

Differences in Gene Content between *Salmonella enterica* Serovar Enteritidis Isolates and Comparison to Closely Related Serovars Gallinarum and Dublin†

S. Porwollik,¹ C. A. Santiviago,¹ P. Cheng,¹ L. Florea,² S. Jackson,³ and M. McClelland^{1*}

Sidney Kimmel Cancer Center, 10835 Road to the Cure, San Diego, California 92121,¹ George Washington University, 801 22nd Street NW, Suite 704, Washington, DC 20052,² and Center for Food Safety and Applied Nutrition, Food and Drug Administration, 8301 Muirkirk Rd., Laurel, Maryland 20708³

Received 22 February 2005/Accepted 15 June 2005

Salmonella enterica serovar Enteritidis is often transmitted into the human food supply through eggs of hens that appear healthy. This pathogen became far more prevalent in poultry following eradication of the fowl pathogen *S. enterica* serovar Gallinarum in the mid-20th century. To investigate whether changes in serovar Enteritidis gene content contributed to this increased prevalence, and to evaluate genetic heterogeneity within the serovar, comparative genomic hybridization was performed on eight 60-year-old and nineteen 10- to 20-year-old serovar Enteritidis strains from various hosts, using a *Salmonella*-specific microarray. Overall, almost all the serovar Enteritidis genomes were very similar to each other. Excluding two rare strains classified as serovar Enteritidis in the *Salmonella* reference collection B, only eleven regions of the serovar Enteritidis phage type 4 (PT4) chromosome (sequenced at the Sanger Center) were absent or divergent in any of the other serovar Enteritidis strains tested. The more recent isolates did not have consistent differences from 60-year-old field isolates, suggesting that no large genomic additions on a whole-gene scale were needed for serovar Enteritidis to become more prevalent in domestic fowl. Cross-hybridization of phage genes on the array with related genes in the examined genomes grouped the serovar Enteritidis isolates into two major lineages. Microarray comparisons of the sequenced serovar Enteritidis PT4 to isolates of the closely related serovars Dublin and Gallinarum (biovars Gallinarum and Pullorum) revealed several genomic areas that distinguished them from serovar Enteritidis and from each other. These differences in gene content could be useful in DNA-based typing and in understanding the different phenotypes of these related serovars.

Salmonella enterica serovar Enteritidis is one of the more than 2,500 serovars of *S. enterica* known. While previously known as a frequent pathogen in rodents, serovar Enteritidis arose in the 1980s as a serious problem in the human food supply, in part due to the fact that infected but asymptomatic chickens can pass on the bacteria in undercooked eggs (23). The incidence of serovar Enteritidis in the United States rose dramatically until the mid-1990s, from a per capita rate of 0.55 per 100,000 in 1976 (5% contribution to total *Salmonella* infections) to a record high of 3.9 per 100,000 in 1995 (more than 10,000 reported incidences; 25% of the total). Subsequently, the number of recorded serovar Enteritidis infections decreased to 5,116 in 2002 (16% of the total) (<http://www.cdc.gov/ncidod/dbmd/phlisdata/salmtab/2002/SalmonellaAnnualSummary2002.pdf>; 25), an effect probably caused by targeted interventions like refrigeration, on-farm prevention and control measures, education of food workers, and egg quality assurance programs (22). However, serovar Enteritidis remains the second most prevalent cause of *Salmonella* infection in humans, after serovar Typhimurium.

Serovar Enteritidis is most closely related to *S. enterica* se-

rovors Dublin and Gallinarum based on gene content (26) and multilocus sequence typing (M. Achtman, personal communication). Serovar Gallinarum is of particular interest because it is a fowl-specific serovar that causes typhoid and diarrhea in poultry (5), with a high mortality rate. The Gallinarum serovar is currently divided into two biovars, Pullorum and Gallinarum, with serovar Gallinarum biovar Pullorum primarily infecting young chicks and serovar Gallinarum biovar Gallinarum being able to cause systemic disease in both young and adult hens (1). Because of the profound adverse effect of serovar Gallinarum on the poultry industry, a nationwide eradication effort by means of culling seropositive birds was implemented in the United States and the United Kingdom in the 1930s, effectively removing serovar Gallinarum from commercial poultry flocks by the mid-1970s (2). Subsequently, isolates of serovar Enteritidis were detected in fowl, usually leaving hens asymptomatic but infecting eggs, and thus humans, by contamination from chicken feces and through infection of the hen reproductive tract. The immunodominant surface antigen of serovar Enteritidis isolates is a tyvelose branching sugar, named O9 (2). Serovar Gallinarum biovar Gallinarum and biovar Pullorum both contain the same O antigen, and it is possible that previous to their eradication they prevented serovar Enteritidis isolates from becoming widespread in domestic poultry (2, 31).

A phage-typing scheme developed by Ward et al. (38) originally differentiated 27 phage types (PT) within serovar Enteritidis. Whereas the current predominant serovar Enteritidis

* Corresponding author. Mailing address: Sidney Kimmel Cancer Center, 10835 Road to the Cure, San Diego, CA 92121. Phone: (858) 450-5990. Fax: (858) 450-3251. E-mail: mmcclelland@skcc.org.

† Supplemental material for this article may be found at <http://jb.asm.org/>.

phage type isolated worldwide is PT4 (18, 25, 37), PT8 isolates were initially the most commonly found in the United States and the United Kingdom (11, 39). Today, these two phage types and PT13a account for the majority of serovar Enteritidis infections reported. The genomes of the different serovar Enteritidis phage types were found to be highly similar, so it was difficult to distinguish them by pulsed-field gel electrophoresis analysis (12, 16). Isolates could be discriminated better by ribotyping techniques, but different phage types did not necessarily result in different ribotypes. Therefore, combinatorial methods of genome characterization have been suggested for epidemiological studies of serovar Enteritidis (15, 32). Two separate lineages of serovar Enteritidis phage types have been proposed, based on differences in the lipopolysaccharide (LPS) core region: PT4-like isolates (including PT1, -4, -4b, -6, -7, and -24) and PT8-like strains (including PT2, -8, -13a, and -23) (9, 17). These differences in the LPS are thought to render the strains resistant or susceptible to a specific phage present in the typing scheme developed by Ward et al.

Sequence and annotation information on the complete genome sequences of four *Salmonella* strains covering three serovars, Typhimurium, Typhi, and Paratyphi A, are currently available (7, 19, 20, 24). In addition, genome sequences for at least seven other serovars are being produced (28), including serovar Enteritidis and both biovars of serovar Gallinarum, as well as the only other species within the genus *Salmonella*, *S. bongori*. An *S. enterica* serovar Enteritidis PT4 strain has been sequenced by the Sanger Center in the United Kingdom, and the sequence is freely available from the Center (<ftp://ftp.sanger.ac.uk/pub/pathogens/Salmonella/SePT4.dbs>). However, the completed genome sequence still awaits final annotation. We used the sequence to complement our custom-made microarray of serovar Typhimurium-Typhi-Paratyphi A genes (19) by adding probes representing genes uniquely present in serovar Enteritidis PT4. Thus, the array now covers four major serovars of *S. enterica*. This array was used to assess genomic differences between serovar Enteritidis strains before and after the eradication of serovar Gallinarum from fowl and between different phage types of recent serovar Enteritidis isolates. In addition, serovar Dublin and serovar Gallinarum (biovar Gallinarum and Pullorum) isolates were investigated to reveal differences from serovar Enteritidis. The results of the genomic comparisons revealed 11 chromosomal regions of diversity within frequently occurring serovar Enteritidis strains and two possible signature divergences each for biovar Pullorum and biovar Gallinarum isolates.

MATERIALS AND METHODS

Strains. The *Salmonella enterica* strains subjected to microarray analysis in this study are listed in Table 1. Genomic DNA was harvested from fresh overnight cultures grown in LB under standard conditions and labeled as described by Porwollik et al. (26).

