a default mechanism for the initiation of polypeptides into the secretion machinery is applicable to the type III pathway of *Y. enterocolitica*.

Ubiquitin (Ub), a 76-residue polypeptide, folds rapidly into a compact, protease-resistant structure of five β -strands and four turns of α -helix (54). When fused to the C terminus of signal peptide-bearing precursor protein, the Sec pathway of *Saccharomyces cerevisiae* cannot translocate folded ubiquitin in a posttranslational manner across the endoplasmic reticulum (ER) membrane (28). Nevertheless, the SRP pathway of yeast does promote cotranslational transport of Ub if a sufficiently long spacer (more than 33 residues) separates the signal peptide and the Ub domain (28). Presumably, synthesis and folding of Ub fusion precursors in the cytoplasm are prevented by SRP-mediated arrest of translation. Once ribosomes are docked on the secretion channel in the ER membrane, cotranslational secretion proceeds at efficient speed and within space constraints that prevent Ub folding.

The Ub-coding sequence was cloned into the middle of reporter genes flanked by 5' *yop* and 3' *npt* sequences, inserted into the low-copy-number plasmid pHSG576 (53), and transformed into yersiniae (13) (Table 1 lists the properties of plasmids and strains used here). Overnight cultures of *Yersinia*

were diluted 1:50 into fresh tryptic soy broth, supplemented with 5 mM EGTA, grown for 2 h at 26°C, and induced at 37°C for 3 h. Cultures were centrifuged at 15,000 \times g for 15 min, and the supernatant (S) was separated from the cell pellet (P). Proteins in both fractions were precipitated with trichloroacetic acid (TCA), washed in acetone, and suspended in sample buffer. Proteins were analyzed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis and immunoblotting. Expression of *yopE*₁₋₂₂₀-Ub gene-*npt* in *Y. enterocolitica* led to the accumulation of the polypeptide in bacterial cells but did not result in type III secretion of YopE₁₋₂₂₀-Ub-Npt (full-length YopE fusion) (Fig. 1A and B). In contrast, YopE₁₋₂₂₀-Npt was secreted efficiently into the extracellular medium (Fig. 1B). A similar result was observed for YopQ-Ub fusions. YopQ₁₋₁₉₈-Npt (full-length YopQ fusion) was secreted via the type III pathway, whereas YopQ₁₋₁₉₈-Ub-Npt was not (Fig. 1C). To test our conjecture that the defect in type III secretion is caused by the folding of Ub domains, we analyzed Ub fusions carrying amino acid substitutions. Ub_{Gly3,13} cannot assume the tightly folded structure of native Ub and does not act as a substrate for Ub proteases (6). Because of their folding defects, Ub_{Gly3,13} fusions are translocated across the Sec channel in the yeast ER membrane in both co- and posttranslational manners (28). Expression of YopE₁₋₂₂₀-Ub_{Gly3,13}-Npt and YopQ₁₋₁₉₈-Ub_{Gly3,13}-Npt in *Y. enterocolitica* resulted in their secretion, indicating that it is indeed the tight folding of the Ub domain that prevents the type III transport of YopE₁₋₂₂₀-Ub-Npt and YopQ₁₋₁₉₈-Ub-Npt (Fig. 1).

YopE can be initiated into the type III pathway in one of two ways. The secretion signal in the first 15 codons promotes secretion of the polypeptide by the type III machinery (3). SycE is a small homodimeric polypeptide that binds to YopE amino acid residues 15 to 100 in the bacterial cytoplasm (13). It is not yet certain whether SycE binding functions as a secretion chaperone in maintaining protein substrates in an unfolded or export-competent conformation (13, 14, 58). Nevertheless, binding of SycE to amino acid residues 15 to 100 also allows the initiation of YopE transport, even in the absence of codons 1 to 15 (13). To test whether fusions of the *yop* secretion signal in the first 15 codons are capable of promoting type III secretion of Ub fusions, we expressed YopE₁₋₁₅-Ub-Npt

