

The Iron Control Element, Acting in Positive and Negative Control of Iron-Regulated *Bradyrhizobium japonicum* Genes, Is a Target for the Irr Protein†

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Bradyrhizobium japonicum, the nitrogen-fixing soybean symbiont, possesses a heme uptake system encoded by the gene cluster *hmuVUT-hmuR-ebxBD-tonB*. Transcription of the divergently oriented *hmuT* and *hmuR* genes was previously found to be induced by iron limitation and to depend on a 21-bp promoter-upstream iron control element (ICE). Here, we show by deletion analysis that the full-length ICE is needed for this type of positive control. Additional genes associated with ICE-like motifs were identified in the *B. japonicum* genome, of which *bll6680* and *blr7895* code for bacterioferritin and rubrerythrin homologs, respectively. Transcription start site mapping revealed that their ICEs directly overlap with either the –10 promoter region or the transcription initiation site, suggesting an involvement of the ICE in negative control of both genes. Consistent with this inference was the observed down-regulation of both genes under iron limitation, which in the case of *bll6680* was shown to require an intact ICE motif. Using a yeast one-hybrid system, we demonstrated *in vivo* interaction of the iron response regulator (Irr) with all three ICEs. Moreover, specific *in vitro* binding of purified Irr protein to the ICE motifs of *bll6680* and *blr7895* was shown in electrophoretic mobility shift experiments. A genome-wide survey for iron-regulated genes with a custom-made Affymetrix gene chip revealed 17 genes to be induced and 68 to be repressed under iron-replete conditions. Remarkably, ICE-like motifs are associated with a large subset of those *B. japonicum* genes. We propose the ICE as an important *cis*-acting element in *B. japonicum* which represents the DNA-binding site for the Irr protein and, depending on its location within promoter regions, is involved in positive or negative control of the associated iron-regulated genes.

Genes involved in iron acquisition or metabolism are not only conserved within and between many bacterial taxa but are also under tight control in response to the external iron supply. Iron availability is usually restricted because the element—even when abundantly present in the environment—occurs mostly in its insoluble, ferric form (2, 4, 21). To get access to this metal as a nutrient, microorganisms have evolved different strategies. Iron-chelating molecules (siderophores), synthesized under iron-limiting conditions, are secreted to bind to ferric iron (Fe³⁺), and the Fe-siderophore complexes are then imported via active transport mechanisms into the cell (11, 60). Alternative iron sources are also used, including heme (both as a free molecule or protein bound, as in hemoglobins), Fe-containing proteins, ferric citrate, or inorganic iron (ideally in the ferrous form). Once the intracellular iron concentration is sufficiently high, transcription of genes that are involved in Fe uptake (e.g., those for siderophore synthesis and transport) is shut off.

After its first identification in *Escherichia coli* (27), the ferric uptake regulator Fur was thought to be the sole “global” regulator for all processes concerned with iron acquisition and

metabolism. In its repressive, Fe²⁺-liganded form, the Fur protein binds to the so-called Fur boxes on DNA, thus preventing transcription of the iron-regulated genes. Although Fur homologs are now known to exist in a vast range of different bacterial species (26), most studies on Fur have been carried out with γ -proteobacteria such as *E. coli*, *Vibrio cholerae*, or *Pseudomonas aeruginosa* (47, 48). Recently, *E. coli* Fur was demonstrated to act also as an indirect activator in that it represses the transcription of a small regulatory RNA, *ryhB*, which in turn inhibits expression of several target genes (38). Unexpectedly, *Neisseria meningitidis* Fur appears to activate RNA transcription directly by binding to upstream sequences of the *pan1*, *norB*, and *nuoA* genes in response to iron accumulation in the cell (9).

Further Fe-dependent regulators have been discovered. One is DtxR of *Corynebacterium diphtheriae*, a repressor of diphtheria toxin production (6), which in gram-positive bacteria such as *Bacillus subtilis* assumes some of the roles that Fur plays in other bacterial species (26). RirA is an iron-responsive regulator in the symbiotic bacterium *Rhizobium leguminosarum*. Null mutation of *rirA* causes high-level expression of at least eight operons whose products are involved in synthesis and uptake of siderophores, uptake of heme, and iron acquisition from other sources (57). The zinc uptake regulator Zur of *Bacillus subtilis* (15) and the manganese uptake regulator Mur of *R. leguminosarum*, which represses transcription of the *sitABCD* operon (10, 15), differ from other members of the Fur

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family, as they do not respond solely to Fe^{2+} but rather to Zn^{2+} or Mn^{2+} .

Another member of the Fur family of transcriptional regulators is the iron-response regulator Irr, first identified in the nitrogen-fixing soybean symbiont, *Bradyrhizobium japonicum* (23). At present, Irr occurrence appears to be restricted to α -proteobacteria, including rhizobia, *Agrobacterium tumefaciens*, the animal pathogen *Brucella abortus*, and *Rhodospseudomonas palustris* (62). It has been argued particularly for the rhizobia that they have a high demand for iron due to the intracellular abundance of iron-sulfur proteins and hemoproteins (nitrogenase complex and cytochromes) during symbiotic nitrogen fixation (42, 20). The Irr protein was shown to repress transcription of *hemB*, the gene for δ -aminolevulinic acid dehydratase in the heme biosynthesis pathway (23). Interestingly, Irr itself is modified at the posttranslational level: under iron-replete conditions, Irr interacts directly with ferrochelatase and becomes degraded in a process that depends on heme being delivered by the ferrochelatase enzyme (49, 50). In addition, transcription of the *irr* gene is repressed in Fe-replete conditions by Fur, even though the *irr* promoter region shares no sequence similarity with canonical Fur boxes (25). Since other roles of Irr, apart from regulating *hemB*, were so far unknown, a DNA sequence motif for Irr binding had yet to be defined.

B. japonicum and *R. leguminosarum* can grow on heme as the sole iron source (43, 46, 63), a property that until recently was thought to be unique to pathogenic bacteria (16). The heme uptake system (Hmu) of *B. japonicum* consists of a TonB-dependent heme receptor (HmuR), a periplasmic heme-binding protein (HmuT), an ABC transporter (HmuUV), and a Ton system (ExbBD and TonB). Transcription of *hmuT* and *hmuR* was shown to be up-regulated by iron limitation via a mechanism that involves a 21-bp A/T-rich incomplete inverted repeat named the iron control element (ICE) (43). This motif is located in between the divergently oriented *hmuT* and *hmuR* genes and might represent the binding site for a regulatory protein that, presumably, acts positively on the expression of both genes (43).

Here, we have followed up on this previous work and have addressed several open questions. Are both of the half-sites of the symmetric ICE needed for control? Does one of the known *B. japonicum* iron control proteins, Irr or Fur, bind to the ICE? Can we solidify the postulated positive control at this ICE? The experiments reported here provide answers to these questions. Moreover, we describe the use of a microarray for the identification of iron-regulated *B. japonicum* genes on a genome-wide scale. Many of those genes are associated with ICE-like motifs which are involved in positive or negative control, depending on their location within promoters.

MATERIALS AND METHODS

Strains and plasmids. Strains and plasmids used in this work are listed in Table 1.

Media and growth conditions. *E. coli* was grown in Luria-Bertani (LB) medium (40) containing the following concentrations of antibiotics for plasmid selection ($\mu\text{g ml}^{-1}$): ampicillin, 200; kanamycin, 30; and tetracycline, 10. *B. japonicum* was cultivated in peptone-salts-yeast extract (PSY) medium (51) containing 0.1% L-arabinose and 1.2 μM FeCl_3 for routine use. Concentrations of antibiotics were as follows ($\mu\text{g ml}^{-1}$): spectinomycin, 100; kanamycin, 100; streptomycin, 50; and tetracycline, 50 (solid media) and 25 (liquid media). For growth under "low-iron" conditions, PSY medium lacked the iron supplement. PSY medium representing "high-iron" conditions contained 50 μM FeSO_4 . Yeast

strains were grown at 30°C in synthetic dropout medium or yeast extract-peptone-dextrose medium (yeast protocols handbook; Clontech, Palo Alto, CA).

DNA work and sequence analysis. Recombinant DNA work was performed according to standard protocols (53). *B. japonicum* chromosomal DNA was isolated as described previously (22). Programs DNA Star MegAlign 5.06, Clone Manager 7.02, GeneDoc version 2.6.002 (<http://www.psc.edu/biomed/genedoc>), and WebLogo version 2.8 (<http://weblogo.berkeley.edu/>) were used for computer-assisted analyses of DNA and protein sequences. Similarity searches were done on the National Center for Biotechnology Information BLAST network server (<http://www.ncbi.nlm.nih.gov/BLAST/>) and RhizoBase, the database for rhizobial genomes (<http://www.kazusa.or.jp/rhizobase/>). Designations for *B. japonicum* genes and open reading frames were also taken from RhizoBase (blr, bll, bsr, and bsl plus 4-digit numbers).

RNA isolation, cDNA synthesis, and microarray analysis. Cultures of *B. japonicum* 110*spc4* were grown to an optical density (at 600 nm) of 0.35 to 0.4 in PSY medium either lacking or containing 50 μM FeSO_4 iron. Grown cultures (200 ml) were immediately transferred into cold tubes containing 1/10 of the culture volume of stop solution (10% Tris-HCl-buffered phenol [pH 8] in ethanol). After centrifugation for 5 min (10,800 \times g; 4°C) the supernatant was decanted, and cells were frozen in liquid nitrogen and stored at -80°C . Isolation of total RNA, cDNA synthesis, and conditions for microarray hybridization were described elsewhere (28). Based on the *B. japonicum* genome sequence (31), a custom Affymetrix-type gene chip representing all 8,317 annotated open reading frames plus 51 RNA genes and intergenic regions was designed (Affymetrix Inc., Santa Clara, CA). Further details of the BJAPETHa520090 microarray will be described elsewhere, and they are also available on request from the authors. Primary data analysis was done with the Affymetrix GeneChip operating software (GCOS). GeneSpring 4.2 software (Silicon Genetics) was used for comparison analysis.