Microarray construction. The construction of the basic *Salmonella enterica* serovar Typhimurium LT2 DNA microarray, as well as its supplementation with serovar Typhi CT18-specific probes, has been previously described (27, 30). In this study, we used an array that is an expansion of this nonredundant microarray. Overall, the current microarray consists of PCR-amplified sequences from the annotated open reading frames in *S. enterica* serovar Typhimurium LT2 (STM), supplemented with gene-specific probes generated by PCR from chromosomal coding sequences from strains Typhi CT18, Paratyphi A SARB42, and Enteritidis PT4, which were more than 10% divergent from Typhimurium LT2 and from each other. Overall, *S. enterica* serovar Typhimurium LT2 genome

coverage for the array is 99.4% (4,466 genes), coverage of the Typhi CT18 genome is 98.3% (4,521 genes), and coverage of the Paratyphi A SARB42 genome is 98.4% (4,193 genes), excluding plasmids. Coverage of the Enteritidis PT4 genome is approximately 92 to 98% (the exact figure awaits the official annotation by the Sanger Center). We automatically annotated the genome sequence of Enteritidis PT4 (<http://www.sanger.ac.uk/Projects/Salmonella/>) using the automatic annotation software programs Generation (Oak Ridge National Laboratory, Oak Ridge, Tennessee) and Glimmer (The Institute for Genomic Research, Rockville, Maryland) (6). Putative genes with 95% identity in a window of 100 bases with any sequence already on the array were removed, as were one copy of duplicates between the two annotation methods. Primers were designed for the remaining 390 “serovar Enteritidis-specific” genes using Primer3 (Massachusetts Institute of Technology, Boston), and products of 100 bp to 1 kb in size were amplified. The DNA was spotted onto Ultra-GAPS glass slides (Corning Inc., Corning, New York) in 50% dimethyl sulfoxide.

Hybridization and data acquisition. Standard protocols for hybridizations in formamide buffer (http://www.corning.com/Lifesciences/technical_information/techDocs/gaps_ii_manual_protocol_5_02_cls_gaps_005.pdf) were applied for prehybridization, hybridization, and posthybridization wash processes. Immediately before use, the labeled genomic DNAs of Enteritidis PT4 (control sample) and one of the query *S. enterica* strains (experimental sample) were unified, the volume was adjusted to 40 μ l, and it was mixed with 40 μ l of 2 \times hybridization buffer (50% formamide, 10 \times SSC [1 \times SSC is 0.15 M NaCl plus 0.015 M sodium citrate], 0.2% sodium dodecyl sulfate) and boiled for 5 min. After application to the array, the samples hybridized to the probes overnight at 42°C and were subsequently washed according to the Corning manual. A ScanArray 5000 laser scanner (Packard BioChip Technologies, Billerica, Massachusetts) with ScanArray 2.1 software was used for image acquisition. Signal intensities were quantified using the QuantArray 3.0 software package (Packard BioChip Technologies, Billerica, Massachusetts).

Microarray data analysis. Three arrays were spotted onto each glass slide, so each experiment resulted in three hybridization ratios per gene. Spots were analyzed by adaptive quantitation. The local background was subtracted from the spot intensities, and the contribution of each “true” spot intensity to the total signal in each channel was calculated for normalization. Subsequently, the median of the three ratios per gene was recorded. Spots representing genes present in PT4 but exhibiting low signal when hybridized with PT4 DNA (median contribution of the three spots to the total signal among the lowest 5% of all PT4 genes) were assigned to the category “uncertain.” Similarly, spots that had background values higher than 20% of the median signal value of all spots in the given channel were set aside as unknown and not processed further. For the remaining spots, absence/presence calls for genes were made according to the following parameters. For chromosomal genes predicted to be present in serovar Enteritidis PT4, unknown-strain/PT4 signal ratios that were higher than 0.67 indicated presence, ratios lower than 0.33 indicated absence, and intermediate ratios indicated uncertain status. For all genes absent from serovar Enteritidis PT4 but present in Typhimurium LT2, Typhi CT18, and/or Paratyphi A SARB42, the following criteria were employed. If the median contribution of the three spots per gene was among the top 70% of all genes represented on the array and the ratio between the query strain and PT4 was over 2.5, the gene was called present. If the median of the three spots was among the bottom 20% of all genes on the array, the gene was called absent. Spots that fell outside of these categories were called “uncertain.” For genes present in the Enteritidis PT4 virulence plasmid, similar thresholds were employed, excluding the necessity for a ratio greater than 2.5 for presence calls.

Comparison of data obtained using our array with available sequenced genomes showed that gene status predictions after a single hybridization experiment on our triplicate array had an estimated error rate of less than 1% (26). Confidence in the predictions of gene absence or presence is consequently high, but not absolute.

Regions that are described as different among strains in this study have been selected based on two parameters: their overall size had to be 1 kb or more, and they had to be represented by at least two spots on the microarray. If elements displayed different absence/presence patterns but were less than 1 kb apart from each other, the two areas are presented separately as subregions.

Note that one of the biovar Gallinarum isolates included in this survey was the unfinished sequenced strain 287/91, the genome sequence of which is generated at the Sanger Center, and predictions of gene presence in this isolate are based on in silico comparisons only. A sliding window of 100 bp was applied for all PCR-amplified sequences on our array, and the best hit for each spot was recorded. If best hits were equal to or over 95% similarity, the gene was called present in the respective sequenced isolate. If similarities were below 85%, the

TABLE 1. Strains used in this study

Strain no.	Strain name	Serovar/biovar	Phage or MLEE type	Isolation ^b	Source
1	MZ 648	Enteritidis	PT1	Denmark, 1985, human	J. Garaizar
2	MZ 649	Enteritidis	PT2	Denmark, 1987, human	J. Garaizar
3	MZ 634	Enteritidis	PT4	Spain, 1983, human	J. Garaizar
4	MZ 636	Enteritidis	PT4b	Spain, 1993, water	J. Garaizar
5	MZ 637	Enteritidis	PT5	Spain, 1993, food	J. Garaizar
6	MZ 638	Enteritidis	PT6a	Spain, 1994, water	J. Garaizar
7	MZ 635	Enteritidis	PT7a	Spain, 1983, human	J. Garaizar
8	SGSC2377	Enteritidis	PT8	Canada	K. Sanderson
9	MZ 650	Enteritidis	PT8	Denmark, 1983, chicken	J. Garaizar
10	21027	Enteritidis	PT13a		P. Fields
11	21046	Enteritidis	PT13a		P. Fields
12	MZ 655	Enteritidis	PT23	Denmark, 1997, chicken	J. Garaizar
13	MZ 646	Enteritidis	PT25	Denmark, 1996, human	J. Garaizar
14	MZ 672	Enteritidis	?	Germany, pigeon	A. Helm
15	MZ743	Enteritidis	PT6a	USA, KS, 1948, guinea pig	P. Fields
16	MZ744	Enteritidis	PT13a	USA, 1948, guinea pig	P. Fields
17	MZ745	Enteritidis	PT35	USA, GA, 1949	P. Fields
18	MZ746	Enteritidis	PT13a	USA, 1949	P. Fields
19	MZ747	Enteritidis	PT8	USA, NJ, 1950	P. Fields
20	MZ748	Enteritidis	PT13a	USA, 1950, mouse	P. Fields
21	MZ749	Enteritidis	PT4b	USA, CA, 1950, rat	P. Fields
22	MZ750	Enteritidis	PT8	USA, TX, 1950,	P. Fields
23	SARB16	Enteritidis	En1, PT13a	USA, Rhode Island	K. Sanderson
24	SARB18	Enteritidis	En3, PT4	USA, Connecticut	K. Sanderson
25	MZ 647	Dublin		Denmark, 1984, human	J. Garaizar
26	SARB12	Dublin	Du1	USA, Idaho, cattle, 1986	K. Sanderson
27	SARB13	Dublin	Du3	France, bovine, 1982	K. Sanderson
28	99113	Gallinarum/Pullorum		Canada, 1999	J. Guard-Bouldin
29	99117	Gallinarum/Pullorum		Canada, 1999	J. Guard-Bouldin
30	7865L	Gallinarum/Pullorum		USA, 1997	J. Frye
31	17368L	Gallinarum/Pullorum		USA, 2003	J. Frye
32	SARB51	Gallinarum/Pullorum	Pu3	Germany	K. Sanderson
33	Sg9	Gallinarum/Gallinarum			T. Wallis
34	SgJ91	Gallinarum/Gallinarum			T. Wallis
35	287/91	Gallinarum/Gallinarum		Sequenced isolate, 1991	Sanger Center
36	SARB52	Gallinarum/Pullorum	Pu4	Germany	K. Sanderson
37	SARB17	Enteritidis	En2, RDNC ^a	Brazil	K. Sanderson
38	SARB20	Enteritidis	En7, RDNC	Switzerland	K. Sanderson
39	MZ630	Enteritidis	PT4	sequenced isolate	Sanger Center

^a RDNC, reactive to phages in serovar Enteritidis phage-typing scheme (38) but do not conform to common phage types.