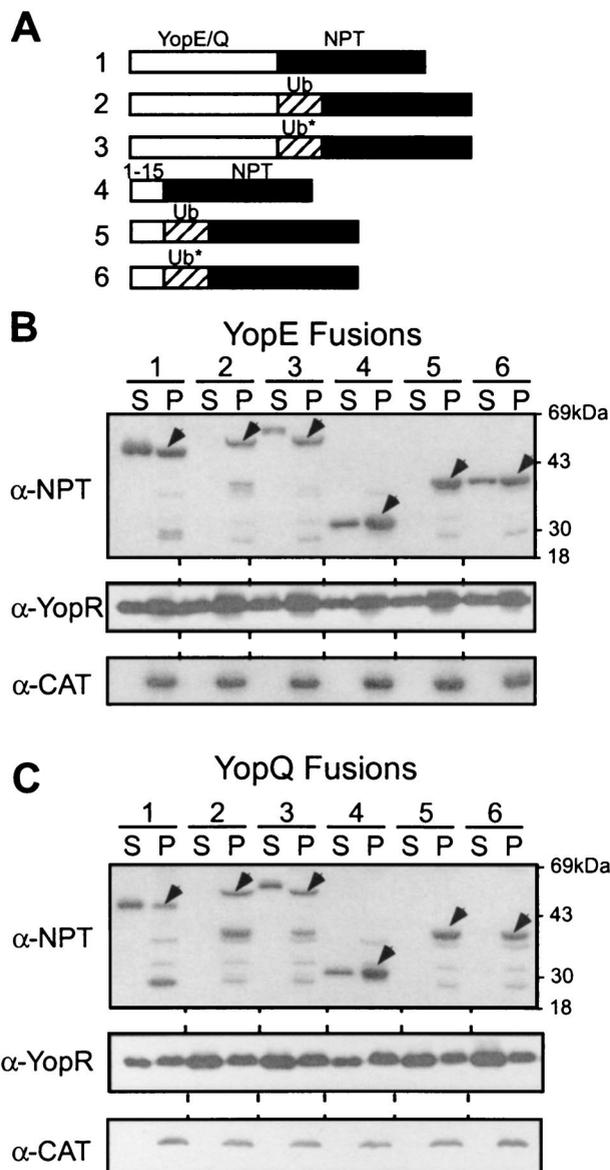


FIG. 1. Ubiquitin fusions are not secreted by the type III machinery of *Y. enterocolitica*. (A) The drawing depicts the primary structures of hybrid polypeptides as follows: line 1, YopE₁₋₂₂₀-Npt (YopQ₁₋₁₉₈-Npt), full-length *yop* fusion; line 2, YopE₁₋₂₂₀-Ub-Npt (YopQ₁₋₁₉₈-Ub-Npt), full-length *yop* ubiquitin fusion; line 3, YopE₁₋₂₂₀-Ub_{Gly3,13}-Npt (YopQ₁₋₁₉₈-Ub_{Gly3,13}-Npt), full-length *yop* fusion to ubiquitin mutant; line 4, YopE₁₋₁₅-Npt (YopQ₁₋₁₅-Npt), *yop* codons 1 to 15 fused; line 5, YopE₁₋₁₅-Ub-Npt (YopQ₁₋₁₅-Ub-Npt), *yop* codons 1 to 15 fused; line 6, YopE₁₋₁₅-Ub_{Gly3,13}-Npt (YopQ₁₋₁₅-Ub_{Gly3,13}-Npt), *yop* codons 1 to 15 fused. Ub represents wild-type ubiquitin, whereas Ub_{Gly3,13} carries two mutations that substitute codons 3 (isoleucine) and 13 (isoleucine) with glycine codons, causing destabilization of the folded polypeptide. Npt, neomycin phosphotransferase. (B) *Y. enterocolitica* strain W22703 (wild type) was transformed with plasmids listed in Table 1, grown in tryptic soy broth supplemented with 5 mM EGTA, and induced for type III secretion by a temperature shift to 37°C. Cultures were centrifuged, and the extracellular medium was separated with the supernatant (S) from the bacterial pellet (P). Proteins were precipitated with TCA, suspended in sample buffer, separated on sodium dodecyl sulfate-polyacrylamide gel electrophoresis, and analyzed by immunoblotting with specific antibody (α -Npt, α -YopR, or α -Cat). (C) *Y. enterocolitica* strain W22703 (wild type) was transformed with plasmids encoding the *yopQ* hybrids described for panel A and analyzed as described for panel B.