In silico search for ICE motifs. Initial searches for ICE-like motifs in intergenic regions of the *B. japonicum* genome were performed with the program "FUZZNUC" (<http://emboss.ch.emblnet.org/Pise>) using TTTA-N_g-TAAA as a query sequence. In subsequent searches, the Darwin software (19) was used. The ICE motif was represented as a position-specific frequency matrix (PSFM), which provides a good approximation of protein-DNA interactions (5). The PSFM is a matrix consisting of the frequency of each nucleotide at each motif position, based on a collection of known motifs. \sqrt{N} pseudocounts (where N is the number of motifs in the PSFM) were used to enable finding new motifs that were similar, but not identical, to the known motifs (32). The genomic sequence 500 bp upstream of each annotated gene was searched for the motif PSFM. Because transcriptional start sites were not known for most genes, the translational start codon was used as a substitute. Each (overlapping) sequence of the same length as the profile was scored by multiplying the frequencies from the PSFM: $\prod_{i=1}^n \text{PSFM}[i,s(i)]$, where n is the length of the PSFM and s is the sequence (of length n) to be scored.

Construction of transcriptional *hmuR-lacZ* fusions. Starting from a 1.8-kb DNA fragment comprising the entire *hmuT-hmuR* intergenic region (plasmid pRJ8710), a set of nested deletion derivatives with variable 5' ends relative to *hmuR* was generated by PCR amplification using *Taq* DNA polymerase and primer pairs GR-3/GR-7, GR-4/GR-7, and GR-5/GR-7 (Table 2). Amplification products of 408 bp, 391 bp, and 375 bp were digested with EcoRI and BamHI, cloned into the pUC19 vector, and verified by sequencing. Plasmid-derived EcoRI-BamHI fragments of 394 bp, 377 bp, and 361 bp were then inserted between the EcoRI and BamHI sites of the transcriptional *lacZ* fusion vector pME3535XhoI, from which the entire transcriptional *lacZ* fusions, including the variable upstream promoter fragments, were transferred into the broad-host-range vector pRK290X by cloning in between the EcoRI and XhoI restriction sites. The final constructs pRJ8722, pRJ8723, and pRJ8724 (Fig. 1) were mobilized by conjugation into *B. japonicum* strains 110*spc4* and GEM4. The correct plasmid contents of these strains were verified by PCR.

Cloning of genes bl16680 and blr7895. *B. japonicum* chromosomal regions comprising bl16680 and blr7895 were amplified by PCR using *Taq* DNA polymerase and primer pairs 6680-1/6680-2 and 7895-1/7895-2 (Table 2). The resulting PCR products of 1,301 bp (bl16680) and 1,306 bp (blr7895) were cloned into vector pGEM-T Easy, yielding plasmids pRJ2562 and pRJ2563. Cloned DNA regions were verified by sequencing.

Construction of translational bl16680'-*lacZ* fusions. A translational bl16680'-*lacZ* fusion was obtained by isolating a 726-bp EcoRI-ScaI fragment from plasmid pRJ2562 and cloning it between the EcoRI and SmaI sites of pNM482. The resulting plasmid, pRJ8767, carried the 5' end of bl16680 and its promoter region, including ICE₆₆₈₀. In parallel, a mutated version of the translational bl16680'-*lacZ* fusion was constructed in which the nucleotides of the 3' portion of ICE₆₆₈₀ were systematically replaced. This resulted in the alteration of the

TABLE 1. Bacterial and yeast strains, cloning vectors, and recombinant plasmids used in this study

Strain or plasmid	Relevant genotype or properties	Source or reference
Strains		
<i>E. coli</i>		
DH5 α	<i>supE44 ΔlacU169 (φ80lacZΔM15) hsdR17 recA1 gyrA96 thi-1 relA1</i>	Bethesda Research Laboratories Inc., Gaithersburg, MD
S17-1	Sm ^r Sp ^r <i>hsdR</i> (RP4-2 <i>kan::Tn7 tet::Mu</i> , integrated in the chromosome)	54
BL21(DE3)	<i>hsdS gal (λCIts857 ind1 Sam7 nin5 lacUV5-T7 gene 1)</i>	55
<i>B. japonicum</i>		
110spc4	Sp ^r wild type	51
GEM4	Sp ^r <i>fur::Ω</i> (parental strain: USDA I110)	24
<i>S. cerevisiae</i>		
YM4271	<i>MATA ura3-52 his3-200 ade2-101 lys2-801 leu2-3,112 trp1-901 tyr1-501 gal4-Δ512 gal80-Δ538 ade5::hisG</i>	36
8730	ICE _{hmu} chromosomally integrated in YM4271	This work
8732	ICE _{hmu} mut chromosomally integrated in YM4271	This work
8733	ICE ₆₆₈₀ chromosomally integrated in YM4271	This work
8734	ICE ₇₈₉₅ chromosomally integrated in YM4271	This work
8765	Fur box chromosomally integrated in YM4271	This work
Plasmids		
pGEM-T Easy	Ap ^r TA cloning vector	Promega, Madison, WI
pME3535XhoI	Tc ^r pME3535 derivative with XhoI site (Klenow fill-in of StuI site, addition of XhoI linker)	64
pASK-IBA3+	Ap ^r expression vector, Tet repressor, <i>tetA</i> -promoter/operator, C-terminal <i>Strep</i> -tag II	IBA, Göttingen, Germany
pRK290X	Tc ^r pRK290 derivative with XhoI site	1
pRKPolScaI	Tc ^r pRK290 derivative with ScaI linker	Laboratory collection
pNM482	Ap ^r pUC8 ' <i>lacZ</i>	41
pSUP482	Tc ^r ' <i>lacZ</i> part from pNM482 in pSUP202pol4	Laboratory collection
pUC18/19	Ap ^r high-copy-number cloning vectors	45
pGAD424	Ap ^r expressing the GAL4 activation domain (AD)	Clontech, Palo Alto, CA
pLacZi	Ap ^r for integration at the <i>URA3</i> locus; carries the <i>lacZ</i> reporter gene	Clontech, Palo Alto, CA
pRJ2562	Ap ^r (pGEM-T Easy) 1,301-bp PCR fragment comprising bll6680 and upstream region	This work
pRJ2563	Ap ^r (pGEM-T Easy) 1,306-bp PCR fragment comprising blr7895 and upstream region	This work
pRJ8710	Ap ^r (pUC18) with 1,852-bp EcoRI/BamHI fragment comprising the <i>hmuT-hmuR</i> intergenic region and <i>hmuR</i>	This work
pRJ8722	Tc ^r (pRK290X) with 5,538-bp <i>hmuR-lacZ</i> fusion	This work
pRJ8723	Tc ^r (pRK290X) with 5,521-bp <i>hmuR-lacZ</i> fusion	This work
pRJ8724	Tc ^r (pRK290X) with 5,505-bp <i>hmuR-lacZ</i> fusion	This work
pRJ8730	Ap ^r (pLacZi) triple tandem copy of ICE _{hmu}	This work
pRJ8732	Ap ^r (pLacZi) triple tandem copy of ICE _{hmu} mut	This work
pRJ8733	Ap ^r (pLacZi) triple tandem copy of ICE ₆₆₈₀	This work
pRJ8734	Ap ^r (pLacZi) triple tandem copy of ICE ₇₈₉₅	This work
pRJ8735	Ap ^r (pGAD424) 493-bp EcoRI fragment comprising the <i>fur</i> gene	This work
pRJ8737	Ap ^r (pGAD424) 504-bp SmaI fragment comprising the <i>irr</i> gene	This work
pRJ8750	Ap ^r (pASK-IBA3+) 491-bp Eco31I fragment comprising the <i>irr</i> gene	This work
pRJ8765	Ap ^r (pLacZi) triple tandem copy of Fur box	This work
pRJ8766	Ap ^r (pGEM-T Easy) 1,017-bp PCR fragment comprising bll6680 and upstream region with 1/2-ICE ₆₆₈₀	This work
pRJ8767	Ap ^r (pNM482) 726-bp EcoRI/ScaI fragment of pRJ2562 comprising bll6680'- <i>lacZ</i> and upstream region	This work
pRJ8768	Ap ^r (pNM482) 730-bp EcoRI/ScaI fragment of pRJ8766 comprising bll6680'- <i>lacZ</i> and upstream region with 1/2-ICE ₆₆₈₀	This work
pRJ8769	Tc ^r (pRKPolScaI) 5,829-bp EcoRI/StuI fragment of pRJ8767	This work
pRJ8770	Tc ^r (pRKPolScaI) 5,829-bp EcoRI/StuI fragment of pRJ8768	This work

wild-type ICE₆₆₈₀ (ATTAGAAAGCGTTCTAAAT) to the mutated 1/2-ICE₆₆₈₀ (ATTAGAAAGCCAAGATTAA; nucleotides in italics are part of an AseI site introduced in the course of the mutagenesis). Using plasmid pRJ2562 as a template, two overlapping fragments comprising the desired mutations were generated in separate amplification reactions with *Pfu* DNA polymerase and primer pairs SP6/6680-19 and 6680-18/F2 (Table 2). Products of these reactions were mixed, annealed via their overlapping ends, and subjected to a third PCR step using the external primers SP6 and F2 according to the method of Higuchi

et al. (29). After attachment of 3' A overhangs in an additional reaction with *Taq* polymerase and dATP, the final amplification product of 1,017 bp, including 1/2-ICE₆₆₈₀ and the 5' portion of bll6680, was cloned into vector pGEM-T Easy, yielding plasmid pRJ8766. The cloned DNA region was verified by sequencing. A translational bll6680'-*lacZ* fusion containing the mutated 1/2-ICE₆₆₈₀ was obtained by cloning a 726-bp EcoRI-ScaI fragment from pRJ8766 between the EcoRI and SmaI sites of pNM482, resulting in plasmid pRJ8768. Finally, 5,829-bp EcoRI-StuI fragments of pRJ8767 and pRJ8768, comprising the desired