^b Location, year, source.

gene was called absent, and for intermediate homologies, an uncertain status was assumed.

Data deposition. The raw microarray data presented in this paper have been deposited at the GEO database of the National Center for Biotechnology at <http://www.ncbi.nlm.nih.gov/geo> under series number GSE2242.

RESULTS

Overview of identified genomic differences. The gene repertoires of 26 serovar Enteritidis strains, 9 serovar Gallinarum strains, and 3 serovar Dublin isolates were investigated in this study (Table 1). These data included serovar Enteritidis isolates that were nearly 60 years old and therefore were isolated before the rise of serovar Enteritidis infections in fowl, as well as younger field isolates. In addition, representative isolates of multilocus enzyme electrophoresis (MLEE) types have been included. These strains are part of the *Salmonella* reference collection B (SARB), a compendium of major MLEE variants of clinically relevant *S. enterica* serovars that was established more than 10 years ago (4).

Absence/presence predictions were generated for every probe on the array for every strain investigated. In addition,

predictions were also made for the *Salmonella* isolates that had complete or partial genome sequences available. The results are illustrated and summarized at single-spot resolution in Table S1 in the supplemental material. Overall, 11 regions of the PT4 chromosome were determined that were absent in at least one of the serovar Enteritidis field isolates tested (regions A01 to A11 in Table 2). Twenty-three additional segments of the PT4 genome were found to be different in strains that represent the most common MLEE types in serovars Gallinarum and Dublin, as well as recent serovar Gallinarum field isolates (regions B01 to B23). Fifty-four more chromosomal PT4 regions were determined that were absent in three unusual and rare SARB isolates—two serovar Enteritidis and one biovar Pullorum strain (see Tables 2, regions C01 to C54, in the supplemental material).

The plasmid present in Enteritidis PT4 has homology to the Typhimurium LT2 pSLT virulence plasmid over at least 57 kb, including the *pef* fimbrial locus, the *spv* locus involved in virulence, and the *par-sam* locus implicated in DNA partitioning and repair. The entire region of homology was absent in one

TABLE 2. Regions differing from the sequenced serovar Enteritidis PT4 isolate in serovar Enteritidis, Dublin, and Gallinarum strains^a

Region no. ^a	Location start	Approx size (kb)	Gene range ^c	Gene and description	Differing strains ^d			
					Enteritidis	Dublin	Gallinarum	Other SARB
Genes present in PT4								
A01 ^b	0249806	1.8			24		28–35	38
A02	0309634	6.7	STM0266–0289		24		28–35	36,37
A03	1275768	7.9	STM1834–1843	<i>rrmA</i>	3			
A04 ^b	1469682	11.6		Phage?	15,17	25–27	28–35	36–38
A05	1659634	3.2	STM1489–1491	<i>ynfK</i>	18			
A06 ^b	2020515	38.4		ST64B phage?	2,8–11,12,15–24		28–35	36–38
A07	2302417	2.5	STM2200–2201	<i>lysP</i>	18–20			
A08	2875524	12.6	STM2844–2858	Hydrogenase 3 operon	7			
A09	2911415	4.6	STM2883–2886	<i>sipBCD</i> , <i>sicA</i> , part of SPII	7			
A10	4366609	50.4	STM4270–4312	<i>dcu</i> , <i>pmr</i> , <i>nrf</i> , <i>mel</i> operon	7			
A11 ^b	4608741	9.4		DNA restriction/methylation?	1			36,37
A12	p_00096	5.4	pSLT001–009	DNA replication, <i>rck</i>	14	27	29	36–38
A13	p_07550	11.2	pSLT012–026	<i>pef</i> fimbriae	14,24	25–27	28–35	36–38
A14	p_19482	23.3	pSLT027–051	<i>spv</i> operon, resolvase, <i>ccd</i>	14		29	36–38
A15_all	p_43200	16.1	pSLT052–055, pSLT068–084	<i>par</i> , <i>sam</i> , SOS inhibition <i>psi</i> , part of <i>tra</i> locus	14	25–27	29,33,35	36–38
A15_sub	p_52504	2.0	pSLT073–077	Part of <i>tra</i> locus	10,11,15	25–27	29,33,35	36–38
B01	0093701	3.7	STM0081–0084			25,26		
B02	0353500	2.4	STM0330–0332	Transcriptional regulator?			32	36,37
B03	0593150	3.7	STM0554–0558	<i>yfd</i> glycosyltransferase		26		36
B04	0866128	1.5	STM0839–0840	<i>ybiV(2)</i>			28,30–32	36,38
B05 ^b	0890919	25.7					29	
B06	1013381	8.2	STM1004–1056	<i>sspH2</i>			28–35	37
B07_all	1242709	7.7	STM0910–0907 plus SPT spec	Similar to parts of phage Fels-1			28,30,31	37
B07_sub	1249229	1.2	STM0910–0909	Similar to parts of phage Fels-1		25–27	28–31	37
B08	1447962	10.8	STM1664–1675	<i>Yersinia invC</i> homolog			28,30–32	
B09 ^b	1520211	5.6		MFS transport?		25–27		36–38
B10 ^b	1533858	1.6					29	
B11	1549742	5.7	STM1590–1594	<i>srfA</i>			34	
B12	1591291	6.4	STM1558–1560	Glycosyl hydrolases		27		
B13_a	1600655	8.9	STM1544–1555	<i>pqaA</i>		25–27	29,33–35	
B13_b	1595657	4.9	STM1556–1558				32	
B14	1668378	2.1	STM1481–1483	Cation transport			32,35	
B15 ^b	1720290	4.7					29	
B16	1873762	6.6	STM1283–1286	Chemotaxis?			29,33–35	
B17 ^b	2128623	22.0					29	
B18	2331001	2.8	STM2226–2228	<i>yejKL</i>			35	
B19	2939355	4.2	STM2912–2915				30	
B20	2947941	3.3	STM2921–2924	<i>rpoS</i>			32	
B21	3071210	12.3	STM3022–3033	<i>std</i> fimbriae			29,33–35	
B22	3898009	2	STM3823–3824	<i>tor</i> system			28,30–32	
B23_a	4602467	2.7	STM4519–4521 plus SPT spec				28–35	
B23_b ^b	4606076	1.8					28–35	36,37
Genes absent in PT4								
A16	0309924	3.9	STM0271–0272		24	25–27	28–35	36,38
A17_a	1099565	4.6	STM1006–1009	Parts of phage Gifsy-2	24	25–27		36
A17_b	1104740	4.3	STM1010–1018	Parts of phage Gifsy-2	24	25–27		
A17_c	1109305	3.7	STM1019–1024	Parts of phage Gifsy-2	24	25–27		36
A17_d	1116857	9.1	STM1031–1040	Parts of phage Gifsy-2	24	25–27	28–35	38
A17_e	1126009	6.7	STM1041–1047	Parts of phage Gifsy-2	24	25–27	28–35	36,38
A18	2762064	1.7	STM2616–2618	Part of phage Gifsy-1	24			
A19_a	2844714	7.2	STM2695–2702	Parts of phage Fels-2	2,8–12,15–20,22,23	25–27		36,38