and YopQ₁₋₁₅-Ub-Npt in *Y. enterocolitica*. Neither YopE₁₋₁₅-Ub-Npt nor YopQ₁₋₁₅-Ub-Npt was transported by the type III pathway (Fig. 1). In contrast, YopE₁₋₁₅-Npt and YopQ₁₋₁₅-Npt as well as YopE₁₋₁₅-Ub_{Gly3,13}-Npt were efficiently secreted, suggesting that the folding of the Ub domain interfered with transport by the type III machinery. We observed only small amounts of secretion (less than 10%) for YopQ₁₋₁₅-Ub_{Gly3,13}-Npt (Fig. 1C). It is conceivable that fusion of the Ub domain affects signaling of the *yopQ*₁₋₁₅ secretion signal. Nevertheless, we cannot yet provide a definitive explanation for the observed differences in secretion between YopE₁₋₁₅-Ub_{Gly3,13}-Npt and YopQ₁₋₁₅-Ub_{Gly3,13}-Npt.

As YopE₁₋₂₂₀-Ub-Npt, YopE₁₋₁₅-Ub-Npt, YopQ₁₋₁₉₈-Ub-Npt, and YopQ₁₋₁₅-Ub-Npt cannot be transported by the type III machinery because of their fused Ub domains, we expected these hybrids to interfere with the secretion of other Yop proteins. We were surprised to find that the secretions of YopE, YopQ, and YopR were not affected by the expression of hybrid Ub fusions (Fig. 1; also data not shown). The results suggested that the type III secretion pathway, a presumed channel within the needle structure (25, 31), cannot be occluded by folded proteins carrying Yop secretion signals. It should be emphasized that this property of the *Yersinia* type III machinery is distinct from that of other secretion pathways in which recognition of signal peptides leads to a productive and irrevocable interaction between a protein substrate and its translocation machinery.

To test the generality of this conjecture, we examined the secretion of YopE fusions to mouse dihydrofolate reductase (DHFR). Translational fusion of genes encoding mitochondrial precursor proteins to DHFR results in hybrid proteins that can be transported into mitochondria, because secretion chaperones unravel the folded precursor prior to translocation (26). Incubation of the *in vitro* translocation substrates with methotrexate, an active-site ligand of DHFR, irreversibly arrests the fused DHFR domain in a fully folded conformation and jams the import machinery (19). Fusion of an N-terminal signal peptide to DHFR causes degradation of the hybrid protein in the cytoplasm of *E. coli* (22). No secretion of the signal peptide DHFR hybrid was observed; however, it was also not clear whether the hybrid protein is capable of jamming the secretory pathway (22). Fusion of the DHFR C terminal to the secretion signal of *E. coli* hemolysin (HlyA) also did not lead to efficient secretion (39). Thus, although DHFR can be transported across membranes by the mitochondrial import machinery, not all membrane translocators can achieve the unfolding of DHFR.

To explore the fate of DHFR in the *Yersinia* type III pathway, YopE-DHFR hybrids were expressed in *Y. enterocolitica* W22703 (wild type) (Fig. 2). Even in the absence of methotrexate, yersiniae secreted only very small amounts of YopE₁₋₂₂₀-DHFR (less than 5%) into the medium and altogether failed to transport YopE₁₋₁₅-DHFR (Fig. 2). Expression of YopE₁₋₂₂₀-DHFR or YopE₁₋₁₅-DHFR in Δ (*yopE*) or Δ (*syncE*) mutant strains did not alter the ability of yersiniae to transport either native Yops or the hybrid proteins (Fig. 2; also data not shown). As previously reported, the Δ (*syncE*) mutant strains contain less YopE or YopE fusion than do wild-type yersiniae (Fig. 2; also data not shown). Together these data suggest that SyncE cannot unfold the fused Ub or DHFR domains and that