TABLE 2. Oligonucleotides used in this work

Designation	Nucleotide sequence (5' to 3') ^{a,b}
GR-3	CCGGAATTCCGGCGAAATTTACAATCGATATAAACTGC _{7,793,570}
GR-4	CCGGAATTCGATATAAACTGCAACCAGACGTC _{7,793,559}
GR-5	CCGGAATTCAGACGTCATTGACCGTCAG _{7,793,547}
GR-7	GCGCGGGTAGATCTCAGCGCTTCT _{822,883}
GR-14	CGGAATTCATGACCGCACTGAAACCTT _{856,706}
GR-15	CGGAATTCGCGAGTCGAAGATGATGAGA _{856,257}
GR-21	TCCCCGGGAATGAGCGAGAATACCG _{823,348}
GR-22	TCCCCGGGTAGATCTCAGCGCTTCT _{822,883}
GR-26	ATGGTAGTCTCAAATGAGCGAGAATACCGCGCCCAT _{823,340}
GR-27	ATGGTAGTCTCAGCGCTGCGCTTCTGCGCAGGCGCA _{822,894}
fwICE-2	AATTC-(AATTTACAATCGATATAAACT) _{3x} -G
revICE-2	TCGAC-(AGTTTATATCGATTGTAATTT) _{3x} -G
5' ICE E/S _{mut}	AATTC-(AATTTACAATCGCGATATAAACT) _{3x} -G
3' ICE E/S _{mut}	TCGAC-(AGTTTATATCGCGATTGTAATTT) _{3x} -G
furbox fw	AATTC-(GATAATGATAATCATTATC) _{3x} -G
furbox rev	TCGAC-(GATAATGATAATCATTATC) _{3x} -G
6680-1	GGGAATTCAGCTGTTCCAGCAGCGATAG _{7,350,980}
6680-2	GGGAATTCACAAGCAGCAGCTGTAGGC _{7,349,730}
6680-7	GTGCCITGTTGAGGTAGTCGATGA _{7,350,328}
6680-8	GTACTGGTTGATCGCAGTCAGCTC _{7,350,296}
6680-9	AATTC-(ATTTAGAAGCGTTCTAAAT) _{3x} -G
6680-10	TCGAC-(ATTTAGAAGCGTTCTAAAT) _{3x} -G
6680-11 ^c	aaactcATTTAGAAGCGTTCTAAATcggtt _{7,350,384}
6680-mut1 ^{c,d}	aaactcATTTAGAAGCGCGTTCTAAATcggtt
6680-16 ^c	aaactcATTTAGAAGCCAAGATTAATcggtt
6680-18	ATTTAGAAGCCAAGATTAATCGGTTTAGATCGAGTTGGACCG
6680-19	CCTAAACCGATTAATCTTGGCTTCTAAATGAGTTTACCG
F2	ATGATCTCCTTGACGTTCT
SP6	ATTTAGGTGACACTATAG
hmu-1 ^c	ggcggaaAATTTACAATCGATATAAACTgcaacc _{7,793,566}
hmu-mut1 ^{c,d}	ggcggaaAATTTACAATCGCGATATAAACTgcaacc
7895-1	GGGAATTCGTTTCGAGGCGATGCGAAAAG _{8,656,690}
7895-2	GGGAATTCGATCGCGATTGATATGTTGGATG _{8,657,942}
7895-8	GAAGTCAITGATGCGGCTGT _{8,656,963}
7895-9	AATTC-(AATTTAGAATCATTCTAAACT) _{3x} -G
7895-10	TCGAC-(AGTTTAGAATGATTCTAAAT) _{3x} -G
7895-11	GCTCTTCTCGGCCATCTGTT _{8,657,032}
GS-7895-11 ^c	ctatatAATTTAGAATCATTCTAAACTagtggg _{8,656,884}
7895-mut1 ^{c,d}	ctatatAATTTAGAATCGCATTCTAAACTagtggg

^a Nucleotide positions in the *B. japonicum* genome are indicated by subscript numbers at the 3' ends of oligonucleotides. 3× indicates that the sequence in parentheses occurs three times in a row.

^b Nucleotides shown in italics belong to restriction sites.

^c Capital letters refer to the 19-to-21-bp ICE motifs of the respective promoter regions; lowercase letters represent adjacent nucleotides (Fig. 2).

^d The spacing between the half-sites of the mutated ICE motifs is increased by the two underlined nucleotides.

lacZ fusions, were cloned between the EcoRI and ScaI restriction sites of the broad-host-range vector pRKPolScaI to yield plasmids pRJ8769 (wild-type ICE₆₆₈₀) and pRJ8770 (1/2-ICE₆₆₈₀). These plasmids were mobilized by conjugation into *B. japonicum* strain 110*spc4*.

Transcript mapping. The transcription start sites of bl16680 and blr7895 were determined by primer extension. *B. japonicum* wild-type cells were grown aerobically to mid-exponential phase in PSY medium either lacking iron or supplemented with 50 μM FeSO₄. RNA was isolated and used for reverse transcription as described previously (3, 44), using primers 6680-7 and 6680-8 (for bl16680) and 7895-8 and 7895-11 (for blr7895). Extension products were analyzed on 6% denaturing polyacrylamide gels adjacent to sequencing ladders generated with the same oligonucleotides and plasmids pRJ2562 and pRJ2563.

Construction of an Irr-*Strep*-tag expression plasmid. The *B. japonicum* 110*spc4* chromosomal region comprising the *irr* gene (bl10768) was amplified by PCR using *Pfu* DNA polymerase and primer pair GR-26/GR-27 (Table 2) containing Eco3II restriction sites (520-bp PCR product). 3' A overhangs were attached through an additional treatment with *Taq* polymerase, which then enabled cloning into the pGEM-T Easy vector. The construct was verified by sequencing. Cloning of the pGEM-T Easy vector-derived 491-bp Eco3II *irr* fragment into the Eco3II-digested expression vector pASK-IBA3+ (encoding the C-terminal *Strep*-tag II; IBA, Göttingen, Germany) resulted in the *irr* expression plasmid pRJ8750.

Overproduction and purification of recombinant Irr protein. *E. coli* strain BL21(DE3) harboring plasmid pRJ8750 was used for overproduction of the Irr-*Strep*-tag II fusion protein. One-liter LB medium cultures in 5,000-ml Erlenmeyer flasks were grown on a rotary shaker (160 rpm) at 37°C until an optical density (at 550 nm) of 0.4 to 0.6 was reached. Expression of *irr* was induced by adding 0.2 μg ml⁻¹ anhydrotetracycline, and the cultures were further incubated for 2 to 3 h. Cells were harvested by centrifugation (5 min, 8,000 × g, 4°C), resuspended in 3 ml of buffer W (100 mM Tris-HCl [pH 8.0], 150 mM NaCl), and lysed by three passages through a French pressure cell (11,000 lb in⁻²). The extract was centrifuged for 30 min (16,000 × g, 4°C), and the supernatant with soluble recombinant Irr protein was applied to a 1-ml *Strep*-Tactin column (IBA, Göttingen, Germany) and affinity purified according to the instructions in the manufacturer's protocol. A further purification step was added by anion ion-exchange chromatography using a 10-ml Mono Q column (4.6 by 100 mm; Amersham Biosciences). The column was developed with 10 mM Tris-HCl (pH 8), 1 mM MgCl₂, 1 mM dithiothreitol, and 5% glycerol using a salt gradient from 0.04 M to 1 M KCl. For desalting, the fraction containing the Irr-*Strep*-tag II fusion protein was gel filtrated over a PD-10 Sephadex G-25 column (Amersham Biosciences) with DNA-binding buffer (10 mM Tris-HCl (pH 8), 40 mM KCl, 1 mM MgCl₂, 1 mM dithiothreitol, 5% glycerol). Protein concentration was determined with the Bio-Rad assay (Bio-Rad Laboratories, Richmond, CA) and the degree of purity was checked by sodium dodecyl sulfate-polyacrylamide gel electrophoresis.

Electrophoretic mobility shift assay. Electrophoretic mobility shift assays were used to monitor Irr binding to the ICE. Irr protein and double-stranded, end-labeled ICE oligonucleotides were incubated for 10 to 15 min at room temperature in a 20-μl volume of DNA-binding buffer to which 1 μg poly(dI-dC) (Amersham Biosciences) was added. Double-stranded ICE DNA probes were

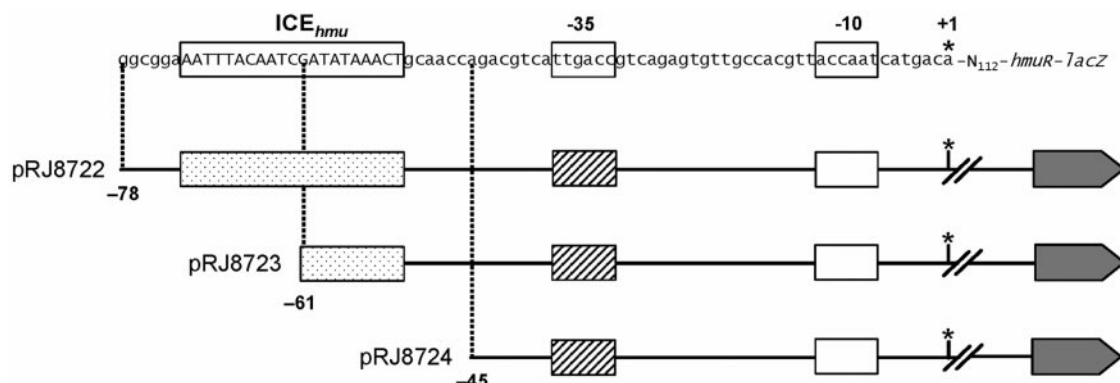


FIG. 1. Promoter-upstream deletions of transcriptional *hmuR-lacZ* fusions. The top line shows the nucleotide sequence of the *hmuR* promoter region comprising the -10/-35 region and the iron regulatory element (ICE_{*hmu*}). The three lower lines represent different 5' deletion constructs. Plasmid names and nucleotide positions of the deletion end points are indicated to the left of each construct. An asterisk marks the transcription start at genome position 7,793,520. The arrow on the far right symbolizes *lacZ*.