Continued on following page

TABLE 2—Continued

Region no. ^a	Location start	Approx size (kb)	Gene range ^c	Gene and description	Differing strains ^d			
					Enteritidis	Dublin	Gallinarum	Other SARB
A19_b	2852212	1.2	STM2703–2704	Parts of phage Fels-2	15,17			
A19_c	2855566	18.0	STM2707–2730	Parts of phage Fels-2	2,8–12,15–20,22,23	25	30,31	36
A20	48285	6.9	pSLT056–067	<i>ssbB</i>	24	27	28,30–35	
A21	67516	25.8	pSLT085–110	<i>tra</i> , <i>trb</i>	15,17,24	27	28,30–35	
A22	2766359	1.7	STY2887–2888	Part of serovar Typhi phage ST27	2,8–12,16–20,22,23	25	28,30,31	36,38
A23	3517867	2.7	STY3662–3667	<i>cII</i> , <i>dam</i> , part of serovar Typhi phage ST35	2,8–12,16,18–20,22,23		28,30,31	36
A24	3526926	12.8	STY3674–3691	<i>nuc</i> , part of ST35	2,8–12,15–20,22,23	25–27	28,30,31	36,38
A25	4498909	4.2	STY4632–4636	<i>Dam</i> , part of serovar Typhi phage SopE _{ST}	15			38
A26	2699408	25.4	SPA2602–2625	Serovar Paratyphi A phage SPA-3	20			38
B24	0306723	9.9	STM0268–0275			25–27		38
B25_a	0318679	13.4	STM0279–0289			25–27		38
B25_b	0332086	1.6	STM0290–0291			25–27		
B26	2873542	4.3	STM2731–2738	Parts of phage Fels-2		25–27		
B27_all	46_00830	31.6	R001–R046			26		
B27_sub	46_14184	7.5	R022–R027	<i>fip</i> , <i>tra</i> , <i>stb</i>		26	34	
B28	1883033	5.2	STY2003–2011	Restr/modification, hydrolase, start of Typhi phage ST18			29,35	
B29	1912209	1.2	STY2043–2045	Part of ST18		26,27		
B30	1921304	1.3	STY2060–2061	Part of ST18		27		
B31	2761219	7.6	STY2882–2889	Part of ST27			28,30,31	
B32	3540251	0.8	STY3693–3694	Part of ST35			28,30,31	
B33	4409652	64.1	STY4521–4597	SPI7: <i>samB pil topB</i>		27		
B34	4508347	21.4	STY4648–4666	SPI7: <i>vex/tvi</i>		27		
B35_a	2488061	7.6	SPA2387–2391	Part of serovar Paratyphi A phage SPA-1			32,34	
B35_b	2496420	10.7	SPA2393–2394	Parts of SPA-1		25,26	32,34	
B35_c	2510504	17.1	SPA2401–2431	Part of SPA-1			32,34	

^a Regions A are different among serovar Enteritidis strains; regions B have been identified in serovar Dublin and Gallinarum strains.

^b Regions are not present in sequenced and annotated genomes of serovar Typhimurium LT2, serovar Typhi CT18, serovar Typhi Ty2, or serovar Paratyphi A SARB42; pSLT, serovar Typhimurium LT2 plasmid; R46, transmission plasmid.

^c Range of homologous region in STM, STY, or SPA is indicated. STM, *S. enterica* serovar Typhimurium LT2; STY, *S. enterica* serovar Typhi CT18; SPA, *S. enterica* serovar Paratyphi A SARB42.

^d Strain numbers are as in Table 1.

serovar Enteritidis, one biovar Pullorum, and three rare SARB isolates tested. In addition, two plasmid regions were identified that were absent in two serovar Enteritidis strains (regions A13 and A15 in Table 2) and the serovar Dublin isolates. Region A13, encoding the *pef* fimbriae, was absent, or divergent, in all eight serovar Gallinarum strains tested by microarray.

The array contained genes present in the sequenced genomes of *S. enterica* serovar Typhimurium LT2, serovar Typhi CT18, and/or Paratyphi A SARB42 but absent in the sequenced Enteritidis PT4 isolate. Some of these regions were predicted to be present in several serovar Enteritidis strains (overall, 11 regions; A16 to A26 in Table 2). Twelve more segments were identified in serovar Dublin and serovar Gallinarum strains (regions B24 to B35), while the three isolates from the SARB collection that represented atypical members of these serovars contained another set of 23 regions that were absent from the PT4 genome (see Table S2, regions C55 to C77, in the supplemental material). The majority of the probes that have no homologue in PT4 but are present in other strains of serovar Enteritidis, serovar Gallinarum, or serovar Dublin include phage genes.

The absence/presence patterns for regions of diversity identified in this study are illustrated for all strains in Fig. 1.

Differences between old isolates and the PT4 genome sequence. The serovar Enteritidis strains isolated in the late 1940s and early 1950s represent five different phage types, PT13a (three isolates), PT8 (two isolates), and PT4b, -6a, and -35 (one isolate each). In general, these strains did not display many pronounced differences compared to the sequenced PT4 strain. Overall, among all eight “old” isolates tested, there were only four regions missing compared to the PT4 chromosome, and only one of these was missing in all eight of them—an ST64B-like phage region (region A06; approximately 38 kb). PT4 may have acquired this phage fairly recently.

Another 12-kb phage region (region A04) was missing from two of the old strains, representing phage types 6a and 35. This region encodes an Enteritidis PT4-specific phage not found in any other *Salmonella* serovar to date. The two remaining areas, regions A05 and A07, are small islands with similarities to three or two Typhimurium LT2 genes of uncertain function.

Among the elements not present in the sequenced Enteritidis PT4, we found six phage regions and one plasmid-derived

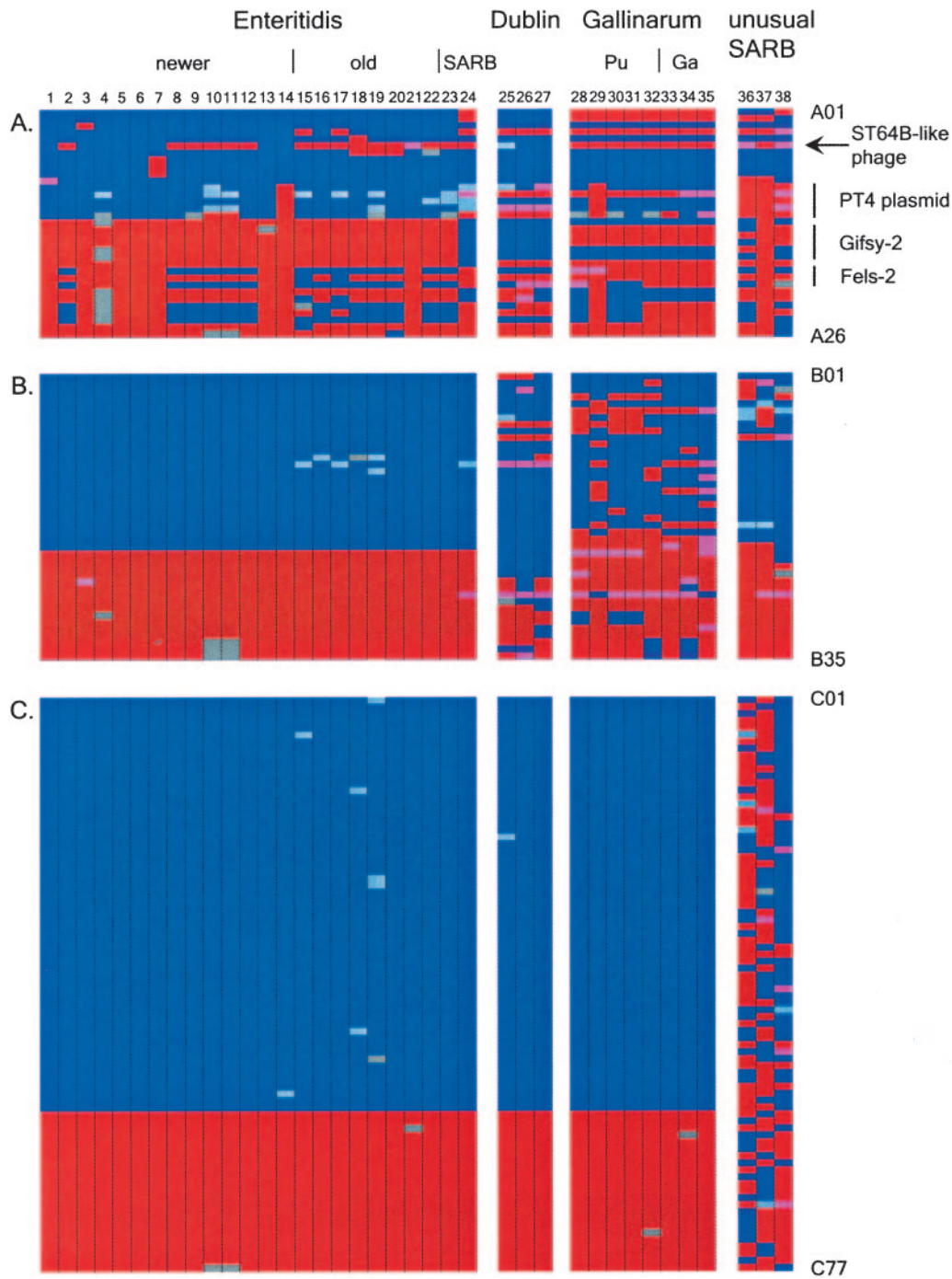


FIG. 1. Genetic regions of diversity in serovar Enteritidis, Dublin, and Gallinarum isolates compared to serovar Enteritidis PT4. (A) Regions that distinguish serovar Enteritidis isolates. (B) Additional regions of difference detected in serovar Dublin and Gallinarum isolates. (C) Further regions of difference detected in unusual SARB isolates. Presence status is color coded: red, absent; light red, mostly absent; gray, uncertain; light blue, mostly present; blue, present. Strain designations correspond to the numbers in Table 1. Regions are described in detail in Table 2 (panels A and B) and Table S2 in the supplemental material (panel C).