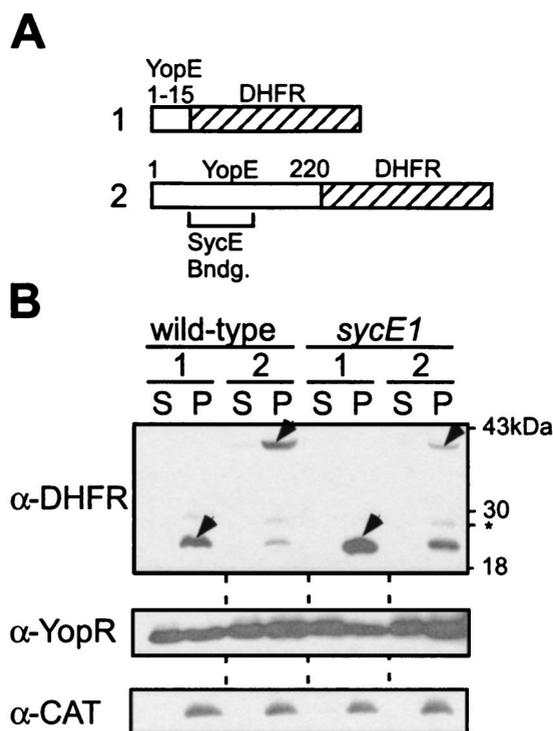


FIG. 2. Dihydrofolate reductase fusions are not secreted by the type III machinery of *Y. enterocolitica*. (A) The drawing depicts the primary structures of hybrid polypeptides as follows: line 1, YopE₁₋₁₅-DHFR, *yop* codons 1 to 15 fused to DHFR; line 2, YopE₁₋₂₂₀-DHFR, *yop* codons 1 to 220 (full-length) fused to dihydrofolate reductase (DHFR). (B) *Y. enterocolitica* strains W22703 (wild type) and LC1 Δ (*syncE1*) were transformed with plasmids described above, and type III secretion was analyzed by immunoblotting with specific antibody (α -DHFR, α -YopR, or α -Cat) as described in the legend to Fig. 1.

the function of SycE is limited to the substrate recognition of YopE.

One simple explanation for a failure of fusion proteins to block the type III pathway could be their aggregation in the bacterial cytoplasm. To test whether fusion proteins are soluble in the cytoplasm, bacterial extracts were generated in a French pressure cell (14,000 lb/in²) and unbroken cells were removed by slow-speed centrifugation (4,000 \times *g* for 15 min). The supernatant was removed, and lysates were subjected to ultracentrifugation at 100,000 \times *g* for 30 min. Soluble supernatant and insoluble sediment were separated and analyzed by immunoblotting. Most of YopE₁₋₂₂₀-Npt, YopE₁₋₂₂₀-Ub-Npt, and YopE₁₋₂₂₀-Ub_{3,13}-Npt did not sediment at 100,000 \times *g* and remained in the supernatant (Fig. 3). Thus, it appears that the fusion proteins are soluble in the bacterial cytoplasm and do not form aggregates that are hindered in their interaction with the type III secretion machinery.

We wish to propose a model that accounts for all of the observations reported here. Yop fusions to folded domains are not permanently engaged by the type III machinery but are rejected from the pathway (presumably once and for all) shortly after the synthesis of the secretion signal has been completed. If the secretion signal is recognized and the substrate is accommodated by the type III machinery, transport will be initiated and completed. If, however, the secretion