TABLE 3. ICE-dependent Fe regulation of plasmid-borne *hmuR-lacZ* fusions in a *B. japonicum* wild-type and *fur* background

Plasmid	ICE _{hmu} genotype	<i>hmuR-lacZ</i> expression ^a							
		110 <i>spc4</i> (wt)			GEM4 (<i>fur</i> mutant)			Ratio (<i>fur</i> mutant:wt)	
		Miller units		Ratio (low:high Fe)	Miller units		Ratio (low:high Fe)	High Fe	Low Fe
		High Fe	Low Fe		High Fe	Low Fe			
pRJ8722	Wild type	51 ± 5	521 ± 67	10.2	26 ± 11	1,045 ± 93	40.2	0.5	2.0
pRJ8723	Deleted 5' half-site	27 ± 1	89 ± 10	3.3	15 ± 8	182 ± 7	12.1	0.6	2.0
pRJ8724	Complete deletion	23 ± 2	69 ± 9	3.0	11 ± 2	83 ± 9	7.5	0.5	1.2

^a Two aerobic cultures per strain and condition were inoculated to an optical density (at 600 nm) of 0.02 and grown for 3 days at 30°C in PSY medium without supplemented FeSO₄ (low Fe) or supplemented with 50 μM FeSO₄ (high Fe) and assayed in duplicate. Mean values ± standard errors of β-galactosidase activities (Miller units) are shown for one out of three independent experiments, which all gave similar results. wt, wild type.

produced by annealing the single-stranded oligonucleotides representing the ICE motifs with their corresponding complementary oligonucleotides (oligonucleotide *hmu*-1 for ICE_{hmu}; 6680-11 for ICE₆₆₈₀; 6680-16 for 1/2-ICE₆₆₈₀; GS-7895-11 for ICE₇₈₉₅; *hmu*-mut1, 6680-mut1, and 7895-mut1 for 2-bp insertions in ICE_{hmu}, ICE₆₆₈₀, and ICE₇₈₉₅, respectively) (Table 2). End labeling and annealing of oligonucleotides was performed as follows: oligonucleotides (30 pmol), T4 polynucleotide kinase (10 units; MBI Fermentas, Vilnius, Lithuania) and [³²P]ATP (10 μCi, 3,000 Ci/mmol) were incubated in polynucleotide kinase-buffer A (MBI Fermentas) for 30 min at 37°C in a total volume of 50 μl. After addition of 2 μl stop solution (4 M Na-acetate, pH 7.5, 25 mM EDTA), 60 pmol of the unlabeled complementary oligonucleotide was added. After 10 min at 95°C, the reaction mixture was slowly cooled down to room temperature. Annealed, labeled oligonucleotides were purified over a NAP-10 Sephadex G-25 column (Amersham Biosciences). Following incubation of protein and DNA (see above), 5 μl of loading dye solution (30% glycerol, 0.02% bromophenol blue) was added and reaction mixtures were analyzed on 6% nondenaturing polyacrylamide gels in electrophoresis buffer (90 mM Tris base, 90 mM boric acid, pH 8) that were prerun for 10 to 20 min at 180 V. After electrophoresis for 25 to 30 min at 180 V, gels were dried and exposed on a phosphorimager screen.

Construction of yeast one-hybrid plasmids and reporter strains. Target DNAs were created by first annealing complementary oligonucleotides (Table 2) representing three tandem copies of the respective ICE (primer pair fwICE-2/revICE-2 for ICE_{hmu}; primer pair 6680-9/6680-10 for ICE₆₆₈₀; primer pair 7895-9/7895-10 for ICE₇₈₉₅; and, as control, primer pair 5'ICE E/S_{-mut}/3'ICE E/S_{mut} for ICE_{hmu}mut or the consensus Fur box (primer pair furbox fw/furbox rev). Annealed oligonucleotides carrying overhanging EcoRI and SalI ends were inserted into vector pLacZi (Clontech) between the EcoRI and SalI sites upstream of the GAL4-dependent *lacZ* reporter gene, resulting in constructs pRJ8730 (ICE_{hmu}), pRJ8732 (ICE_{hmu}mut), pRJ8733 (ICE₆₆₈₀), pRJ8734 (ICE₇₈₉₅), and pRJ8765 (Fur box). Correct orientation and sequence of the ICEs or Fur box were confirmed by DNA sequencing. ICE- or Fur box-containing plasmids were linearized by digestion with NcoI and integrated into the genome of yeast strain YM4271 upon transformation according to the lithium acetate method (17). The five resulting yeast strains are listed in Table 1. For the construction of translational fusions between the activation domain of yeast GAL4 and *B. japonicum* prey proteins, *B. japonicum fur* (bl0797) and *irr* (bl0768) were PCR amplified with primer pairs GR-14/GR-15 and GR-21/GR-22, respectively. Genes were cloned in frame with the GAL4 activation domain-encoding region on the EcoRI- or SmaI-linearized cloning vector pGAD424 (Matchmaker one-hybrid system; Clontech), yielding plasmids pRJ8735 and pRJ8737 that were confirmed by sequencing. These plasmids were transferred to all five reporter yeast strains (Table 1) by lithium acetate transformation (17).

β-Galactosidase assays. Assays for β-galactosidase activity in *B. japonicum* were carried out as described previously (12). β-Galactosidase activity assays with *Saccharomyces cerevisiae* were performed as described in the user manual for the Matchmaker one-hybrid system (Clontech).

RESULTS

Maximal *hmuR* expression depends on full-length ICE_{hmu}. The ICE is located in the promoter regions between the divergently oriented *hmuT* and *hmuR* genes and is necessary for low-iron-inducible expression of these genes. In order to determine the length of the ICE needed for control of *hmuR*, it

was functionally characterized by 5' deletion analysis. A set of transcriptional *hmuR-lacZ* fusions with variable 5' ends relative to *hmuR* was generated (see Materials and Methods and Fig. 1). Expression from the plasmid-borne *hmuR-lacZ* fusions was monitored as β-galactosidase activity measured in *B. japonicum* wild-type (110*spc4*) and *fur* mutant (GEM4) cells grown under high- and low-iron conditions (Table 3). The latter strain was included to investigate a potential role of the Fur protein in iron regulation of *hmuR*.

Maximal expression was observed under iron-depleted conditions in both *B. japonicum* backgrounds harboring pRJ8722 with full-length ICE_{hmu}. Deletion of the 5' ICE half-site (pRJ8723) resulted in a sixfold decrease of expression in both backgrounds. Complete elimination of the ICE (pRJ8724) further decreased *hmuR-lacZ* expression, maximally twofold. Thus, maximal *hmuR* activation under iron-depleted conditions depended on full-length ICE_{hmu}. Under iron-replete conditions, expression of *hmuR* was repressed in both backgrounds, with slightly higher residual expression levels when full-length ICE was present (pRJ8722).

Because the regulatory pattern of *hmuR* expression was largely the same in wild-type and *fur* mutant cells, the Fur protein is unlikely to be directly involved. Yet, Fur may contribute indirectly (via control of *irr*; see Discussion), because gene activation factors in the *fur* background were about fourfold higher than in the wild type when assayed with pRJ8722 and pRJ8723.

The fact that significant iron regulation was retained in both backgrounds, even when ICE_{hmu} was completely deleted, may point to the existence of an (unknown) additional regulatory mechanism which is independent of the ICE and Fur.

***B. japonicum* possesses several ICE-like sequences.** A systematic search was performed to look for the possible presence of additional ICEs in the *B. japonicum* strain 110 genome. Indeed, using the TTTA-N₆-TAAA motif as a probe with the "FUZZNUC" program, six additional ICE-like motifs were identified, four of them located in presumed intergenic regions upstream of blr0586 (ICE₀₅₈₆), blr4157 (ICE₄₁₅₇), blr6680 (ICE₆₆₈₀), and blr7895 (ICE₇₈₉₅) (Fig. 2). The open reading frames blr0586 and blr4157 code for conserved hypothetical proteins without significant similarities to functionally characterized proteins. They were not further analyzed in this study. By contrast, ICE₆₆₈₀ and ICE₇₈₉₅ are associated with genes that code for proteins likely to contain iron. The predicted product of blr6680 (162 amino acids) shares a 50% identity (68% similarity) with bacterioferritin from *E. coli*. The

TABLE 4. Expression of ICE-*lacZ* and Fur box-*lacZ* reporter fusions in *S. cerevisiae* strains synthesizing Gal4'-Irr or Gal4'-Fur hybrid prey proteins^a

Yeast strain	Target	β-Galactosidase activity (Miller units)	
		Gal4'-Irr	Gal4'-Fur
8730	ICE _{hmu}	36.1 ± 9.7	0.0
8732	ICE _{hmu} mut	0.0	ND
8733	ICE ₆₆₈₀	10.1 ± 1.3	0.0
8734	ICE ₇₈₉₅	93.5 ± 8.4	0.0
8765	Fur box	ND	5.3 ± 0.3

^a Cells of yeast reporter strains were transfected with prey plasmid pRJ8737 (Gal4'-Irr) or pRJ8735 (Gal4'-Fur). Cultures grew in synthetic dropout medium (-URA, -LEU) for 3 to 5 h until they reached mid-exponential phase (optical density at 600 nm of 0.5 to 0.8), and one-hybrid interactions were examined as β-galactosidase activity. Activity was measured from two cultures per strain, each assayed in duplicate. Control yeast strains 8730, 8733, 8734 and 8765 harboring the vector pGAD424 (encoding Gal4' only) showed no β-galactosidase activity. Data are shown for one out of four independent experiments, all yielding similar results. ND, not done.

were candidate regulators for ICE-dependent iron control. Based on the genetic evidence presented above (Table 3), Fur is unlikely to interact with the ICE. Unfortunately, we were unable to apply the same strategy for testing the potential involvement of Irr, because repeated attempts to construct a null mutation in the respective gene (*blI0768*) of our *B. japonicum* wild-type strain 110*spc4* were not successful. Therefore, a yeast one-hybrid system, employing Irr- and Fur-expressing yeast strains, was used to examine whether any of these regulatory proteins might be able to interact with the ICE directly. Yeast reporter strains were constructed as described in Material and Methods. Reproducible one-hybrid interaction was observed between Irr and all three of the ICES associated with *hmuR*, *blI6680*, and *blr7895* (Table 4). To validate that interaction between Irr and the three ICES was specific, a negative control strain carrying a mutated ICE_{hmu} with a 2-bp insertion between the ICE half-sites was used (ICE_{hmu}mut). No interaction between Irr and the mutant ICE was detected (Table 4). Similarly, no interaction was observed when Gal4'-Fur (pRJ8735) or Gal4' alone (pGAD424) was expressed in the ICE-containing reporter strains. When pRJ8735 was present in the yeast strain 8765 which contains an *E. coli* Fur box upstream of the *lacZ* reporter gene, β-galactosidase activity was detected, indicating that the Gal4'-Fur hybrid protein was functional.