sequence that were apparently present in one or more of the old isolates (Table 2). Three of the phage regions (A19, A22, and A24) are present in all old isolates except the PT4b representative. Region A19 has similarity to Typhimurium LT2 Fels-2, and regions A22 and A24 are parts of the Typhi CT18 P2-like phages ST27 and ST35, respectively (36). These areas

are also found in the unfinished genome sequence of the serovar Enteritidis isolate partially sequenced at the University of Illinois (data not shown), a PT8 strain originally isolated from a chicken (14). The old isolates representing phage types 13a and 8 all contained another region of ST35, region A23. In addition, one old PT13a isolate harbors a phage with high

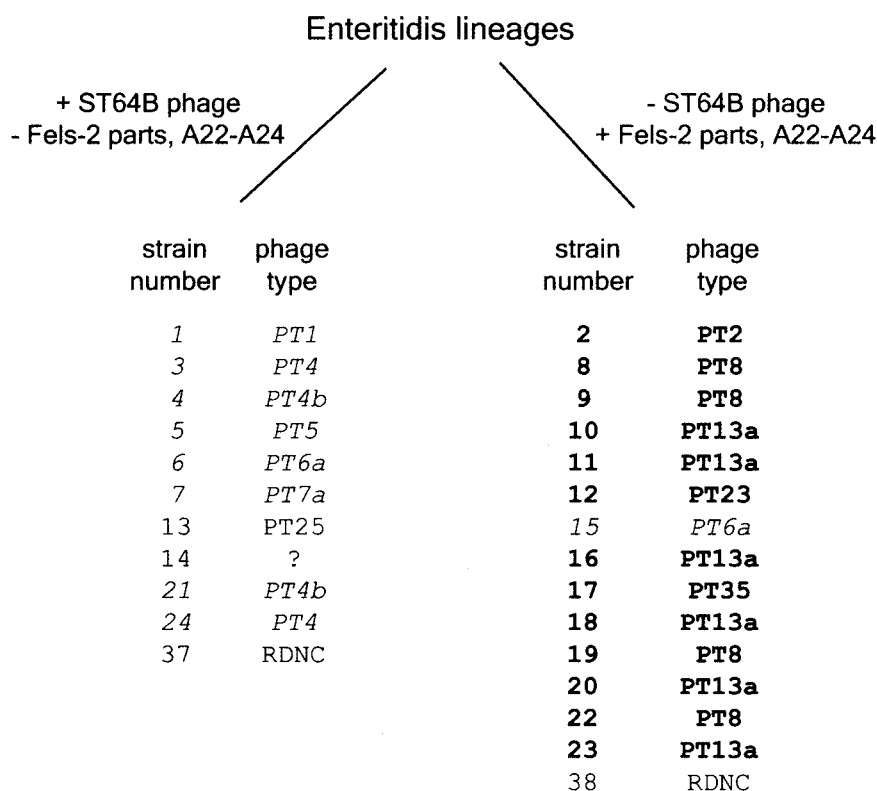


FIG. 2. Lineages of serovar Enteritidis strains based on presence of a subset of phage regions. Isolates in this study were grouped based on absence/presence patterns of regions A06 (ST64B-like phage), A19 (Fels-2 regions), and the Typhi CT18 phage regions A22 to A24. Deviation of one character status was allowed. The strains in italics are phage types of the PT4-like lineage, and strains in boldface are PT8-like phage types, according to Guard-Petter (9). Strain numbers correspond to Table 1.

similarity to the 25.4-kb Paratyphi A SARB42 phage SPA-3 (region A26). Finally, a Typhi CT18-derived region of 4.2 kb, which is part of the P2-like phage SopE_{ST} (36), was found in the PT6a old strain (A25).

Gene contents of more recent serovar Enteritidis isolates. We investigated the gene contents of 13 serovar Enteritidis strains that were more recently collected, the majority of which were isolated in Denmark and Spain between 1983 and 1994 (Table 1). These isolates represented 10 phage types, including the most prevalent types in Europe and the United States (PT4, -8, and -13a). The two recent PT13a strains included in this data set have been characterized in a previous study (21). The recent isolates of phage types 1, 4, 4b, 5, 6a, 7a, and 25 contained genes homologous to the ST64B-like phage present in the sequenced PT4. However, the four remaining phage types represented by six recent strains in this study, PT2, -8, -13a, and -23, lacked this phage.

Notably, these four phage types contained the same non-PT4 phage regions as all of the old isolates except the PT4b representative (regions A19, A22, and A24) (see above). Region A23, present in the old PT8 and PT13a isolates, was also detected in these six recent strains. Applying the five phage regions A06, A19, and A22 to A24 for separation of the investigated strains, with two differences considered a new lineage, two major groups can be distinguished—those that lack A06 and contain the others and those that contain A06 and lack the others. Figure 2 outlines the clusters obtained and their almost

perfect overlap with the separation into PT4-like and PT8-like lineages based on LPS core structure, as proposed by Guard-Petter (9).

Three genomic PT4 regions that are present in all genomic *Salmonella* sequences to date are missing in the recent PT7a isolate: regions A08 to A10. The 12.6 kb of region A08 encompass the entire hydrogenase 3 operon of *S. enterica* serovar Typhimurium LT2 (15 genes), which enables the bacterium to produce hydrogen from endogenously produced formate in cooperation with the formate hydrogenlyase enzyme (34). Region A09 encodes three cell invasion proteins, SipB, SipC, and SipD, as well as the chaperone SicA, and lies within *Salmonella* pathogenicity island 1. SipB, SipC, and SicA are necessary for *Salmonella* invasion of eukaryotic cells (13). In addition, SipB activates caspase 1 to induce apoptosis of the mammalian macrophage (10) and results in release of interleukin-18 (8). The third region missing in the PT7a isolate (A10) is the biggest segment found to be missing in any of the serovar Enteritidis strains investigated (more than 50 kb). It contains 43 genes that include two two-component regulatory systems, *pmrAB* and *dcuRS*, as well as an anaerobic dimethyl sulfoxide reductase complex, a formate-dependent nitrite reductase (*nrf* operon), genes involved in melibiose transport and utilization (*melABR*), and the central metabolic enzyme acetyl-coenzyme A synthetase *Acs*. The *pmrAB* regulatory system confers resistance to a variety of antibacterial peptides, including polymyxin, and is also involved in lipid A modification (35). In

conclusion, the lack of these regions in the investigated PT7a isolate could render the strain nonvirulent, if not nonviable, under environmental conditions. The described deletions might have occurred after isolation of the strain from a human host in 1983.

The strain MZ672, an isolate from Germany, did not display any differences from the sequenced PT4 strain except the notable lack of the entire virulence plasmid. Since this plasmid is known to enhance the virulence of serovar Enteritidis isolates, it can be speculated that the strain may have lost this plasmid after its initial isolation.

Genome content of common SARB serovar Enteritidis. The four serovar Enteritidis isolates included in the SARB collection (4) were also subjected to microarray analysis. Two of the four MLEE types of serovar Enteritidis represented in this collection were found only in a singular isolate (out of at least 362 serovar Enteritidis strains surveyed), and their genome contents were found to be very dissimilar to those of the remaining serovar Enteritidis strains in this study. These differences are described in detail below. The other two more common isolates were SARB16, representing the major MLEE type (357 isolates), and SARB18, representing a minor type (3 isolates).

SARB16, a PT13a strain (12), displayed a genome that was essentially identical to those of the recent PT2, PT8, PT13a, and PT23 isolates included in this study. The minor SARB isolate SARB18, a PT4 strain isolated approximately 25 years ago in Connecticut, lacked the PT4 phage regions A04 and A06, providing further evidence that the sequenced PT4 isolate may have acquired the ST64B-like phage fairly recently, and possibly in Europe. Lack of the ST64B phage did not coincide with presence of the Fels-2 and serovar Typhi phage regions as in the isolates of the PT2/8/13a/23 cluster. SARB18 is also devoid of the plasmid-derived *pef* fimbriae. These and all other deviations from PT4 are summarized in Table 2.