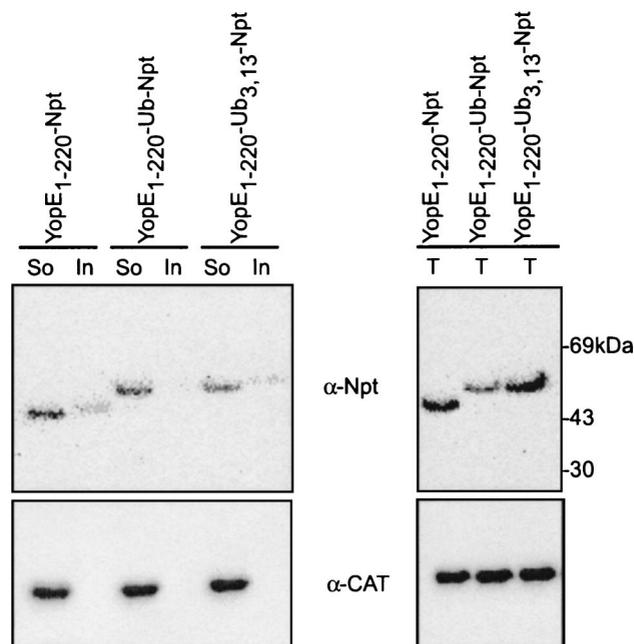


FIG. 3. YopE₁₋₂₂₀-Npt, YopE₁₋₂₂₀-Ub-Npt, and YopE₁₋₂₂₀-Ub_{Gly3,13}-Npt are soluble proteins. *Y. enterocolitica* W22703 expressing either YopE₁₋₂₂₀-Npt, YopE₁₋₂₂₀-Ub-Npt, or YopE₁₋₂₂₀-Ub_{Gly3,13}-Npt was lysed in a French pressure cell. Crude extracts were subjected to ultracentrifugation at 100,000 \times *g*. The soluble supernatant (So) and insoluble sediment (In) were separated, and proteins were precipitated with TCA and analyzed by immunoblotting (α -Npt or α -CAT). In the right panel, the signal intensities of Yop fusions in cell extracts are compared with the signal intensities in samples obtained by TCA precipitation of total cultures (T).

signal is recognized and the substrate cannot be accommodated, the protein will be rejected and cannot be reconsidered (or reinitiated) for transport by the type III machinery. Are this model and the data reported here compatible with the signal peptide hypothesis whereby an amphipathic helical peptide initiates substrates into the type III pathway? We think the results argue against a signal peptide mechanism of type III secretion, as this model would predict the initiation of signal peptide-bearing substrates as well as the block of a pathway charged with substrates that cannot be transported. In contrast, the RNA signal hypothesis may account for the observed substrate rejection within the type III secretion pathway. Assuming that the RNA signals of *yop* transcripts couple translation to secretion of the polypeptide, one could predict that the N-terminal portion of Yops is initiated into the type III pathway. Once the polypeptide has been completed and is folded, the rejected Yop-Ub or Yop-DHFR hybrids cannot reenter the pathway because the protein product has been separated from the RNA signal.

We think it unlikely that an entire Yop protein is transported in a cotranslational, unfolded manner, as the needle structures are longer than a linear polypeptide of 220 residues (3). Yop fusions to small (76-residue ubiquitin, 8-kDa) or medium-sized (187-residue DHFR, 21-kDa) domains are obviously permitted to fold and can be rejected at an undefined step along the pathway. We wondered whether the size of the type III machinery is incompatible with the size of folded

ubiquitin or DHFR. By using the crystallographic coordinates and computer simulation (40, 54), conservative estimates for the smallest diameters of folded ubiquitin and DHFR are 22 and 27 Å, respectively (data not shown). The diameter of the lumen of type III needle, a conduit through which Yop proteins are presumed to travel, is measured at about 20 Å (11, 25, 31). Thus, it seems that folded Yop proteins (52) or Yop fusions to folded ubiquitin or DHFR are too large to be transported through the lumen of the needle structure. In summary, we conclude that substrate recognition of type III machines occurs by unique mechanisms with governing principles that are distinct from those of co- and posttranslational translocation by the Sec pathway.