Irr protein binds to ICE₆₆₈₀ and ICE₇₈₉₅ in an electrophoretic mobility shift assay. *Strep*-tagged Irr protein was overproduced, purified (see Materials and Methods), and tested for possible binding to the ICE in gel retardation experiments. While Irr binding to ICE_{hmu} was very weak and not reproducibly detected (data not shown), we observed clear Irr-dependent retardation in the cases of ICE₆₆₈₀ and ICE₇₈₉₅ (Fig. 4A). To test whether Irr binds specifically to these two ICE motifs, competitive gel retardation experiments were done. Non-labeled, wild-type ICE₆₆₈₀ (31 bp) and ICE₇₈₉₅ (33 bp) were used as competing DNAs, whereas two variants, termed ICE₆₆₈₀* (33 bp) and ICE₇₈₉₅* (35 bp), were assumed to be noncompetitive. ICE₆₆₈₀* and ICE₇₈₉₅* are derivatives of ICE₆₆₈₀ and ICE₇₈₉₅, respectively, which contain two additional nucleotides (GC) inserted between the ICE half-sites. The same mutation inserted in ICE_{hmu} was shown previously

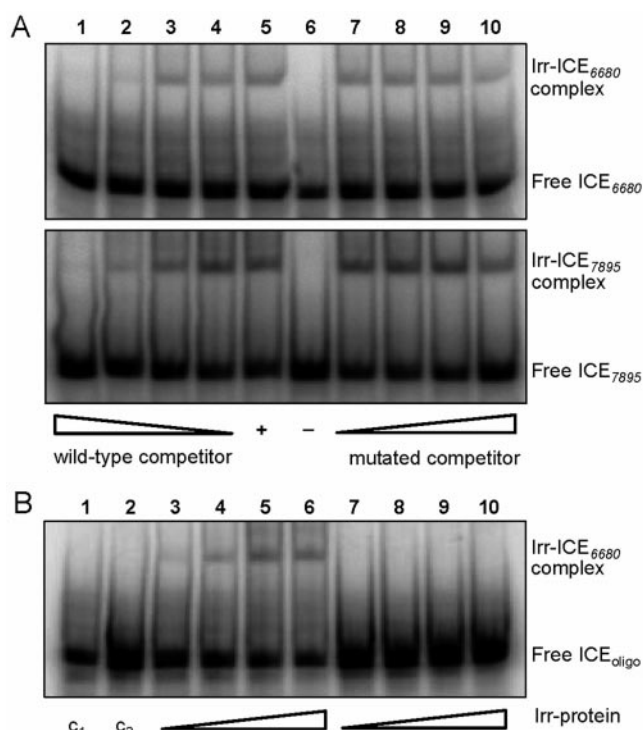


FIG. 4. Analysis of Irr binding to ICE targets by electrophoretic mobility shift assays. (A) Constant amounts of purified, recombinant Irr protein (1.5 μM) and double-stranded ³²P-labeled oligonucleotides ICE₆₆₈₀ (upper) and ICE₇₈₉₅ (lower), each at approximately 2 nM, were incubated with increasing amounts of the corresponding nonlabeled wild-type (ICE₆₆₈₀ and ICE₇₈₉₅) or mutated (ICE₆₆₈₀* and ICE₇₈₉₅*) competitor oligonucleotides. Molar ratios between labeled and nonlabeled oligonucleotides were 1:1 (lanes 4 and 7), 1:3 (lanes 3 and 8), 1:30 (lanes 2 and 9), and 1:300 (lanes 1 and 10). No competitor oligonucleotide and no Irr protein was added to the control reactions in lanes 5 and 6, respectively. (B) Increasing amounts of purified, recombinant Irr protein were incubated with constant amounts of double-stranded ³²P-labeled oligonucleotide ICE₆₆₈₀ (lanes 3 through 6) or 1/2-ICE₆₆₈₀ (lanes 7 through 10), each at approximately 2 nM. Concentrations of Irr protein were 0.26 μM (lanes 3 and 7), 0.52 μM (lanes 4 and 8), 1.0 μM (lanes 5 and 9), and 2.1 μM (lanes 6 and 10). No Irr protein was added to the control reactions C₁ and C₂ with oligonucleotides ICE₆₆₈₀ (lane 1) and 1/2-ICE₆₆₈₀ (2), respectively. Samples were run on 6% non-denaturing polyacrylamide gels and visualized by exposing the dried gels to a phosphorimager screen. ICE_{oligo}, free digonucleotide ICE₆₆₈₀ or 1/2-ICE₆₆₈₀.

to abolish activation of *hmuR* and *hmuT* under iron-depleted conditions. Indeed, the mutated oligonucleotides ICE₆₆₈₀* and ICE₇₈₉₅* showed only little competition even when added at a 300-fold molar excess, whereas competition with the wild-type oligonucleotides ICE₆₆₈₀ and ICE₇₈₉₅ became detectable already at a 3-fold molar excess (compare lanes 7 to 10 and 1 to 4 in Fig. 4A). This suggested a specific binding of Irr to ICE₆₆₈₀ and ICE₇₈₉₅.

It was obvious to test whether iron or heme has an effect on Irr binding to its target DNA sequences. Addition of iron (FeSO₄), heme, or hemin (all at 0.3 to 30 μM) to the in vitro binding reactions had no effect on specific binding of Irr to ICE₆₆₈₀ or ICE₇₈₉₅ (data not shown).

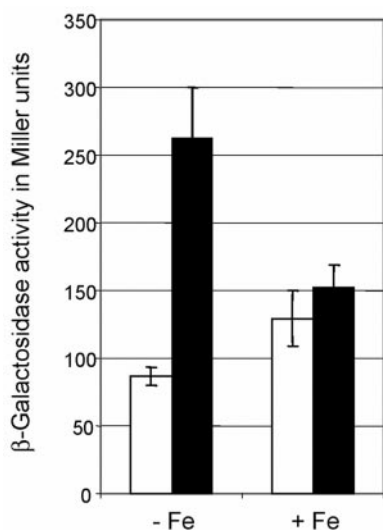


FIG. 5. β -Galactosidase activity derived from expression of plasmid-borne *bll6680'*-*lacZ* fusions with wild-type ICE₆₆₈₀ (pRJ8769, open columns) or mutant 1/2-ICE₆₆₈₀ (pRJ8770, filled columns) in *B. japonicum* 110*spc4* cells. Cultures were inoculated to an optical density (at 600 nm) of 0.02 and grown aerobically for about 36 h at 30°C in PSY medium lacking (–Fe) or supplemented (+Fe) with iron (50 μ M FeSO₄) and assayed in duplicate. Shown are mean values \pm standard errors of β -galactosidase activities (Miller units) derived from two independent experiments, each comprising two cultures per strain and condition.

ICE₆₆₈₀ mediates negative control of *bll6680*, and its function depends on both half-sites. One of the two newly identified Irr binders, ICE₆₆₈₀, was chosen for further analysis of its role in iron regulation of the associated *bll6680* gene. To do so, a variant of ICE₆₆₈₀, termed 1/2-ICE₆₆₈₀, was designed in which eight nucleotides at the 3' end of ICE₆₆₈₀ were simultaneously exchanged (Table 2). Based on our transcript mapping results (Fig. 2 and 3), we anticipated that this mutation would not interfere with the function of the –10 box of the *bll6680* promoter.

Plasmids pRJ8769 and pRJ8770, harboring a *bll6680'*-*lacZ* translational fusion with full-length ICE₆₆₈₀ or 1/2-ICE₆₆₈₀, respectively, were transferred to the *B. japonicum* 110*spc4* wild-type strain, and the resulting reporter strains were grown at low-iron (no iron supplement) or high-iron (50 μ M FeSO₄) conditions. Expression of *bll6680'*-*lacZ* with full-length ICE₆₆₈₀ (pRJ8769) was reproducibly about 1.5-fold higher under iron-replete conditions than under iron limitation (Fig. 5). Under the latter conditions, the strain with plasmid pRJ8770 showed more than threefold higher β -galactosidase activity than that containing pRJ8769. As expected, both fusions yielded comparable β -galactosidase activities under iron-replete conditions; however, for reasons that are not obvious, they did not reach the same expression level as did the strain with plasmid pRJ8770 grown under iron limitation. Iron regulation of *bll6680* was further substantiated by measuring expression from a chromosomally integrated *bll6680'*-*lacZ* reporter fusion. β -Galactosidase activity was consistently 1.7-fold higher in cells grown under iron-replete conditions than in iron-depleted cells, and the same result was obtained with the fusion integrated in the chromosome of the *fur* mutant strain GEM4 (data not shown). Taken together, these results indi-

cated that *bll6680* is under negative control and that intact ICE₆₆₈₀, but not Fur, is crucial for repression of the promoter under iron-limiting conditions.

Finally, Irr binding to 1/2-ICE₆₆₈₀ was also tested in vitro in the mobility shift assay. In contrast to ICE₆₆₈₀, no interaction was observed between 1/2-ICE₆₆₈₀ and the Irr protein (0.26 to 2.1 μ M) (Fig. 4B). Thus, both half-sites of ICE₆₆₈₀ are critical for productive Irr binding.

Identification of iron-regulated, ICE-associated genes by microarray analysis. Microarray analysis was used to identify iron-regulated *B. japonicum* genes on a genome-wide scale. All genes whose transcription changed reproducibly at least two-fold ($P \leq 0.01$) in the presence of iron (50 μ M FeSO₄) than in iron-depleted conditions are listed in Table S2 in the supplemental material. For each condition, four biological replicates were analyzed. We identified 17 genes to be induced and 68 genes to be repressed under iron-replete conditions. Remarkably, almost 40% of the genes are predicted to code for iron-containing proteins or for proteins involved in iron or energy metabolism.