Differences found in closely related serovars, but not serovar Enteritidis. In addition to a survey of old and recent serovar Enteritidis isolates, the gene contents of three strains of the related *S. enterica* serovar Dublin and nine isolates of serovar Gallinarum were investigated. Many of the regions found to be divergent in the serovar Enteritidis isolates were also different in the investigated serovar Dublin, biovar Pullorum, and biovar Gallinarum isolates (Table 2 shows the details). Of the nine serovar Gallinarum strains, six were biovar Pullorum and three were biovar Gallinarum. Two of the biovar Pullorum isolates were represented in the SARB collection, with the rare variant SARB52 displaying an aberrant genome content. This isolate is discussed below, together with the unusual serovar Enteritidis SARB isolates.

The three serovar Dublin isolates included in this study had a total of six chromosomal PT4 regions of divergence that were universally present in all of the serovar Enteritidis isolates tested (Table 2). Three of these, regions B09, B13_a, and sub_B07, were absent in all three of the strains. The products of B09 are possibly involved in starvation sensing and are present in all common serovar Enteritidis and Gallinarum isolates investigated here. Region B13_a contains several genes that may be of phage origin and has been shown to be missing in other isolates of *Salmonella*, including strains from serovars Dublin, Montevideo, Choleraesuis, Abortusovis, Typhi, Typhisuis, and Paratyphi A, B, and C (26). This segment is also absent in all biovar Gallinarum isolates. Sub_B07 is a small region similar to a segment of phage Fels-1.

Nine non-PT4 regions were detected in the serovar Dublin isolates but not in any serovar Enteritidis isolates in this panel. Among these is region B24, an area of the *S. enterica* serovar Typhimurium LT2 genome between STM0268 and STM0275, which includes a gene with similarities to a Shiga-like toxin. As previously noted (26), the Dublin SARB13 isolate contained almost the entire Typhi CT18 SPI7, including the Vi antigen but excluding phage SopE_{ST}. Remarkably, Dublin SARB12 contained all genes of the *Escherichia coli* plasmid R46 that were present on our array.

The isolates of serovar Gallinarum, including both biovars Gallinarum and Pullorum, contributed a total of 19 deletions of regions that were present in all of the serovar Enteritidis or serovar Dublin strains investigated. Two of these regions were absent in all serovar Gallinarum strains included in the panel: region B06, 8 kb of phage origin with similarity to the STM1004-to-STM1056 region, including *sph2*, and region B23, comprised of 4.4 kb and found in its entirety only in serovar Dublin and serovar Enteritidis genome sequences to date.

The array experiments identified three PT4 regions that were absent in biovar Pullorum strains (except in the atypical strains 29 and 36) but were present in all other field isolates of serovar Enteritidis, serovar Dublin, or biovar Gallinarum. These were regions B22, B08, and B04. Region B22 is a 2-kb island encoding part of the *tor* regulatory system. This particular genomic area has been found to be missing only in one serovar Abortusovis isolate to date (26) but was ubiquitous in all other *Salmonella* isolates. The *torRS* locus encodes a two-component regulator for the bacterial trimethylamine *N*-oxide reductase respiratory system, which may also aid in protection of the bacterium against high pH (3). Similarly, the 10.8 kb of region B08 was also missing in that serovar Abortusovis isolate only and was present in every other *Salmonella* isolate investigated to date. Finally, region B04, a 1.5-kb island containing a hydrolase, has been found to be absent in many different serovars of *Salmonella* (26) and is therefore a highly mobile, and dispensable, area in the genome.

The biovar Pullorum isolate SARB51 apparently lacked a homolog of the *rpoS* gene (in region B20), which should render the strain less resistant to stress in stationary phase. Mutations in *rpoS* have been observed quite frequently in clinical serovar Typhi isolates but not in clinical serovar Typhimurium (33). However, *rpoS* mutations were found commonly in highly passaged *Salmonella* strains. Strains that cause systemic disease only, like serovar Typhi, are known to have a high natural mutation frequency of *rpoS*. Since biovar Pullorum isolates do not cause systemic disease exclusively but also result in diarrhea, the loss of this region may have occurred after initial isolation of SARB51.

Two regions are missing in all three biovar Gallinarum strains included in Table 2 but are present in all other strains investigated here. Region B21 is a 12.3-kb cluster that encodes *std* fimbriae. The assortment of fimbriae is known to be very irregular and specific for different serovars within *Salmonella*, but the *std* locus has so far been observed to be absent only in *Salmonella* outside subspecies I (26). The other region absent

from all three serovar Gallinarum strains is region B16. Its gene products may play a role in chemotaxis, since STM1265, one of the two regulatory genes predicted to be within this region, has a *cheY*-like receiver domain. This is the first report of the absence of this region in any *Salmonella* isolate.

Strain 29, a recent biovar Pullorum isolate from Canada, differed quite substantially from the other biovar Pullorum isolates investigated. It harbored regions B07, B08, and B22, which are all missing in the other common biovar Pullorum strains. Moreover, it lacked regions B16 and B21, deletions otherwise found only in strains of biovar Gallinarum. Based on the genetic profile, it may be more closely related to biovar Gallinarum than to biovar Pullorum.

Unusual serovar Enteritidis and biovar Pullorum isolates in the SARB collection. This study also included investigations of some SARB serovars that were very rare at the time of their isolation and/or exhibited gene contents that were quite different from the genomic reservoir present in the other isolates of the same serovar. These included strains SARB17 and SARB20, both originally serotyped as serovar Enteritidis, and SARB52, serotyped as serovar Gallinarum biovar Pullorum. The presence/absence results of the regions in these three “unusual” strains can be found as part of Table S2 in the supplemental material. Overall, 77 regions were identified with unique differences within these three isolates compared to Enteritidis PT4.

In particular, isolate SARB17 exhibited a very unusual genome. Some elements that were previously defined as subspecies I specific were found to be present, but some elements that had been originally characterized as subspecies I predictive (29) were apparently absent. There are currently 31 predicted genes characteristic of subspecies I and invariably present in subspecies I isolates (see Table S3 in the supplemental material), three of which were found to be absent in SARB17: STM0058 and STM0062, involved in citrate lyase synthesis, and STM2516, *sinI*, a gene with similarity to a sporulation inhibition gene in *Bacillus*. Overall, this isolate lacked approximately 363 kb of the entire PT4 chromosome. Some of these regions, amounting to at least 155 kb, were absent only in SARB17 and no other isolate within the panel of strains presented here. In phylogenetic trees using whole-genome absence/presence data based on serovar Typhimurium LT2 comparisons with approximately 200 strains, including 79 isolates representing the 25 most prevalent serovars, SARB17 clustered consistently outside subspecies I. However, it was also separated from the isolates of all other subspecies (data not shown).

SARB20, the other non-phage-typeable rare MLEE type of serovar Enteritidis included in the SARB collection, had less dramatic differences from PT4. It followed the PT2/8/13a/23 pattern of phage gene presence but also lacked any plasmid-derived genes. It contained seven unique differences from the PT4 sequence that had not been observed in any other serovar Enteritidis isolate. These included the absence of the complete *ste* fimbria locus, a sugar kinase locus, and the *rtc* locus.

Another unusual strain, the biovar Pullorum isolate SARB52, lacked approximately 263 kb of PT4 regions, 75 kb of which were uniquely missing in this isolate and no other strain in this study. Apart from the absence of the *stc* fimbriae, a cluster that is also missing in serovar Typhimurium and serovar

Typhi, no functions could be assigned to many of the genes in these regions. Some putative roles included heat shock resistance (*hsc*), O-antigen transport, ABC transport, dehydrogenases, and racemases. SARB52 contains the LT2 *stj* fimbriae and the *sta* fimbriae found in Typhi CT18, both of which are absent in Enteritidis PT4.

DISCUSSION

Among the recent serovar Enteritidis isolates investigated here, members of the PT8-like clade were found to harbor a specific set of phage genes, which were similar to phage elements occurring in serovars Typhimurium and Typhi. This phage presence pattern is probably the molecular expression of the distinction between the two phage clusters in the current serovar Enteritidis phage-typing scheme. However, the presence of the detected phage may not be the cause of the lineage distinction. A different LPS core structure in these PT8-like phage types may have rendered the isolates more susceptible to lysogenic phage that were detected with our array.