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REFERENCES

- Anderson, D. M., D. Fouts, A. Collmer, and O. Schneewind. 1999. Reciprocal secretion of proteins by the bacterial type III machines of plant and animal pathogens suggests universal recognition of mRNA targeting signals. *Proc. Natl. Acad. Sci. USA* **96**:12839–12843.
- Anderson, D. M., K. S. Ramamurthi, C. Tam, and O. Schneewind. 2002. YopD and LcrH regulate the expression of *Yersinia enterocolitica* YopQ at a post-transcriptional step and bind to *yopQ* mRNA. *J. Bacteriol.* **184**:1287–1295.
- Anderson, D. M., and O. Schneewind. 1997. A mRNA signal for the type III secretion of Yop proteins by *Yersinia enterocolitica*. *Science* **278**:1140–1143.
- Anderson, D. M., and O. Schneewind. 1999. Type III machines of Gram-negative pathogens: injecting virulence factors into host cells and more. *Curr. Opin. Microbiol.* **2**:18–24.
- Anderson, D. M., and O. Schneewind. 1999. *Yersinia enterocolitica* type III secretion: an mRNA signal that couples translation and secretion of YopQ. *Mol. Microbiol.* **31**:1139–1148.
- Baker, R. T., J. T. Tobias, and A. Varshavsky. 1992. Ubiquitin-specific proteases of *Saccharomyces cerevisiae*. *J. Biol. Chem.* **267**:23364–23375.
- Benson, S. A., M. N. Hall, and T. J. Silhavy. 1985. Genetic analysis of protein export in *Escherichia coli* K12. *Annu. Rev. Biochem.* **54**:101–134.
- Bieker, K. L., and T. J. Silhavy. 1990. PrlA (SecY) and PrlG (SecE) interact directly and function sequentially during protein translocation in *E. coli*. *Cell* **61**:833–842.
- Blobel, G. 1980. Intracellular protein topogenesis. *Proc. Natl. Acad. Sci. USA* **77**:1496–1500.
- Blobel, G., and B. Dobberstein. 1975. Transfer of proteins across membranes. I. Presence of proteolytically processed and unprocessed nascent immunoglobulin light chains on membrane-bound ribosomes of murine myeloma. *J. Cell Biol.* **67**:835–851.
- Blocker, A., N. Jouihri, E. Larquet, P. Gounon, F. Ebel, C. Parsot, P. Sansonetti, and A. Allaoui. 2001. Structure and composition of the *Shigella flexneri* 'needle complex', a part of its type III secretin. *Mol. Microbiol.* **39**:652–663.
- Brown, J. L., D. M. Brown, and I. Zabin. 1967. Thiogalactoside transacetylase. Physical and chemical studies of subunit structure. *J. Biol. Chem.* **242**:4254–4258.
- Cheng, L. W., D. M. Anderson, and O. Schneewind. 1997. Two independent type III secretion mechanisms for YopE in *Yersinia enterocolitica*. *Mol. Microbiol.* **24**:757–765.
- Cheng, L. W., and O. Schneewind. 1999. *Yersinia enterocolitica* type III secretion: on the role of SycE in targeting YopE into HeLa cells. *J. Biol. Chem.* **274**:22102–22108.
- Cornelis, G. R., A. Boland, A. P. Boyd, C. Geuijen, M. Iriarte, C. Neyt, M.-P. Sory, and I. Stainier. 1998. The virulence plasmid of *Yersinia*, an antihost genome. *Microbiol. Mol. Biol. Rev.* **62**:1315–1352.
- Cornelis, G. R., and C. Colson. 1975. Restriction of DNA in *Yersinia enterocolitica* detected by the recipient ability for a derepressed R factor from *Escherichia coli*. *J. Gen. Microbiol.* **87**:285–291.
- Dalbey, R. E., and W. Wickner. 1985. Leader peptidase catalyzes the release of exported proteins from the outer surface of the *Escherichia coli* plasma membrane. *J. Biol. Chem.* **260**:15925–15931.
- Economou, A., J. A. Pogliano, J. Beckwith, D. B. Oliver, and W. Wickner. 1995. SecA membrane cycling at SecYEG is driven by distinct ATP binding and hydrolysis events and is regulated by SecD and SecF. *Cell* **83**:1171–1181.
- Eilers, M., and G. Schatz. 1986. Binding of a specific ligand inhibits import of a purified precursor protein into mitochondria. *Nature* **322**:228–232.
- Emr, S., and T. J. Silhavy. 1980. Mutations affecting localization of an *Escherichia coli* outer membrane protein, the bacteriophage λ receptor. *J. Mol. Biol.* **141**:63–90.
- Emr, S. D., S. Hanley-Way, and T. J. Silhavy. 1981. Suppressor mutations that restore export of a protein with a defective signal sequence. *Cell* **23**:79–88.
- Gentz, R., Y. Kuys, C. Zwieb, D. Taatjes, H. Taatjes, W. Bannwarth, D. Stueber, and I. Ibrahim. 1988. Association of degradation and secretion of three chimeric polypeptides in *Escherichia coli*. *J. Bacteriol.* **170**:2212–2220.
- Glaser, P., H. Sakamoto, J. Bellalou, A. Ullmann, and A. Danchin. 1988. Secretion of cyclolysin, the calmodulin-sensitive adenylate cyclase-hemolysin bifunctional protein of *Bordetella pertussis*. *EMBO J.* **7**:3997–4004.