The overlap of iron-regulated genes and genes associated with putative ICE motifs consisted of 6 genes whose expression was increased and 11 genes whose expression was decreased in the presence of iron (Table 5). Among the genes whose expression was increased were those encoding fumarate hydratase and aconitase, which both contain an iron-sulfur cluster. While the change value of *blr7895* (2.7-fold) was above the cutoff, *bll6680* was excluded from the selection because its change value was too low (see Discussion). Members of the *hmuVUT-hmuR-exbBD-tonB* gene cluster were found in the group of genes whose expression was decreased under high-iron conditions. The *hmuR* and *hmuT* genes which share the ICE_{*hmu*} motif showed very high change values of 51-fold and 18-fold, respectively.

DISCUSSION

The work reported here has expanded our knowledge of the *B. japonicum* regulatory circuitry that responds to external iron. The new and important facets are concerned with the functional analysis of the *cis*-acting ICE motif, the identification of the Irr protein as the putative *trans*-acting factor binding to ICEs, and the identification of additional iron-regulated target genes, many of them associated with an ICE-like motif.

In a previous study, the ICE motif was shown to be involved in transcriptional activation of *B. japonicum* heme uptake genes when iron was limited (43) (see the introduction). It was suggested that the ICE represents the binding site for a transcription activator protein, yet the element was only poorly characterized and the identity of the postulated binding protein remained unknown. 5'-end deletion analysis indicated that both half-sites of ICE_{*hmu*} are required and sufficient for maximal iron regulation of *hmuR*. As shown for *bll6680* and *blr7895*, ICE motifs can be involved also in repression under low-iron conditions, and at least in the case of ICE₆₆₈₀, both half-sites are critical for this type of negative control.

The *B. japonicum* ferric uptake regulator Fur (encoded by *bll0797*) is not directly involved in ICE-dependent regulation of *hmuR* and *bll6680*, as concluded from the retained regulation in the *fur* mutant strain GEM4. Likewise, lack of Fur did

TABLE 5. Fe-regulated *B. japonicum* genes or open reading frames associated with an ICE-like motif^a

Gene no.	Gene name	Description	Fold change	Putative ICE-like motif ^b	Fe-regulated genes possibly organized in an operon with the gene listed in column 1 ^c
Increased expression under iron repletion					
bll5796		Fumarate hydratase	4.5	TTTGAAAATCATGATAAA ₋₂₆₀	
bll0466	<i>acnA</i>	Aconitase	3.6	GGTTAGAAAAGCTTCTATA ₋₁₁₇	
blr2219		Dehydrogenase	3.5	CTTTTGACGAATTCCTAAA ₋₁₅₂	blr2217 ^d
blr6742		Putative glutamate synthase, small subunit	3.4	CTTCGGAAGAGTTCTAAT ₋₁₀₇	blr6743
blr0488	<i>leuC</i>	3-Isopropylmalate dehydratase, large subunit	3.3	TCTTGGAATGTTTCCTAGA ₋₁₉₆	
blr7895		Rubrerythrin-like protein	2.7	ATTTAGAATCATTCTAAA ₋₂₀	
Decreased expression under iron repletion					
bll7076	<i>hmuR</i>	Hemin receptor precursor	-50.8	ATTTACAATCGATATAAAA ₋₁₄₉	<i>tonB</i> , <i>exbD</i> , <i>exbB</i> , bll7074, bll7075
blr3555	<i>fcuA</i>	Probable ferrichrome receptor precursor	-49.8	CTTTGGAAGTTTTCCAAA ₋₂₃₂	bsr3556
bll7968		Probable TonB-dependent receptor	-35.6	GCTTAGAACCAATTCCTAAA ₋₉₈	bll7967, bll7966 ^e
blr4504		TonB-dependent receptor	-27.0	GTCTAGAAGGATTCCTAAA ₋₁₅₁	blr4505
blr7077	<i>hmuT</i>	Hemin ABC transporter, hemin-binding protein	-18.1	GTTTATATCGATTGTAAA ₋₂₈₄	<i>hmuU</i> , <i>hmuV</i>
blr3904		Probable iron transport protein	-16.1	TTTTAGAACGACTCCAAT ₋₁₆₆	blr3905 blr3906 blr3907 blr3908 blr5541
blr5540		Hypothetical protein	-12.7	GATCACAATCGGTCTAAA ₋₆₁	
bll0597		Similar to nickel-dependent hydrogenase, cytochrome B subunit	-6.5	CGTTAGAATATCTCTAAA ₋₄₆₀	
blr0697		Hypothetical protein	-3.7	TTTCAGAAGGCTTGTGAA ₋₃₈	
blr7418		Hypothetical glutathione S-transferase like protein	-2.9	TTTTCGAGTTTTTCTATT ₋₁₀₀	
bll2216		Transcriptional regulatory protein TetR family	-2.5	ATTTGGAATTCGTCTAAA ₋₅₅	

^a Fe-regulated genes were identified by microarray analysis using the BJAPETHa520090 gene chip and cDNA derived from *B. japonicum* cells grown under Fe-depleted (no Fe supplement) or Fe-replete (50 μM FeSO₄) conditions. Change values (*n*-fold) are based on four biological replicas for each growth condition. A comprehensive list of all Fe-regulated genes is shown in Table S2 in the supplemental material.

^b ICE-like motifs were identified as described in the text. A complete list of all ICE-like motifs found in this search is presented in Table S1 in the supplemental material. Numbers refer to the distance of the 3' nucleotide from the translational start codon of the ICE-associated gene.

^c Genes considered to be part of an operon are oriented in the same direction and separated by less than 32 nucleotides or by less than 100 nucleotides in case they are functionally related.

^d The ICE motif is associated with blr2217, which is probably organized in an operon with blr2219, yet no significant iron regulation was found for blr2217.

^e Expression of bll7967 and bll7966 was iron regulated; however, change (-fold) and *P* values were below the standard cutoff values.

not eliminate repression of bll6680 under low-iron conditions (data not shown). Somewhat unexpectedly, iron regulation of *hmuR* was not completely abolished in both backgrounds when ICE_{*hmu*} was partially (pRJ8723) or completely (pRJ8724) deleted. This may point to the existence of an additional ICE- and Fur-independent iron control mechanism.

The potential involvement of the *B. japonicum* Irr protein in ICE-dependent iron control could not be tested directly by genetic means, as we failed to delete the respective gene (bll0768) in our *B. japonicum* wild-type strain 110*spc4*. While the reason for this remains unknown, it contrasts with what was previously described for a different *B. japonicum* strain (LO) from which a mutant derivative carrying a Tn5 insertion in the 5' portion of the *irr* gene had been isolated (23). Therefore, Irr binding to ICE_{*hmu*}, ICE₆₆₈₀, and ICE₇₈₉₅ was probed in a yeast one-hybrid system and with electrophoretic mobility shift assays. While Irr bound to all three targets in vivo, specific in vitro binding of Irr was observed only for ICE₆₆₈₀ and ICE₇₈₉₅. This may reflect different binding affinities between negatively and positively regulated Irr target promoters. No indication for an

interaction between Fur and the ICE was found in the yeast reporter system, which is in accordance with the *lacZ* expression studies in the *B. japonicum* *fur* mutant. Collectively, the results from the ICE deletion analysis and the interaction studies with Irr provide compelling evidence that the Irr protein is the regulator that mediates ICE-dependent, positive or negative iron control. This conclusion is well compatible with the elevated activation of *hmuR* in the *fur* mutant background (Table 3), as the *irr* gene is under negative control by Fur (14, 25).

The Irr protein of *B. japonicum* was originally characterized as a repressor of the heme biosynthesis gene *hemB* in iron-limited cells (23). An increase of iron supply leads to *hemB* derepression. In addition to transcriptional control by Fur, the level of Irr protein is controlled at the posttranslational level via iron-dependent degradation mediated by heme (49). Specifically, it was shown that in the presence of iron, Irr interacts directly with the heme biosynthesis enzyme ferrochelatase, which leads to inactivation and subsequent heme-dependent degradation of Irr (50, 65). Since heme acts as an effector molecule in Irr degradation, *B.*

japonicum possesses means to coordinate iron availability with heme biosynthesis. In light of the dual control (transcriptional and posttranslational) which is imposed on Irr, there is no need for additional regulation of its DNA binding properties by iron and/or heme. Indeed, heme or iron had no effect on in vitro binding of Irr to ICE₆₆₈₀ and ICE₇₈₉₅.

Given that *B. japonicum* is able to utilize external heme as an iron source, it makes sense that cells induce heme uptake via Irr when Fe becomes limiting. As opposed to repression of *hemB*, *bll6680*, and *blr7895*, the *hmu* heme uptake genes appear to be activated by Irr under low-iron conditions. The same type of Irr-dependent regulation was observed previously for uptake of ferric iron, yet the respective genes had not been characterized in that study (23). Based on our microarray results, one may envisage the *fcuA* ferrichrome receptor gene (*blr3555*) and/or the putative iron uptake system encoded by the *blr3904* to *-3908* gene cluster as candidate genes for this phenotypic property, because they are repressed by iron and associated with ICE-like motifs (Table 5). Target-dependent positive or negative control by iron regulatory proteins is not unprecedented, as it had been described previously for the *N. meningitidis* Fur and the *Mycobacterium tuberculosis* IdeR regulators (9, 18). Notably, the latter is an essential protein in *M. tuberculosis* (52), which may also hold true for Irr in *B. japonicum*.

The identification of new ICEs and ICE-associated genes is another advancement reported here. Given the lack of any ICE consensus sequences at the onset of this work, we looked for *B. japonicum* genome sequences that shared closest similarity with the ICE near *hmuR*. Two of the four initially identified ICE-like sequences were of immediate interest because they are associated with genes that encode putative proteins involved in iron metabolism (*bll6680* for bacterioferritin and *blr7895* for rubrerythrin). Having determined the transcription start sites of *bll6680* and *blr7895*, it became evident that ICEs and promoters overlapped, a situation that is reminiscent of negatively regulated genes (Fig. 2). A repression mechanism at these genes is indeed likely, because we could demonstrate in vivo and in vitro binding of Irr to ICE₆₆₈₀ and ICE₇₈₉₅ (Table 4; Fig. 4).