When old *S. enteritidis* strains, isolated before the eradication of serovar Gallinarum in domestic fowl flocks, were compared with newer isolates, including strains isolated from humans and chickens, no significant consistent differences in gene content were observed. However, differences on a minor scale, like transcriptional changes due to point mutations, silencing of genes, and small deletions, were not monitored using our whole-gene array. Further, the loss of genetic segments may have contributed to the ability to move into fowl. These deletions could not be monitored on the array, since it is based on the genome sequence of a recent PT4 strain. The information obtained by the comparative genomic hybridization experiments is by definition unidirectional—we can monitor only genes that are present in the sequenced strain, but not those specific to other isolates of the same serovar. Changes in the status of genes not present in the sequenced isolate will not be detected by our array. However, it is entirely possible that the loss of competition from serovar Gallinarum strains alone may have enabled serovar Enteritidis to infect the domestic chicken flocks (2, 31).

The serovar Enteritidis strains investigated here displayed remarkable chromosomal homogeneity on whole-gene level, given the different phage types, isolation years, geographical locations, and hosts they were recovered from. This is consistent with a possible recent, and clonal, serovar history. However, two very rare serovar Enteritidis isolates represented in the SARB collection showed major differences, with one isolate exhibiting a genome very aberrant from the PT4 sequence. This isolate may be an example of how the genes affecting serotyping, i.e., the LPS biosynthetic and flagellum genes, may be transferred into a different *Salmonella* isolate with a distant phylogenetic background, resulting in serovar assignments that are unrelated to possible pathogenicity and specificity of the isolate in question. It is intriguing that SARB17 appears to have characteristics intermediate between subspecies I and all other subspecies of *Salmonella*. In phylogenetic trees based on genomic content using absence/presence predictions of genes, this isolate clusters between subspecies I and its closest related lineage, subspecies VI (data not shown), an observation confirmed by recent multilocus sequence typing analyses (M.

Achtman, personal communication). The subspecies assignment, and consequently the serovar, of this strain is uncertain.

Two regions were identified in this study that were missing in all common biovar Pullorum isolates and only one other isolate in the extensive studies across the entire *Salmonella* clade: a similarly host-specific serovar, Abortusovis. One of these missing regions contained the *tor* regulatory system, and the other region has a possible function in invasion of eukaryotic cells. Two other distinct regions were identified to be specifically missing in the three biovar Gallinarum isolates included in this study, both of which are probably involved in motility, chemotaxis, or adherence traits of the bacterium: the *std* fimbriae and a 6.6-kb region (B16) consisting of genes of unknown function that is present in all other isolates of *Salmonella* investigated to date. The one biovar Pullorum strain (other than the unusual SARB52) that contained *tor* and lacked *std* and region B16, strain 29, was later confirmed to be of unusual phenotype, not conforming to the established biovar Pullorum infectivity. This strain had been isolated together with strain 28 in the same outbreak. It is currently classified as “intermediate” biovar Pullorum (J. Guard-Petter, personal communication). Therefore, the observed genetic differences may be predictive of the different biovars. Other genes that are specifically present in either biovar Gallinarum or biovar Pullorum isolates may be elucidated soon, since both biovars are awaiting completion of their genome sequences. It will be interesting to determine whether the number of genomic differences between the two very similar biovars of serovar Gallinarum will be small enough to allow correlation of the observed phenotypic differences between the biovars with the genomic dissimilarities between the two lineages.

The whole-genome microarray was able to differentiate between the two major phage type lineages within serovar Enteritidis based on the comparatively few phage genes represented on the array. It was not successful in discriminating between all different phage types of serovar Enteritidis. In principle, phage type in serovar Enteritidis, based on a profile of phage that can infect the bacterial isolate and shown to be dependent on LPS structure (9), may also generally have genetic manifestations in the presence and absence of lysogenic phages in the respective genomes. An array specifically designed to detect phage gene patterns would have more discriminatory power than our present array. The costs of microarray analysis are constantly decreasing, making it possible that a more complete “phage array” might be able to type *Salmonella* strains without having to do the conventional, cost-effective but cumbersome, lysis experiments. In addition, an inexpensive PCR-based approach to classification within and between serovars can be envisioned, also based on the presence and absence of sets of specific chromosomal genes, cistrons, and phage identified in this study.

ACKNOWLEDGMENTS

We are grateful for the policy of the Wellcome Trust Sanger Institute to release its sequencing data into the public domain as the data are being generated. These sequence data were produced by the *Salmonella* sp. comparative sequencing group at the Sanger Institute and can be obtained from <ftp://ftp.sanger.ac.uk/pub/pathogens/Salmonella/SePT4.dbs>. We thank Ken Sanderson (University of Calgary, Calgary, Alberta, Canada), Jean Guard-Bouldin (USDA Agricultural Research Service, Athens, Ga.), and Patti Fields (Centers for Disease Control

and Prevention) for strains and critical reading of the manuscript. In addition, we acknowledge Jonathan Frye, Allan Helm, Javier Garaizar, and Timothy Wallis for sending us isolates for this study.

Financial support came from NIH grant AI34829 to M.M. and the generosity of Sidney Kimmel.

REFERENCES

- Barrow, P. A., J. M. Simpson, M. A. Lovell, and M. M. Binns. 1987. Contribution of *Salmonella gallinarum* large plasmid toward virulence in fowl typhoid. *Infect. Immun.* **55**:388–392.
- Baumler, A. J., B. M. Hargis, and R. M. Tsois. 2000. Tracing the origins of *Salmonella* outbreaks. *Science* **287**:50–52.
- Bordi, C., L. Theraulaz, V. Mejean, and C. Jourlin-Castelli. 2003. Anticipating an alkaline stress through the Tor phosphorelay system in *Escherichia coli*. *Mol. Microbiol.* **48**:211–223.
- Boyd, E. F., F. S. Wang, P. Beltran, S. A. Plock, K. Nelson, and R. K. Selander. 1993. *Salmonella* reference collection B (SARB): strains of 37 serovars of subspecies I. *J. Gen. Microbiol.* **139**:1125–1132.
- Bullis, K. L. 1977. The history of avian medicine in the U.S. II. Pullorum disease and fowl typhoid. *Avian Dis.* **21**:422–429.
- Delcher, A. L., D. Harmon, S. Kasif, O. White, and S. L. Salzberg. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res.* **27**:4636–4641.
- Deng, W., S. R. Liou, G. Plunkett III, G. F. Mayhew, D. J. Rose, V. Burland, V. Kodoyianni, D. C. Schwartz, and F. R. Blattner. 2003. Comparative genomics of *Salmonella enterica* serovar Typhi strains Ty2 and CT18. *J. Bacteriol.* **185**:2330–2337.
- Dreher, D., M. Kok, C. Obregon, S. G. Kiama, P. Gehr, and L. P. Nicod. 2002. *Salmonella* virulence factor SipB induces activation and release of IL-18 in human dendritic cells. *J. Leukoc. Biol.* **72**:743–751.
- Guard-Petter, J. 1999. Phage type and other outer-membrane characteristics of *Salmonella enterica* serovar Enteritidis associated with virulence, p. 221–232. *In* A. M. Saeed, R. K. Gast, M. E. Potter, and P. G. Wall (ed.), *Salmonella enterica* serovar Enteritidis in humans and animals, vol. 1. Iowa State University Press, Ames.
- Hernandez, L. D., M. Pypaert, R. A. Flavell, and J. E. Galan. 2003. A *Salmonella* protein causes macrophage cell death by inducing autophagy. *J. Cell Biol.* **163**:1123–1131.
- Hickman-Brenner, F. W., A. D. Stubbs, and J. J. Farmer III. 1991. Phage typing of *Salmonella enteritidis* in the United States. *J. Clin. Microbiol.* **29**:2817–2823.
- Hudson, C. R., M. Garcia, R. K. Gast, and J. J. Maurer. 2001. Determination of close genetic relatedness of the major *Salmonella enteritidis* phage types by pulsed-field gel electrophoresis and DNA sequence analysis of several *Salmonella* virulence genes. *Avian Dis.* **45**:875–886.
- Kaniga, K., S. Tucker, D. Trollinger, and J. E. Galan. 1995. Homologs of the *Shigella* IpaB and IpaC invasins are required for *Salmonella typhimurium* entry into cultured epithelial cells. *J. Bacteriol.* **177**:3965–3971.
- Keller, L. H., D. M. Schifferli, C. E. Benson, S. Aslam, and R. J. Eckroade. 1997. Invasion of chicken reproductive tissues and forming eggs is not unique to *Salmonella enteritidis*. *Avian Dis.* **41**:535–539.
- Liebana, E., C. Clouting, L. Garcia-Migura, F. A. Clifton-Hadley, E. Lindsay, E. J. Threlfall, and R. H. Davies. 2004. Multiple genetic typing of *Salmonella enteritidis* phage-types 4, 6, 7, 8 and 13a isolates from animals and humans in the UK. *Vet. Microbiol.* **100**:189–195.
- Liebana, E., L. Garcia-Migura, M. F. Breslin, R. H. Davies, and M. J. Woodward. 2001. Diversity of strains of *Salmonella enterica* serotype Enteritidis from English poultry farms assessed by multiple genetic fingerprinting. *J. Clin. Microbiol.* **39**:154–161.
- Liebana, E., L. Garcia-Migura, J. Guard-Petter, S. W. McDowell, S. Rankin, H. M. Opitz, F. A. Clifton-Hadley, and R. H. Davies. 2002. *Salmonella enterica* serotype Enteritidis phage types 4, 7, 6, 8, 13a, 29 and 34: a comparative analysis of genomic fingerprints from geographically distant isolates. *J. Appl. Microbiol.* **92**:196–209.
- Ling, M. L., and G. C. Wang. 2001. Epidemiological analysis of *Salmonella enteritidis* isolates in Singapore. *J. Infect.* **43**:169–172.
- McClelland, M., K. E. Sanderson, S. W. Clifton, P. Latreille, S. Porwollik, A. Sabo, R. Meyer, T. Bieri, P. Ozersky, M. McLellan, C. R. Harkins, C. Wang, C. Nguyen, A. Berghoff, G. Elliott, S. Kohlberg, C. Strong, F. Du, J. Carter, C. Kremizki, D. Layman, S. Leonard, H. Sun, L. Fulton, W. Nash, T. Miner, P. Minx, K. Delehaunty, C. Fronick, V. Magrini, M. Nhan, W. Warren, L. Florea, J. Spieth, and R. K. Wilson. 2004. Comparison of genome degradation in Paratyphi A and Typhi, human-restricted serovars of *Salmonella enterica* that cause typhoid. *Nat. Genet.* **36**:1268–1274.
- McClelland, M., K. E. Sanderson, J. Spieth, S. W. Clifton, P. Latreille, L. Courtney, S. Porwollik, J. Ali, M. Dante, F. Du, S. Hou, D. Layman, S. Leonard, C. Nguyen, K. Scott, A. Holmes, N. Grewal, E. Mulvaney, E. Ryan, H. Sun, L. Florea, W. Miller, T. Stoneking, M. Nhan, R. Waterston, and R. K. Wilson. 2001. Complete genome sequence of *Salmonella enterica* serovar Typhimurium LT2. *Nature* **413**:852–856.
- Morales, C. A., S. Porwollik, J. G. Frye, H. Kinde, M. McClelland, and J.