- Hartl, F. U., S. Lecker, E. Schiebel, J. P. Hendrick, and W. Wickner. 1990. The binding cascade of SecB to SecA to SecY/E mediates preprotein targeting to the *E. coli* plasma membrane. *Cell* **63**:269–279.
- Hoiczky, E., and G. Blobel. 2001. Polymerization of a single protein of the pathogen *Yersinia enterocolitica* into needles punctures eukaryotic cells. *Proc. Natl. Acad. Sci. USA* **98**:4669–4674.
- Hurt, E. C., B. Pesold-Hurt, and G. Schatz. 1984. The amino-terminal region of an imported mitochondrial precursor polypeptide can direct cytoplasmic dihydrofolate reductase into the mitochondrial matrix. *EMBO J.* **3**:3149–3156.
- Ito, K., P. J. J. Bassford, and J. Beckwith. 1981. Protein localization in *E. coli*: is there a common step in the secretion of periplasmic and outer membrane proteins? *Cell* **24**:707–717.
- Johnsson, N., and A. Varshavsky. 1994. Ubiquitin-assisted dissection of protein transport across membranes. *EMBO J.* **13**:2686–2698.
- Kalnins, A., K. Otto, U. Ruether, and B. Mueller-Hill. 1983. Sequence of the *lacZ* gene of *Escherichia coli*. *EMBO J.* **2**:593–597.
- Kiino, D. R., and T. J. Silhavy. 1984. Mutation in *prf1* relieves the lethality associated with export of beta-galactosidase hybrid proteins in *Escherichia coli*. *J. Bacteriol.* **158**:878–883.
- Kubori, T., Y. Matsushima, D. Nakamura, J. Uralil, M. Lara-Tejero, A. Sukhan, J. E. Galan, and S.-I. Aizawa. 1998. Supermolecular structure of the *Salmonella typhimurium* type III protein secretion system. *Science* **280**:602–605.
- Kumamoto, C. A. 1989. *Escherichia coli* SecB protein associates with exported protein precursors *in vivo*. *Proc. Natl. Acad. Sci. USA* **86**:5320–5324.
- Langley, K. E., M. R. Villarejo, A. V. Fowler, P. J. Zamenhof, and I. Zabin. 1975. Molecular basis of beta galactosidase alpha-complementation. *Proc. Natl. Acad. Sci. USA* **72**:1254–1257.
- Lipp, J., B. Dobberstein, and M. T. Haeuptle. 1987. Signal recognition particle arrests elongation of nascent secretory and membrane proteins at multiple sites in a transient manner. *J. Biol. Chem.* **262**:1680–1684.
- Lloyd, S. A., M. Norman, R. Rosqvist, and H. Wolf-Watz. 2001. *Yersinia* YopE is targeted for type III secretion by N-terminal, not mRNA, signals. *Mol. Microbiol.* **39**:520–531.
- Lloyd, S. A., M. Sjostrom, S. Andersson, and H. Wolf-Watz. 2002. Molecular characterization of the type III secretion signals via analysis of synthetic N-terminal amino acid sequences. *Mol. Microbiol.* **43**:51–59.
- Meyer, D. I., E. Krause, and B. Dobberstein. 1982. Secretory protein translocation across membranes—the role of the “docking protein.” *Nature* **297**:647–650.
- Michiels, T., and G. R. Cornelis. 1991. Secretion of hybrid proteins by the *Yersinia* Yop export system. *J. Bacteriol.* **173**:1677–1685.
- Nakano, H., Y. Kawakami, and H. Nishimura. 1992. Secretion of genetically-engineered dihydrofolate reductase from *Escherichia coli* using an *E. coli* alpha-hemolysin membrane translocation system. *Appl. Microbiol. Biotechnol.* **37**:765–771.
- Oefner, C., A. D'Arcy, and F. K. Winkler. 1988. Crystal structure of human dihydrofolate reductase complexed with folate. *Eur. J. Biochem.* **174**:377–385.
- Oliver, D. B., and J. Beckwith. 1981. *E. coli* mutant pleiotropically defective in the export of secreted proteins. *Cell* **25**:765–772.
- Poritz, M. A., H. D. Bernstein, K. Strub, D. Zopf, H. Wilhelm, and P. Walter. 1990. An *E. coli* ribonucleoprotein containing 4.5S RNA resembles mammalian signal recognition particle. *Science* **250**:1111–1117.
- Powers, T., and P. Walter. 1997. Co-translational protein targeting catalyzed by the *Escherichia coli* signal recognition particle and its receptor. *EMBO J.* **16**:4880–4886.
- Ramamurthi, K. S., and O. Schneewind. 2002. *Yersinia enterocolitica* type III secretion: mutational analysis of the *yopQ* secretion signal. *J. Bacteriol.* **184**:3321–3328.
- Randall, L. L. 1992. Peptide binding by chaperone SecB: implications for recognition of non-native structure. *Science* **257**:241–245.
- Randall, L. L., and S. J. Hardy. 1986. Correlation of competence for export with lack of tertiary structure of the mature species: a study *in vivo* of maltose-binding protein in *E. coli*. *Cell* **46**:921–928.
- Reiss, B., R. Sprengel, and H. Schaller. 1984. Protein fusions with the kanamycin resistance gene from transposon Tn5. *EMBO J.* **3**:3317–3322.