The predicted functions of bacterioferritin and rubrerythrin are in accordance with the need to negatively regulate the corresponding genes in response to iron limitation. Bacterioferritin is a well-known iron-mineralizing and storage protein (7, 8) that should be synthesized only when the iron supply tends to exceed the cellular demand for iron incorporation into other proteins. Rubrerythrin, which ligates three iron atoms at two different sites, is also a structurally and biophysically well-characterized protein (33, 35); its function in cells, however, is much less clear. The current hypothesis is that rubrerythrin provides oxidative-stress protection via catalytic reduction of intracellular hydrogen peroxide (37, 56, 61), although this function was recently disputed (30). It is likely that such a function becomes more relevant at increasing iron concentrations with the concomitant risk that Fe²⁺ or Fe³⁺ reacts with H₂O₂ to form hydrogen peroxide radicals (Fenton reaction) (58). Conversely, it would make physiological sense if cells down-regulated rubrerythrin gene expression under iron limitation, which in fact holds true for *blr7895* (Fig. 3; Table S2 in the supplemental material).

A more global search for ICE-like motifs in the *B. japonicum* genome yielded a large number (172) of putative ICE motifs whose significance is difficult to evaluate. Yet, about 15% of them are linked to genes which either encode iron-containing proteins or whose function is related to iron metabolism. To identify those ICE-associated genes which are iron regulated, microarray analysis was performed using a recently developed *B. japonicum* whole-genome Affymetrix gene chip (F. Hauser et al., unpublished material). Strikingly, 41% (7 of 17) of the genes showing increased expression under high-iron conditions and 40% (27 of 68) of the genes repressed under these conditions are either directly associated with a predicted ICE-like motif or may belong to a putative operon that is preceded by such a motif. This suggests that ICE-dependent regulation through the Irr protein is a widely used mechanism for cellular iron homeostasis in *B. japonicum* and possibly also in other α -proteobacteria encoding an Irr homolog (62). It also may indicate that direct repression by the Fur protein is less common in *B. japonicum* than, e.g., in *E. coli* or *Bacillus subtilis* (4, 39). The sole *B. japonicum* target gene for which this regulatory function of Fur has been well documented is the *irr* gene (13), and only indirect evidence suggests Fur-dependent repression also at the *hemA* (25) and the *fegA* genes (34). This situation is reminiscent to that in *R. leguminosarum* and *Sinorhizobium meliloti*, where RirA, and not Fur, is the general iron regulator (59, 62).

Unexpectedly, the microarray analysis failed to detect iron regulation of *bll6680*, although this was clearly observed in the primer extension experiment (Fig. 3A) and, albeit weakly, also with the *bll6680'*-*lacZ* fusion (Fig. 5). It turns out that *bll6680* indeed showed increased expression under high-iron conditions in all four replicates of the microarray experiments, yet the average change value was below our cutoff (≥ 2 -fold change). This means that the number of genes listed in Table S2 is probably an underestimation which does not include all iron-regulated genes.

The ICE consensus sequence (Fig. 6) which was defined on the basis of all putative ICE motifs associated with iron-regulated genes (Table 5) reveals that critical nucleotides of the ICE comprise not only the TTTA-N₉-TAAA motif which was used in the initial search but rather a more extended palindrome (TTTAGAA-N₃-TTCTAAA) with individual positions showing different degrees of invariance. It is interesting to note that ICE₆₆₈₀ and ICE₇₈₉₅, which both bound to Irr in vitro, perfectly match the consensus sequence, whereas ICE_{*hmu*}, which showed no consistent in vitro binding of Irr (data not shown), deviates from the consensus sequence at three positions (TTTACAA-N₃-ATATAAA; italicized nucleotides mark deviations).

It is somewhat puzzling that we found no convincing ICE-like DNA sequence in the promoter region of the *irr*-controlled *hemB* gene. To our knowledge, Irr binding to an operator near *hemB* has not been shown directly, which leaves open the formal possibility that an Irr-dependent unknown regulator controls *hemB*. As the nucleotides of the ICE that make contact with Irr have not been ascertained precisely, the ICE consensus sequence proposed in Fig. 6 must be regarded as tentative.

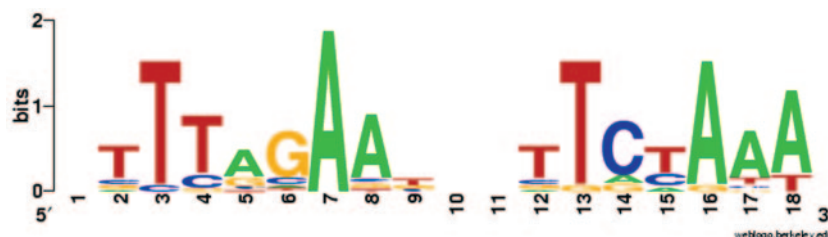


FIG. 6. ICE sequence logo created by using “WebLogo” version 2.8. The consensus motif is based on all individual ICE motifs listed in Table 4, plus that of bll6680 (ICE₆₆₈₀).

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REFERENCES

1. Alvarez-Morales, A., M. Betancourt-Alvarez, K. Kaluza, and H. Hennecke. 1986. Activation of the *Bradyrhizobium japonicum* *nifH* and *nifDK* operons is dependent on promoter-upstream DNA sequences. *Nucleic Acids Res.* **14**: 4207–4227.
2. Andrews, S. C., A. K. Robinson, and F. Rodriguez-Quinones. 2003. Bacterial iron homeostasis. *FEMS Microbiol. Rev.* **27**:215–237.
3. Babst, M., H. Hennecke, and H. M. Fischer. 1996. Two different mechanisms are involved in the heat-shock regulation of chaperonin gene expression in *Bradyrhizobium japonicum*. *Mol. Microbiol.* **19**:827–839.
4. Baichoo, N., T. Wang, R. Ye, and J. D. Helmann. 2002. Global analysis of the *Bacillus subtilis* Fur regulon and the iron starvation stimulon. *Mol. Microbiol.* **45**:1613–1629.
5. Benos, P. V., M. L. Bulyk, and G. D. Stormo. 2002. Additivity in protein-DNA interactions: how good an approximation is it? *Nucleic Acids Res.* **30**:4442–4451.
6. Boyd, J., M. N. Oza, and J. R. Murphy. 1990. Molecular cloning and DNA sequence analysis of a diphtheria toxin iron-dependent regulatory element (*dtxR*) from *Corynebacterium diphtheriae*. *Proc. Natl. Acad. Sci. USA* **87**: 5968–5972.
7. Braun, V. 1997. Avoidance of iron toxicity through regulation of bacterial iron transport. *Biol. Chem.* **378**:779–786.
8. Carrondo, M. A. 2003. Ferritins, iron uptake and storage from the bacterioferritin viewpoint. *EMBO J.* **22**:1959–1968.
9. Delany, I., R. Rappuoli, and V. Scarlato. 2004. Fur functions as an activator and as a repressor of putative virulence genes in *Neisseria meningitidis*. *Mol. Microbiol.* **52**:1081–1090.
10. Diaz-Mireles, E., M. Wexler, G. Sawers, D. Bellini, J. D. Todd, and A. W. Johnston. 2004. The Fur-like protein Mur of *Rhizobium leguminosarum* is a Mn²⁺-responsive transcriptional regulator. *Microbiology* **150**:1447–1456.
11. Ferguson, A. D., and J. Deisenhofer. 2004. Metal import through microbial membranes. *Cell* **116**:15–24.
12. Fischer, H. M., M. Babst, T. Kaspar, G. Acuña, F. Arigoni, and H. Hennecke. 1993. One member of a *groESL*-like chaperonin multigene family in *Bradyrhizobium japonicum* is co-regulated with symbiotic nitrogen fixation genes. *EMBO J.* **12**:2901–2912.
13. Friedman, Y. E., and M. R. O’Brien. 2004. The ferric uptake regulator (Fur) protein from *Bradyrhizobium japonicum* is an iron-responsive transcriptional repressor *in vitro*. *J. Biol. Chem.* **279**:32100–32105.
14. Friedman, Y. E., and M. R. O’Brien. 2003. A novel DNA-binding site for the ferric uptake regulator (Fur) protein from *Bradyrhizobium japonicum*. *J. Biol. Chem.* **278**:38395–38401.
15. Gaballa, A., T. Wang, R. W. Ye, and J. D. Helmann. 2002. Functional analysis of the *Bacillus subtilis* Zur regulon. *J. Bacteriol.* **184**:6508–6514.
16. Genco, C. A., and D. W. Dixon. 2001. Emerging strategies in microbial haem capture. *Mol. Microbiol.* **39**:1–11.
17. Gietz, D., A. St. Jean, R. A. Woods, and R. H. Schiestl. 1992. Improved method for high efficiency transformation of intact yeast cells. *Nucleic Acids Res.* **20**:1425.
18. Gold, B., G. M. Rodriguez, S. A. Marras, M. Pentecost, and I. Smith. 2001. The *Mycobacterium tuberculosis* IdeR is a dual functional regulator that controls transcription of genes involved in iron acquisition, iron storage and survival in macrophages. *Mol. Microbiol.* **42**:851–865.
19. Gonnet, G. H., M. T. Hallett, C. Korostensky, and L. Bernardin. 2000. Darwin v. 2.0: an interpreted computer language for the biosciences. *Bioinformatics* **16**:101–103.
20. Guerinot, M. L. 1991. Iron uptake and metabolism in the rhizobia legume symbioses. *Plant Soil* **130**:199–209.
21. Guerinot, M. L. 1994. Microbial iron transport. *Annu. Rev. Microbiol.* **48**: 743–772.
22. Hahn, M., and H. Hennecke. 1986. Genomstruktur von *Bradyrhizobium japonicum*: gemeinsames Vorkommen von repetitiven Sequenzen und Genen für die Wurzelknöllchensymbiose. Dissertation ETHZ Nr. 8104. Eidgenössische Technische Hochschule, Zürich, Switzerland.
23. Hamza, I., S. Chauhan, R. Hassett, and M. R. O’Brien. 1998. The bacterial Irr protein is required for coordination of heme biosynthesis with iron availability. *J. Biol. Chem.* **273**:21669–21674.
24. Hamza, I., R. Hassett, and M. R. O’Brien. 1999. Identification of a functional *fur* gene in *Bradyrhizobium japonicum*. *J. Bacteriol.* **181**:5843–5846.
25. Hamza, I., Z. Qi, N. D. King, and M. R. O’Brien. 2000. Fur-independent regulation of iron metabolism by Irr in *Bradyrhizobium japonicum*. *Microbiology* **146**:669–676.
26. Hantke, K. 2001. Iron and metal regulation in bacteria. *Curr. Opin. Microbiol.* **4**:172–177.
27. Hantke, K. 1981. Regulation of ferric iron transport in *Escherichia coli* K12: isolation of a constitutive mutant. *Mol. Gen. Genet.* **182**:288–292.
28. Hauser, F., A. Lindemann, S. Vuilleumier, A. Patrignani, R. Schlapbach, H. M. Fischer, and H. Hennecke. 17 November 2005. Design and validation of a partial-genome microarray for transcriptional profiling of the *Bradyrhizobium japonicum* symbiotic gene region. *Mol. Genet. Genomics* doi: 10.1007/s00438-005-0059-7.
29. Higuchi, R., B. Krummel, and R. K. Saiki. 1988. A general method of *in vitro* preparation and specific mutagenesis of DNA fragments: study of protein and DNA interactions. *Nucleic Acids Res.* **16**:7351–7367.
30. Jean, D., V. Briolat, and G. Reysset. 2004. Oxidative stress response in *Clostridium perfringens*. *Microbiology* **150**:1649–1659.
31. Kaneko, T., Y. Nakamura, S. Sato, K. Minamisawa, T. Uchiumi, S. Sasamoto, A. Watanabe, K. Idesawa, M. Iriguchi, K. Kawashima, M. Kohara, M. Matsumoto, S. Shimpo, H. Tsuruoka, T. Wada, M. Yamada, and S. Tabata. 2002. Complete genomic sequence of nitrogen-fixing symbiotic bacterium *Bradyrhizobium japonicum* USDA110. *DNA Res.* **9**:189–197.
32. Lawrence, C. E., S. F. Altschul, M. S. Boguski, J. S. Liu, A. F. Neuwald, and J. C. Wootton. 1993. Detecting subtle sequence signals: a Gibbs sampling strategy for multiple alignment. *Science* **262**:208–214.
33. LeGall, J., B. C. Prickril, I. Moura, A. V. Xavier, J. J. Moura, and B. H. Huynh. 1988. Isolation and characterization of rubrerythrin, a non-heme iron protein from *Desulfovibrio vulgaris* that contains rubredoxin centers and a hemerythrin-like binuclear iron cluster. *Biochemistry* **27**:1636–1642.
34. LeVier, K., and M. L. Guerinot. 1996. The *Bradyrhizobium japonicum* *vegA* gene encodes an iron-regulated outer membrane protein with similarity to hydroxamate-type siderophore receptors. *J. Bacteriol.* **178**:7265–7275.
35. Li, M., M. Y. Liu, J. LeGall, L. L. Gui, J. Liao, T. Jiang, J. P. Zhang, D. C. Liang, and W. R. Chang. 2003. Crystal structure studies on rubrerythrin: enzymatic activity in relation to the zinc movement. *J. Biol. Inorg. Chem.* **8**:149–155.
36. Liu, J., T. E. Wilson, J. Milbrandt, and M. Johnston. 1993. Identifying DNA-binding sites and analyzing DNA-binding domains using a yeast selection system. *Methods* **5**:125–137.
37. Lumpio, H. L., N. V. Shenvi, A. O. Summers, G. Voordouw, and D. M. Kurtz, Jr. 2001. Rubrerythrin and rubredoxin oxidoreductase in *Desulfovibrio vulgaris*: a novel oxidative stress protection system. *J. Bacteriol.* **183**:101–108.
38. Masse, E., and S. Gottesman. 2002. A small RNA regulates the expression of genes involved in iron metabolism in *Escherichia coli*. *Proc. Natl. Acad. Sci. USA* **99**:4620–4625.
39. McHugh, J. P., F. Rodriguez-Quinones, H. Abdul-Tehrani, D. A. Svistunenko,