- Guard-Bouldin.** 2005. Correlation of phenotype with the genotype of egg-contaminating *Salmonella enteritidis*. *Appl. Environ. Microbiol.*, in press.
22. **Mumma, G. A., P. M. Griffin, M. I. Meltzer, C. R. Braden, and R. V. Tauxe.** 2004. Egg quality assurance programs and egg-associated *Salmonella enteritidis* infections, United States. *Emerg. Infect. Dis.* **10**:1782–1789.
 23. **Parker, C. T., B. Harmon, and J. Guard-Petter.** 2002. Mitigation of avian reproductive tract function by *Salmonella enteritidis* producing high-molecular-mass lipopolysaccharide. *Environ. Microbiol.* **4**:538–545.
 24. **Parkhill, J., G. Dougan, K. D. James, N. R. Thomson, D. Pickard, J. Wain, C. Churcher, K. L. Mungall, S. D. Bentley, M. T. Holden, M. Sebahia, S. Baker, D. Basham, K. Brooks, T. Chillingworth, P. Connor, A. Cronin, P. Davis, R. M. Davies, L. Dowd, N. White, J. Farrar, T. Feltwell, N. Hamlin, A. Haque, T. T. Hien, S. Holroyd, K. Jagels, A. Krogh, T. S. Larsen, S. Leather, S. Moule, P. O'Gaora, C. Parry, M. Quail, K. Rutherford, M. Simmonds, J. Skelton, K. Stevens, S. Whitehead, and B. G. Barrell.** 2001. Complete genome sequence of a multiple drug resistant *Salmonella enterica* serovar Typhi CT18. *Nature* **413**:848–852.
 25. **Patrick, M. E., P. M. Adcock, T. M. Gomez, S. F. Altekruse, B. H. Holland, R. V. Tauxe, and D. L. Swerdlow.** 2004. *Salmonella enteritidis* infections, United States, 1985–1999. *Emerg. Infect. Dis.* **10**:1–7.
 26. **Porwollik, S., E. F. Boyd, C. Choy, P. Cheng, L. Florea, E. Proctor, and M. McClelland.** 2004. Characterization of *Salmonella enterica* subspecies I genovars by use of microarrays. *J. Bacteriol.* **186**:5883–5898.
 27. **Porwollik, S., J. Frye, L. D. Florea, F. Blackmer, and M. McClelland.** 2003. A non-redundant microarray of genes for two related bacteria. *Nucleic Acids Res.* **31**:1869–1876.
 28. **Porwollik, S., and M. McClelland.** 2003. Lateral gene transfer in *Salmonella*. *Microbes Infect.* **5**:977–989.
 29. **Porwollik, S., R. M. Wong, and M. McClelland.** 2002. Evolutionary genomics of *Salmonella*: gene acquisitions revealed by microarray analysis. *Proc. Natl. Acad. Sci. USA* **99**:8956–8961.
 30. **Porwollik, S., R. M. Wong, S. H. Sims, R. M. Schaaper, D. M. DeMarini, and M. McClelland.** 2001. The DeltauvrB mutations in the Ames strains of *Salmonella* span 15 to 119 genes. *Mutat. Res.* **483**:1–11.
 31. **Rabsch, W., B. M. Hargis, R. M. Tsois, R. A. Kingsley, K. H. Hinz, H. Tschape, and A. J. Baumler.** 2000. Competitive exclusion of *Salmonella enteritidis* by *Salmonella gallinarum* in poultry. *Emerg. Infect. Dis.* **6**:443–448.
 32. **Ridley, A. M., E. J. Threlfall, and B. Rowe.** 1998. Genotypic characterization of *Salmonella enteritidis* phage types by plasmid analysis, ribotyping, and pulsed-field gel electrophoresis. *J. Clin. Microbiol.* **36**:2314–2321.
 33. **Robbe-Saule, V., G. Algorta, I. Rouilhac, and F. Norel.** 2003. Characterization of the RpoS status of clinical isolates of *Salmonella enterica*. *Appl. Environ. Microbiol.* **69**:4352–4358.
 34. **Sawers, R. G., D. J. Jamieson, C. F. Higgins, and D. H. Boxer.** 1986. Characterization and physiological roles of membrane-bound hydrogenase isoenzymes from *Salmonella typhimurium*. *J. Bacteriol.* **168**:398–404.
 35. **Tamayo, R., S. S. Ryan, A. J. McCoy, and J. S. Gunn.** 2002. Identification and genetic characterization of PmrA-regulated genes and genes involved in polymyxin B resistance in *Salmonella enterica* serovar Typhimurium. *Infect. Immun.* **70**:6770–6778.
 36. **Thomson, N., S. Baker, D. Pickard, M. Fookes, M. Anjum, N. Hamlin, J. Wain, D. House, Z. Bhutta, K. Chan, S. Falkow, J. Parkhill, M. Woodward, A. Ivens, and G. Dougan.** 2004. The role of prophage-like elements in the diversity of *Salmonella enterica* serovars. *J. Mol. Biol.* **339**:279–300.
 37. **van Duijkere, E., W. J. Wannet, D. J. Houwers, and W. van Pelt.** 2002. Serotype and phage type distribution of *Salmonella* strains isolated from humans, cattle, pigs, and chickens in the Netherlands from 1984 to 2001. *J. Clin. Microbiol.* **40**:3980–3985.
 38. **Ward, L. R., J. D. de Sa, and B. Rowe.** 1987. A phage-typing scheme for *Salmonella enteritidis*. *Epidemiol. Infect.* **99**:291–294.
 39. **Ward, L. R., J. Threlfall, H. R. Smith, S. J. O'Brien, H. Riemann, P. Kass, D. Cliver, A. J. Baumler, B. M. Hargis, and R. M. Tsois.** 2000. *Salmonella enteritidis* epidemic. *Science* **287**:1753c.