48. **Schatz, P. J., and J. Beckwith.** 1990. Genetic analysis of protein export in *Escherichia coli*. *Annu. Rev. Genet.* **24**:215–248.
49. **Schesser, K., E. Fritz-Lindsten, and H. Wolf-Watz.** 1996. Delineation and mutational analysis of the *Yersinia pseudotuberculosis* YopE domains which mediate translocation across bacterial and eukaryotic cellular membranes. *J. Bacteriol.* **178**:7227–7233.
50. **Sory, M.-P., A. Boland, I. Lambermont, and G. R. Cornelis.** 2002. 1995. Identification of the YopE and YopH domains required for secretion and internalization into the cytosol of macrophages, using the *cyaA* gene fusion approach. *Proc. Natl. Acad. Sci. USA* **92**:11998–12001.
51. **Sory, M.-P., and G. R. Cornelis.** 1994. Translocation of a hybrid YopE-adenylate cyclase from *Yersinia enterocolitica* into HeLa cells. *Mol. Microbiol.* **14**:583–594.
52. **Stuckey, J. A., H. L. Schubert, E. B. Fauman, Z. Y. Zhang, J. E. Dixon, and M. A. Saper.** 1994. Crystal structure of *Yersinia* protein tyrosine phosphatase at 2.5 Å and the complex with tungstate. *Nature* **370**:571–575.
53. **Takeshita, S., M. Sato, M. Toba, W. Masahashi, and T. Hashimoto-Gotoh.** 1987. High-copy-number and low-copy-number plasmid vectors for LacZ alpha-complementation and chloramphenicol- or kanamycin-resistance selection. *Gene* **61**:63–74.
54. **Vijay-Kumar, S., C. E. Bugg, and W. J. Cook.** 1987. Structure of ubiquitin refined at 1.8 Å resolution. *J. Mol. Biol.* **194**:531–544.
55. **Walter, P., and G. Blobel.** 1980. Purification of a membrane-associated protein complex required for protein translocation across the endoplasmic reticulum. *Proc. Natl. Acad. Sci. USA* **77**:7112–7116.
56. **Walter, P., and G. Blobel.** 1982. Signal recognition particle contains a 7S RNA essential for protein translocation across the endoplasmic reticulum. *Nature* **299**:691–698.
57. **Walter, P., R. Keenan, and U. Schmitz.** 2000. SRP—where the RNA and membrane worlds meet. *Science* **287**:1212–1213.
58. **Wattiau, P., and G. R. Cornelis.** 1993. SycE, a chaperone-like protein of *Yersinia enterocolitica* involved in the secretion of YopE. *Mol. Microbiol.* **8**:123–131.