- R. K. Poole, C. E. Cooper, and S. C. Andrews. 2003. Global iron-dependent gene regulation in *Escherichia coli*. A new mechanism for iron homeostasis. *J. Biol. Chem.* **278**:29478–29486.
40. Miller, J. H. 1972. Experiments in molecular genetics. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.
41. Minton, N. P. 1984. Improved plasmid vectors for the isolation of translational *lac* gene fusions. *Gene* **31**:269–273.
42. Nadler, K. D., A. W. B. Johnston, J. W. Chen, and T. R. John. 1990. A *Rhizobium leguminosarum* mutant defective in symbiotic iron acquisition. *J. Bacteriol.* **172**:670–677.
43. Nienaber, A., H. Hennecke, and H. M. Fischer. 2001. Discovery of a haem uptake system in the soil bacterium *Bradyrhizobium japonicum*. *Mol. Microbiol.* **41**:787–800.
44. Nienaber, A., A. Huber, M. Göttfert, H. Hennecke, and H. M. Fischer. 2000. Three new NifA-regulated genes in the *Bradyrhizobium japonicum* symbiotic gene region discovered by competitive DNA-RNA hybridization. *J. Bacteriol.* **182**:1472–1480.
45. Norrander, J., T. Kempe, and J. Messing. 1983. Construction of improved M13 vectors using oligodeoxynucleotide-directed mutagenesis. *Gene* **26**:101–106.
46. Noya, F., A. Arias, and E. Fabiano. 1997. Heme compounds as iron sources for nonpathogenic *Rhizobium* bacteria. *J. Bacteriol.* **179**:3076–3078.
47. Ochsner, U. A., and M. L. Vasil. 1996. Gene repression by the ferric uptake regulator in *Pseudomonas aeruginosa*: cycle selection of iron-regulated genes. *Proc. Natl. Acad. Sci. USA* **93**:4409–4414.
48. Panina, E. M., A. A. Mironov, and M. S. Gelfand. 2001. Comparative analysis of FUR regulons in gamma-proteobacteria. *Nucleic Acids Res.* **29**:5195–5206.
49. Qi, Z., I. Hamza, and M. R. O'Brian. 1999. Heme is an effector molecule for iron-dependent degradation of the bacterial iron response regulator (Irr) protein. *Proc. Natl. Acad. Sci. USA* **96**:13056–13061.
50. Qi, Z., and M. R. O'Brian. 2002. Interaction between the bacterial iron response regulator and ferroxidase mediates genetic control of heme biosynthesis. *Mol. Cell* **9**:155–162.
51. Regensburger, B., and H. Hennecke. 1983. RNA polymerase from *Rhizobium japonicum*. *Arch. Microbiol.* **135**:103–109.
52. Rodriguez, G. M., M. I. Voskuil, B. Gold, G. K. Schoolnik, and I. Smith. 2002. *ideR*, an essential gene in *Mycobacterium tuberculosis*: role of IdeR in iron-dependent gene expression, iron metabolism, and oxidative stress response. *Infect. Immun.* **70**:3371–3381.
53. Sambrook, J., E. F. Fritsch, and T. Maniatis. 1989. Molecular cloning: a laboratory manual, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.
54. Simon, R., U. Priefer, and A. Pühler. 1983. Vector plasmids for in vivo and in vitro manipulation of Gram-negative bacteria, p. 98–106. *In* A. Pühler (ed.), Molecular genetics of the bacteria-plant interaction. Springer-Verlag, Heidelberg, Germany.
55. Studier, F. W., and B. A. Moffatt. 1986. Use of bacteriophage T7 RNA polymerase to direct selective high-level expression of cloned genes. *J. Mol. Biol.* **189**:113–130.
56. Sztukowska, M., M. Bugno, J. Potempa, J. Travis, and D. M. Kurtz, Jr. 2002. Role of rubrerythrin in the oxidative stress response of *Porphyromonas gingivalis*. *Mol. Microbiol.* **44**:479–488.
57. Todd, J. D., M. Wexler, G. Sawers, K. H. Yeoman, P. S. Poole, and A. W. B. Johnston. 2002. RirA, an iron-responsive regulator in the symbiotic bacterium *Rhizobium leguminosarum*. *Microbiology* **148**:4059–4071.
58. Touati, D. 2000. Iron and oxidative stress in bacteria. *Arch. Biochem. Biophys.* **373**:1–6.
59. Viguier, C., P. Ó. Cuív, P. Clarke, and M. O'Connell. 2005. RirA is the iron response regulator of the rhizobactin 1021 biosynthesis and transport genes in *Sinorhizobium meliloti* 2011. *FEMS Microbiol. Lett.* **246**:235–242.
60. Wandersman, C., and P. Delepelaire. 2004. Bacterial iron sources: from siderophores to hemophores. *Annu. Rev. Microbiol.* **58**:611–647.
61. Weinberg, M. V., F. E. Jenney, Jr., X. Cui, and M. W. Adams. 2004. Rubrerythrin from the hyperthermophilic archaeon *Pyrococcus furiosus* is a rubredoxin-dependent, iron-containing peroxidase. *J. Bacteriol.* **186**:7888–7895.
62. Wexler, M., J. D. Todd, O. Kolade, D. Bellini, A. M. Hemmings, G. Sawers, and A. W. B. Johnston. 2003. Fur is not the global regulator of iron uptake genes in *Rhizobium leguminosarum*. *Microbiology* **149**:1357–1365.
63. Wexler, M., K. H. Yeoman, J. B. Stevens, N. G. de Luca, G. Sawers, and A. W. B. Johnston. 2001. The *Rhizobium leguminosarum tonB* gene is required for the uptake of siderophore and haem as sources of iron. *Mol. Microbiol.* **41**:801–816.
64. Winteler, H. V., and D. Haas. 1996. The homologous regulators ANR of *Pseudomonas aeruginosa* and FNR of *Escherichia coli* have overlapping but distinct specificities for anaerobically inducible promoters. *Microbiology* **142**:685–693.
65. Yang, J., K. Ishimori, and M. R. O'Brian. 2005. Two heme binding sites are involved in the regulated degradation of the bacterial iron response regulator (Irr) protein. *J. Biol. Chem.* **280**:7671–7676.



ERRATUM

The Iron Control Element, Acting in Positive and Negative Control of Iron-Regulated *Bradyrhizobium japonicum* Genes, Is a Target for the Irr Protein

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Volume 188, no. 2, p. 733–744, 2006. Page 743, Fig. 6 legend: “. . . ICE motifs listed in Table 4 . . .” should read “. . . ICE motifs listed in Table 5. . . .”